

SYSTEMATIC REVIEW

Prevalence of KRAS, PIK3CA, BRAF and AXIN2 gene mutations in colorectal cancer and their relationship with dental agenesis: a systematic review

Prevalencia de mutaciones en los genes KRAS, PIK3CA, BRAF y AXIN2 en cáncer colorrectal y su relación con agenesia dental: revisión sistemática

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Open access

Received: 07/05/2021

Accepted: 25/11/2021

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Keywords: Anodontia; Mutación; Prevalence; Genes; Colorectal Cancer; Colorectal Neoplasms (MeSH).

Palabras clave: Anodoncia; Mutación; Prevalencia; Genes; Cáncer colorrectal; Neoplasma colorrectal (DeCS).

How to cite: Sir-Mendoza F, Madera M, González-Martínez F. Prevalence of KRAS, PIK3CA, BRAF and AXIN2 gene mutations in colorectal cancer and their relationship with dental agenesis: a systematic review. Rev. Fac. Med. 2023;71(1):e95595. English. doi: <https://doi.org/10.15446/revfacmed.v71n1.95595>.

Cómo citar: Sir-Mendoza F, Madera M, González-Martínez F. [Prevalencia de mutaciones en los genes KRAS, PIK3CA, BRAF y AXIN2 en cáncer colorrectal y su relación con agenesia dental: revisión sistemática]. Rev. Fac. Med. 2023;71(1):e95595. English. doi: <https://doi.org/10.15446/revfacmed.v71n1.95595>.

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Abstract

Introduction: The study of allelic and genotypic frequencies contributes to determining the distribution of genetic variants in different populations and their possible association with biomarkers. This knowledge could improve the decision-making process regarding the management of some diseases such as colorectal cancer (CRC), in which the detection of clinical biomarkers such as dental agenesis could be crucial in clinical practice.

Objective: To evaluate the available scientific evidence on the prevalence of mutations in KRAS, PIK3CA, BRAF and AXIN2 genes and their possible association with tooth agenesis in people with CRC.

Materials and methods: A systematic search was conducted in PubMed, EMBASE and Cochrane Library databases using the following search strategy: type of studies: observational studies reporting the prevalence of KRAS, PIK3CA, BRAF and AXIN2 mutations in people diagnosed with CRC and their possible association with dental agenesis; publication language: English and Spanish; publication period: 2010-2020; search terms: “Genes”, “RAS”, “Kras”, “PIK3CA”, “BRAF”, “AXIN2”, “Mutation”, “Polymorphism”, “Colorectal Neoplasms” and “Colorectal Cancer”, used in different combinations (“AND” and “OR”).

Results: The initial search yielded 403 records, but only 30 studies met the eligibility criteria. Of these, 11, 5, 5 and 1 only reported the prevalence of PIK3CA, KRAS, BRAF and AXIN2 mutations, respectively; while 8 reported the prevalence of more than one of these mutations in patients with CRC. The prevalence of KRAS (p.Gly12Asp), PIK3CA (p.Glu545Lys), and BRAF (p.Val600Glu) mutations ranged from 20.5% to 54%, 3.5% to 20.2%, and 2.5% to 12.1%, respectively. There were no findings regarding the association between the occurrence of these mutations and dental agenesis.

Conclusions: KRAS mutations were the most prevalent; however, there is no evidence on the association between dental agenesis and the occurrence of KRAS, PIK3CA and BRAF germline mutations in individuals with CRC.

Resumen

Introducción. El estudio de frecuencias alélicas y genotípicas contribuye a determinar la distribución de variantes genéticas en diferentes poblaciones y su posible asociación con biomarcadores. Este conocimiento podría mejorar la toma de decisiones respecto al manejo de algunas enfermedades como el cáncer colorrectal (CCR), en el cual la detección de biomarcadores clínicos como la agenesia dental podría ser crucial en la práctica clínica.

Objetivo. Evaluar la evidencia científica sobre la prevalencia de mutaciones en los genes KRAS, PIK3CA, BRAF y AXIN2 y su posible asociación con la agenesia dental en individuos con CCR.

Materiales y métodos. Se realizó una búsqueda sistemática en PubMed, Embase y Cochrane Library empleando la siguiente estrategia de búsqueda: tipos de estudio: estudios observacionales que reportaran la prevalencia de mutaciones en los genes KRAS, PIK3CA, BRAF y AXIN2 en personas con CCR y su posible asociación con agenesia dental; idioma: inglés y español; periodo de publicación: 2010-2020; términos de búsqueda: “Genes”, “RAS”, “Kras”, “PIK3CA”, “BRAF”, “AXIN2”, “Mutation”, “Polymorphism”, “Colorectal Neoplasms” y “Colorectal Cancer” en diferentes combinaciones (“AND” y “OR”).

Resultados. Se identificaron 403 registros, pero solo 30 cumplieron con los criterios de elegibilidad. De estos, 11, 5, 5 y 1 solo reportaron la prevalencia de mutaciones en PIK3CA, KRAS, BRAF y AXIN2, respectivamente, mientras que 8 reportaron la prevalencia de más de una de estas mutaciones en pacientes con CCR. La prevalencia de mutaciones en los genes KRAS (p.Gly12Asp), PIK3CA (p.Glu545Lys), y BRAF (p.Val600Glu) varió entre 20.5% y 54%, 3.5% y 20.2%, y 2.5% y 12.1%, respectivamente. No hubo hallazgos respecto a la asociación entre la ocurrencia de estas mutaciones y la agenesia dental.

Conclusiones. Las mutaciones en el gen KRAS fueron las más prevalentes; sin embargo, no hay evidencia de la asociación entre agenesia dental y la ocurrencia de mutaciones en los genes KRAS, PIK3CA y BRAF en individuos con CCR.

Introduction

Colorectal cancer (CRC) is one of the leading causes of morbimortality worldwide.¹ According to demographic and time projections, its global incidence is expected to increase by 60% by 2030, resulting in more than 2.2 million new cases and 1.1 million deaths per year.² The pathogenesis of this type of cancer is complex and is not yet fully understood. However, genetic factors reportedly play a critical role in tumorigenesis.³ In this regard, Kolligs⁴ reported that up to one third of the risk of developing CRC can be attributed to hereditary factors. Likewise, people with a family history of CRC are at a higher risk of developing it. Overall, genetic mutations are critical in the development of CRC and several genes and signaling pathways have been associated with its occurrence, including *KRAS*, *BRAF*, *PIK3CA*, *RAS-RAF-MAPK* and *PI3K-PTEN-AKT*.⁵⁻⁷

The main therapeutic approach in CRC patients includes surgery and subsequent chemotherapy, and it has been described that over 75% of cases that receive this treatment can be effectively cured.⁸ However, it has also been reported that more than 30% of these patients could develop new neoplastic polyps,⁸ suggesting that this treatment is not completely effective.⁹ Thus, new therapies have been developed, such as anti-epidermal growth factor receptor (anti-EGFR) monoclonal antibodies, considering that EGFR is the major therapeutic target in CRC.¹⁰

However, it has been described that the efficacy of anti-EGFR therapy is affected when there are mutations in *KRAS*, *BRAF* and *PIK3CA* genes.^{11,12} Therefore, these genes constitute important biomarkers in CRC, both in terms of diagnosis and prognosis. In addition, it has been reported that mutations in *AXIN2* gene associated with tooth agenesis could act as a diagnostic biomarker for CRC, so the presence of variants in this gene and of this congenital developmental anomaly of the oral cavity have been proposed as predictive factors of this type of cancer.¹³

Non-syndromic tooth agenesis, the most common human malformation,¹⁴ is the congenital absence of one or more permanent teeth due to alterations occurring during early stages of dental development.¹⁵ In addition, an association between mutations in *AXIN2* gene and teeth development anomalies has been reported in mice, suggesting their possible participation in human dental development.¹⁶ In this sense, Lammi *et al.*¹⁶ found that a nonsense mutation (p.Arg656Stop) in this gene was associated with the occurrence of tooth agenesis (oligodontia) and predisposition to CRC. Likewise, Rosales-Reynoso *et al.*¹⁷ reported that patients with the homozygous T/T genotype of the single nucleotide polymorphism rs2240308 in the *AXIN2* gene have a higher risk of CRC.

Taking this information into account, adequate knowledge about the prevalence of these genetic mutations and their possible association with clinical biomarkers such as tooth agenesis by health professionals is of great importance to improve the average time to diagnosis of CRC and, therefore, the prognosis of these patients, particularly dentists, who have an important role in detecting tooth agenesis. Thus, the aim of this study was to evaluate the available scientific evidence on the prevalence of mutations in genes *KRAS*, *PIK3CA*, *BRAF*, *AXIN2* mutations and their possible association with dental agenesis in people with CRC.

Materials and methods

We conducted a systematic review following the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement.¹⁸

Search strategy

A structured and systematic search using MeSH and DeCS terms was performed in Medline (Via PubMed), EMBASE (Via Ovid) and Cochrane Library databases using the following search strategy: type of studies: observational studies reporting the prevalence of *KRAS*, *PIK3CA*, *BRAF* and *AXIN2* mutations in people diagnosed with CRC and their possible association with dental agenesis; publication period: January 2010-September 2020; publication languages: English and Spanish; search terms: “Genes”, “RAS”, “Kras”, “PIK3CA”, “BRAF”, “AXIN2”, “Prevalence”, “Mutation”, “Polymorphism”, “Colorectal Neoplasms”, “Colorectal Cancer” and “dental agenesis”, used in different combinations (“AND” and “OR”). The search equation used in each database is shown in Appendix 1.

Screening and selection process

The titles and abstracts of the records retrieved in the searches were managed using the reference manager software EndNote® (Version X8, Thomson Reuters). After removing duplicates, two reviewers (FS and FG) independently screened all titles/abstracts to exclude those studies that were not relevant for the objective of this systematic review. Then, the full texts of the screened articles were read by the two reviewers to confirm if they addressed the topics of interest for this review and make a decision on their final inclusion for full analysis taking into account the following inclusion criterion: being case-control, cohort or cross-sectional studies addressing the prevalence of *KRAS*, *PIK3CA*, *BRAF* and *AXIN2* mutations/polymorphisms in people with primary (adenocarcinoma) or metastatic CRC and their possible association with tooth agenesis. In addition, studies conducted in animals, those published before 2010, and those addressing other types of genetic alterations and reporting other associations with CRC or in concomitance with other cancers were excluded.

Disagreements were resolved by consensus, and when necessary, a third reviewer (MM) participated in the discussion until an agreement was reached.

Methodological quality assessment

The methodological quality of the selected studies was evaluated independently by two appraisers following criteria previously reported.¹⁹ The following criteria were evaluated: a) research question/aim of the research (1 item); b) participants (5 items); c) comparability between groups studied (4 items); d) definition and measurement of the main variables (4 items); e) statistical analysis and confounders (4 items), global assessment of internal validity; f) results (4 items); g) conclusions, external validity and applicability of results (4 items), and h) conflicts of interest (1 item). Each item was assessed using the following options: “very good”, “good”, “regular”, “bad”, “not reported” and “does not apply”. Regarding the global assessment of the quality of the studies, “high”, “medium” or “low” categories were used.

Two authors (FS and FG) established a grading system by assigning “5”, “4”, “3”, “2”, “1” and “0” points to the “very good”, “good”, “regular”, “bad”, “not reported” and “does not apply” assessment criteria, with 135 and 27 being the maximum and minimum scores, respectively. Studies with a score between 81 and 107 and with regular internal validity were classified as having “medium” methodological quality, while those with a score >108, as having a “high” methodological quality.

Finally, it should be noted that in those studies in which the assessment of the “comparability between groups studied” criteria was not possible due to the study design, the maximum and minimum scores were 115 and 23, respectively. In these cases, studies with regular internal validity and with a score between 69 and 91 and those with a score >92 were considered to have “medium” and “high” methodological quality, respectively.

Data extraction and analysis

The following information was extracted for each study: first author, publication year, geographic region in which the study was conducted, sample size, general prevalence of the mutation, mutation prevalence by sex (male and female), mutation (changes in amino acids) and sequencing techniques. The information was included in tables and organized by genes.

Results

Selection and characterization of studies

The study selection process is presented in Figure 1. In total, 30 articles were included for full analysis.

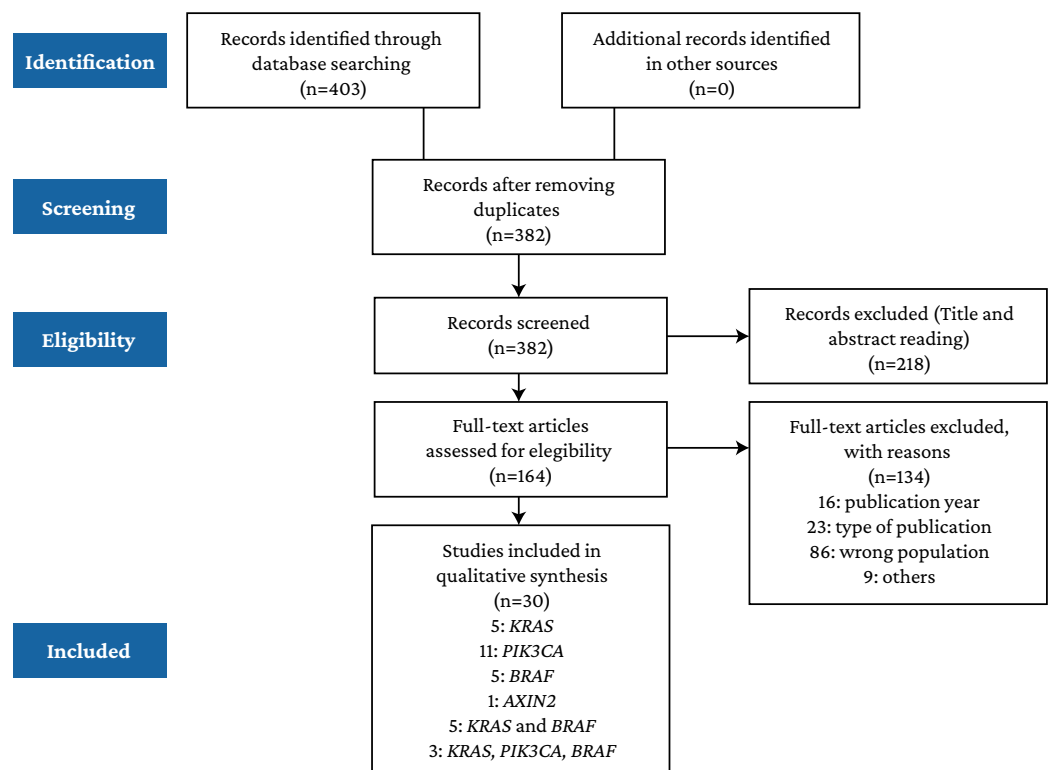


Figure 1. Study selection process.

Source: own elaboration.

Regarding their geographical distribution, 10 studies were conducted in Asia; 8, in America; 5, in the Middle East; 3, in Europe; 2, in Oceania, and 1, in Africa. Regarding sample size, studies addressing KRAS, PIK3CA and BRAF mutations were conducted in samples ranging from 49 to 5 732, 61 to 2 299, and 17 to 1 110 individuals, respectively. Besides, all studies were cross-sectional (Table 1).

Table 1. General characteristics of the studies included for full analysis.

Authors (year)	Country	Study design	Sample	Genes assessed	MQA
Gavin <i>et al.</i> ²⁰ (2012)	United States	CSS	2 299	<i>PIK3CA</i>	High
Palomba <i>et al.</i> ²¹ (2012)	Italy	CSS	384	<i>PIK3CA</i>	High
Mao <i>et al.</i> ²² (2012)	China	CSS	61	<i>PIK3CA</i>	High
Liao <i>et al.</i> ²³ (2012)	United States	CSS	1 170	<i>PIK3CA</i>	High
Watanabe <i>et al.</i> ²⁴ (2013)	Japan	CSS	5 732	<i>KRAS</i>	High
Shen <i>et al.</i> ²⁵ (2013)	China	CSS	674	<i>KRAS, BRAF</i>	High
Patil <i>et al.</i> ²⁶ (2013)	India	CSS	1 323	<i>KRAS</i>	High
Chang <i>et al.</i> ²⁷ (2013)	Taiwan	CSS	165	<i>KRAS, BRAF</i>	Medium
Rosty <i>et al.</i> ²⁸ (2013)	Australia	CSS	757	<i>PIK3CA</i>	High
Kang <i>et al.</i> ²⁹ (2013)	United States	CSS	150	<i>PIK3CA</i>	Medium
Marchoudi <i>et al.</i> ³⁰ (2013)	Morocco	CSS	92	<i>BRAF</i>	High
Samadder <i>et al.</i> ³¹ (2013)	United States	CSS	563	<i>BRAF</i>	Medium
Baskin <i>et al.</i> ³² (2014)	Turkey	CSS	49	<i>KRAS</i>	Medium
Imamura <i>et al.</i> ³³ (2014)	United States	CSS	1 267	<i>KRAS</i>	High
Bader & Ismail ³⁴ (2014)	Saudi Arabia	CSS	83	<i>KRAS</i>	High
Chen <i>et al.</i> ³⁵ (2014)	China	CSS	214	<i>KRAS, PIK3CA, BRAF</i>	High
Bisht <i>et al.</i> ³⁶ (2014)	India	CSS	204	<i>PIK3CA</i>	High
Russo <i>et al.</i> ³⁷ (2014)	United States	CSS	222	<i>PIK3CA</i>	High
Siraj <i>et al.</i> ³⁸ (2014)	Saudi Arabia	CSS	757	<i>BRAF</i>	High
Ye <i>et al.</i> ³⁹ (2015)	China	CSS	535	<i>KRAS, BRAF</i>	Medium
Zhang <i>et al.</i> ⁷ (2015)	China	CSS	1 110	<i>KRAS, PIK3CA, BRAF</i>	High
Phipps <i>et al.</i> ⁴⁰ (2015)	United States, Canada, Australia	CSS	377	<i>PIK3CA</i>	High
Foltran <i>et al.</i> ⁴¹ (2015)	Italy	CSS	194	<i>PIK3CA</i>	High
Allard <i>et al.</i> ⁴² (2015)	France	CSS	1 428	<i>BRAF</i>	High
Vatandoust <i>et al.</i> ⁴³ (2016)	Australia	CSS	3 318	<i>KRAS, BRAF</i>	Medium
Watson <i>et al.</i> ⁴⁴ (2016)	United States	CSS	447	<i>KRAS, BRAF</i>	High
Al-Shamsi <i>et al.</i> ⁴⁵ (2016)	Arab countries	CSS	99	<i>KRAS, PIK3CA, BRAF</i>	High
Molaei <i>et al.</i> ⁴⁶ (2016)	Iran	CSS	85	<i>BRAF</i>	Medium
Jauhri <i>et al.</i> ⁴⁷ (2017)	India	CSS	112	<i>PIK3CA</i>	High
Chang <i>et al.</i> ⁴⁸ (2020)	Taiwan	CSS	161	<i>AXIN2</i>	High

MQA: Methodological quality assessment; CSS: Cross-sectional study.

Source: Own elaboration.

Methodological quality assessment

Based on the scores obtained using the critical appraisal of epidemiological cross-sectional studies instrument, it was determined that 23 studies had “high” methodological quality, while the remaining 7 studies had “medium” quality (Table 1). The domains with the highest mean scores were “participants” and “results”, whereas the lowest mean scores were observed in the “statistical analysis and confounders” domain.

Prevalence of mutations

KRAS

The highest overall prevalence of mutations in this gene was 54% in a sample of 447 individuals and the lowest, 20.5% in a sample of 1 323 people. The highest prevalence by sex was 65.7% and 50% in males and females, respectively. Moreover, in 46.15% and 23.08% of the studies, direct sequencing and next-generation sequencing (NGS) were carried out to identify mutations. The most frequent mutation consisted of an amino acid change from glycine into aspartic acid in codon 12 (Table 2).

PIK3CA

The highest general prevalence of mutations in *PIK3CA* was 20.2% in a sample of 2 299 people and the lowest, 3.5% in a sample of 1 110 individuals. In addition, the highest prevalence of mutations in men was 65.4%, and in women, 50%. Mutations were identified by means of direct sequencing and NGS in 57.14% and 21.43% of the studies, respectively. The most frequent mutation was the substitution of glutamic acid by lysine in codon 545 (p.Glu545Lys) (Table 2).

BRAF

The highest overall prevalence of mutations was 27% in a sample of 563 individuals and the lowest, 2.5% in a sample of 757 individuals. Besides, it is worth noting that BRAF mutations were not identified in two studies. The highest prevalence in men was 55.6%, and in women, 44.4%. Mutations were detected using direct sequencing and NGS in 69.23% and 23.08% of the studies, respectively. The most frequent variant was an amino-acid change of valine by glutamate in codon 600 (p.Val600EGLu) (Table 2).

AXIN2

Only one study (conducted in Taiwan) addressed AXIN2 mutations.⁴⁸ The general prevalence of mutations was 21.7% in a sample of 161 individuals. Mutations were identified using NGS, being p.A603P the most frequent mutation (Table 2).

Table 2. Prevalence of mutations by gene and sequencing technique reported by the studies included in the review.

Gene	Author (year)	Prevalence of mutations (%)			Most frequent mutation (%)	Sequencing technique
		General (%)	Sex (%)			
			Male	Female		
KRAS	Watanabe <i>et al.</i> ²⁴ (2013)	37.6	21.4	16.1	NR	DS, LA
	Shen <i>et al.</i> ²⁵ (2013)	35.9	19.0	16.5	G12D (13.6)	DS
	Patil <i>et al.</i> ²⁶ (2013)	20.5	13.8	6.6	G12A (36.5)	DS
	Chang <i>et al.</i> ²⁷ (2013)	35.7	NR	NR	G12D (35.5)	PEA
	Baskin <i>et al.</i> ³² (2014)	32.1	20.2	14.2	G12D (12.2)	ARMS
	Imamura <i>et al.</i> ³³ (2014)	39.8	19.8	20.0	G12D (12)	PS
	Bader & Ismail ³⁴ (2014)	42.2	27	14	G12D (45.7)	AA
	Chen <i>et al.</i> ³⁵ (2014)	44.9	25.4	19.8	G12D (35.4)	DS
	Ye <i>et al.</i> ³⁹ (2015)	37.9	20.6	17.3	G12D (18.4)	DS, ARMS
	Zhang <i>et al.</i> ⁷ (2015)	45.4	25.7	19.6	G12D (40.7)	DS, ARMS, NGS
	Vatandoust <i>et al.</i> ⁴³ (2016)	38.9	NR	NR	NR	NR
	Watson <i>et al.</i> ⁴⁴ (2016)	54	NR	NR	NR	PS, NGS, SNUPE
	Al-Shamsi <i>et al.</i> ⁴⁵ (2016)	44.4	25.2	19.1	NR	NGS
PIK3CA	Gavin <i>et al.</i> ²⁰ (2012)	20.1	10.4	9.7	NR	AA
	Palomba <i>et al.</i> ²¹ (2012)	17.4	9.8	7.5	E545A (14)	DS
	Mao <i>et al.</i> ²² (2012)	8.2	4.9	3.2	H1047L (7)	DS
	Liao <i>et al.</i> ²³ (2012)	16	8.2	7.8	NR	PS
	Rosty <i>et al.</i> ²⁸ (2013)	14	6.8	7.2NR	E542K (35)	DS
	Kang <i>et al.</i> ²⁹ (2013)	12	NR	NR	NR	DS
	Chen <i>et al.</i> ³⁵ (2014)	12.3	8.0	4.2.	H1047R (31)	DS
	Bisht <i>et al.</i> ³⁶ (2014)	5.9	2.9	2.9	E545K (3.4)	DS
	Russo <i>et al.</i> ³⁷ (2014)	13	NR	NR	NR	DS
	Zhang <i>et al.</i> ⁷ (2015)	3.5	1.9	1.5	H1047R (3.5)	DS, ARMS, NGS
	Phipps <i>et al.</i> ⁴⁰ (2015)	11	4.5	6.6	E542K,E545K (64)	PS
	Foltran <i>et al.</i> ⁴¹ (2015)	16.49	NR	NR	E545K (56)	PS
	Al-Shamsi <i>et al.</i> ⁴⁵ (2016)	13.1	5.1	8.08	NR	NGS
	Jauhri <i>et al.</i> ⁴⁷ (2017)	16.1	13.3	2.6	E545A, E545K, H1047R (15.8)	NGS
BRAF	Chang <i>et al.</i> ²⁷ (2013)	4.24	NR	NR	V600E (100)	HRM
	Shen <i>et al.</i> ²⁵ (2013)	6.96	4.1	2.8	V600E (1.8)	DS
	Marchoudi <i>et al.</i> ³⁰ (2013)	5.4	NR	NR	V600E (100)	DS
	Samadder <i>et al.</i> ³¹ (2013)	27	NR	NR	V600E (100)	DS
	Chen <i>et al.</i> ³⁵ (2014)	4.2	2.3	1.8	V600E (89)	DS
	Siraj <i>et al.</i> ³⁸ (2014)	2.5	1.4	1.0	V600E (89.5)	DS
	Allard <i>et al.</i> ⁴² (2015)	6.4	NR	NR	V600E (100)	HRM, DS
	Zhang <i>et al.</i> ⁷ (2015)	3.1	1.6	1.4	V600E (100)	DS, ARMS, NGS
	Ye <i>et al.</i> ³⁹ (2015)	4.4	1.5	2.8	V600E (80)	DS, ARMS
	Vatandoust <i>et al.</i> ⁴³ (2016)	12.1	NR	NR	NR	NR
	Al-Shamsi <i>et al.</i> ⁴⁵ (2016)	4.0	2.0	2.0	NR	NGS
	Molaei <i>et al.</i> ⁴⁶ (2016)	0	0	0	NR	DS
	Watson <i>et al.</i> ⁴⁴ (2016)	0	0	0	NR	PS, NGS
AXIN2	Chang <i>et al.</i> ⁴⁸ (2020)	21.7	NR	NR	A603P (11.4)	NGS

DS: Direct sequencing; LA: Luminex assay; PEA: Primer extension assay; ARMS: Amplification refractory mutations system-PCR, PS: Pyrosequencing; AA: Array analysis; NGS: Next-generation sequencing; NR: Not reported; SNUPE: Single-nucleotide primer extension; HRM: High resolution melting. Source: Own elaboration.

Discussion

The fact that cancer cells have multiple genetic mutations suggests that the development and progression of tumors could be partially caused by mutagenesis. Additionally, these mutations can contribute to developing resistance to conventional oncological therapies, such as chemotherapy.⁴⁹ Currently, scientific evidence shows that, despite the development of new drugs, therapy against cancer is limited, since new ways of resistance have emerged, such as drug inactivation, alteration of drug targets, drug efflux, and cell death inhibition.⁵⁰

Taking the above into account, understanding the distribution of mutations in oncogenes in cancer patients is essential to improve both the knowledge of the genomic profile of malignant diseases and personalized medicine, since a better understanding of the cancer genome is important to choose the best treatment for each oncological patient according to their individual characteristics.⁵¹

Several studies have reported the presence of mutations in multiple genes involved in the development and progression of CRC, being *KRAS*, *PIK3CA* and *BRAF* the ones with the highest prevalence of mutations.^{6,7} In addition, *AXIN2* has been identified as a potentially useful gene in the early diagnosis of CRC through clinical markers such as tooth agenesis.¹⁶

Regarding *KRAS* gene, it has been described that mutations in this gene are the mutations most frequently identified in the development of human tumors (approximately 30%).^{52,53} *KRAS* encodes for a protein constituted by 188 residues of amino acids involved in molecular pathways activation that allows signal transduction from the cell surface to the nucleus.⁵⁴ *KRAS* is found in chromosome 12 and is a member of the RAS gene family; mutations in this gene comprise 86% of all RAS family mutations, being most frequently observed in codons 12 and 13 of exon 2,⁵⁵ and less frequently, in codons 61⁵⁶ and 146.^{57,58} In addition, the main mutation in *KRAS* consists of a G>A transition followed by a G>T transversion in exon 1.⁵⁹

KRAS has been studied as a predictive molecular marker of anti-EGFR monoclonal antibody resistance in primary and metastatic CRC patients.^{11,60,61} In the presence of *KRAS* mutations, GTPase activity decreases and the *KRAS* mutant protein remains bound to GTP in its active conformation, transmitting signals continuously. As a result, signal transmission is not blocked by EGFR inhibitors and the effects of this therapy are scarce or cannot be observed.⁶²⁻⁶⁴

In the present review, the prevalence of *KRAS* mutations ranged between 20.5% to 54.^{7,24-27,32-35,39,43-45} Regarding the prevalence of these mutations in different regions of the world, the following was found:

- i) In America, the highest prevalence was 54% in a sample of 447 individuals,⁴⁴ while the lowest was 40% in a sample of 1 267 people.³³ In relation to sex distribution, the highest prevalence of mutations in males was 19.8, while in females it was 20%.³³
- ii) In Asia, the highest prevalence of *KRAS* mutations was found in China (45.4% in a total sample of 1 110 individuals),⁷ while the lowest was found in India (20.5% in a sample of 1 323 people).²⁶ Regarding sex distribution, the highest prevalence of mutations in males was 25.7%,⁷ while in females it was 19.8%.³⁵
- iii) In the Middle East, the highest prevalence of *KRAS* mutations was 44.4% in a sample of 99 individuals from several countries,⁴⁵ and the lowest prevalence was 30.6% in a sample of 49 people from Turkey.³² In addition, the highest prevalence of mutations in males was 27%,³⁴ while in females it was 19.1%.⁴⁵
- iv) Only one study included in this review provided data on *KRAS* mutations in Oceania, reporting a prevalence of 38.9% in a sample of 3 318 Australian subjects; however, no data on prevalence by sex were reported.⁴³

Finally, the most frequent mutation in the *KRAS* gene was the substitution of glycine for aspartate (p.Gly12Asp). However, the substitution of glycine by alanine (p.Gly12Ala) was the most common mutation in one study.²⁶ These conformational biochemical changes have been associated with a poor prognosis in terms of survival and with increased tumoral aggressiveness.^{32,65,66}

PIK3CA is a gene located in chromosome 3 that codes for PI3K protein. PI3K is part of the lipid kinase family, which is involved in cell proliferation, growth and survival.^{67,68} PI3K protein is also involved in the PI3K/AKT pathway, which catalyzes AKT phosphorylation, activating the downstream signaling pathway.⁶⁹

It has been reported that the presence of mutations in this gene stimulates said pathway and promotes cell growth in various types of cancers.⁷⁰ The prevalence of mutations in the *PIK3CA* gene ranges from 7% to 32% in CRC patients, being the G>A transversion in exons 9 and 20 the most common mutation (80% of mutations). Furthermore, there is contradictory evidence in relation to the presence of these mutations as a predictor of response to cancer treatment.^{28,71,72} On the one hand, mutations in exon 20 have been associated with a low response to treatment with cetuximab and chemotherapy.⁷³ On the other, clinical trials such as the one conducted by Soeda *et al.*⁷⁴ state that their presence might not contribute to the prediction of the response to monoclonal therapy with cetuximab in patients with advanced and/or metastatic CRC.

According to the data retrieved in the present systematic review, the prevalence of mutations in *PIK3CA* ranged between 3.5% to 20.1%.^{7,20-23,28,29,35,36,37,40,41,45,47} Regarding the prevalence of these mutations in different regions of the world, the following was found:

- i) In America, the highest prevalence was 20.2%, in a sample of 2 299 individuals from the United States,²⁰ while the lowest prevalence was 11%, in a sample of 377 people from the United States and Canada.⁴⁰ In addition, the highest prevalence of mutations in males was 10.4% and in females, 9.7%.²⁰
- ii) In Asia, the highest prevalence of *PIK3CA* mutations was reported in India (16.1% in a sample of 112 patients),⁴⁷ while the lowest was found in China (3.5% in a sample of 1 110 people).⁷ In addition, the highest prevalence of mutations in males was 13.3%⁴⁷, and in females, 4.2%.³⁵
- iii) Only one study reported data for people from the Middle East, finding a prevalence of *PIK3CA* mutations of 13.1% (5.1. in males and 8% in females) in 99 individuals from Middle Eastern countries.⁴⁵
- iv) Similarly, only one study reported data for population from Western Europe, finding a prevalence of *PIK3CA* mutations of 17.4% (9.8% in males and 7.5% in females) in a sample of 384 Italian individuals.²¹
- v) Only one study provided data on *PIK3CA* mutations in Oceania, reporting a prevalence of 14% (6.8% in males and 7.2 in females) in a sample of 757 individuals from Australia.²⁸

Finally, the most frequent variant in the *PIK3CA* gene was the replacement of glutamic acid by lysine (p.Glu545Lys) due to alterations in exon 9. However, a high mutation index was also reported in exon 20, resulting in the substitution of histidine by leucine (p.His1047Leu) and of histidine by arginine (p.His1047Arg).

BRAF is a gene involved in cell proliferation and differentiation, as well as in apoptosis pathways;⁷⁵ besides, it has been described that the presence of mutations in in this gene might lead to phenotypic alterations in the colorectal tissue.⁷⁵ In the present review, the prevalence of mutations in *BRAF* ranged from 2.5% to 27%.^{7,25,27,30,31,35,38,39,42-46} Regarding the prevalence of these mutations in different regions of the world, it was found that:

- i) In America, the highest prevalence was 27% in a sample of 563 individuals.³¹ In contrast, the lowest prevalence (0%) was reported by Watson *et al.*⁴⁴ in a study conducted in 17 people; both studies were carried out in the United States. No data on the prevalence of these mutations in men and women were reported.
- ii) In Asia, the highest prevalence was 6.96% in a sample of 674 people²⁵ and the lowest was 3.1% in a sample of 1 110 individuals,⁷ with both studies being conducted in China. With regard to the prevalence of *BRAF* mutations by sex, the highest prevalence in men was 4.1% and in women, 2.8%.^{25,39}
- iii) In the Middle East, the highest prevalence of mutations was 4% in a sample of 99 individuals from several countries,⁴⁵ while the lowest prevalence (0%) was reported by Molaei *et al.*⁴⁶ in a study conducted in 85 people from Iran. Furthermore, the highest prevalence of mutations was 2% in both males and females.⁴⁵
- iv) Only one study reported data on *BRAF* mutations in population from Western Europe, finding a prevalence of 6.4% in 1428 individuals from France.⁴² No data on the prevalence of these mutations by sex were reported.
- v) In Africa, a study conducted in 92 people from Morocco found a prevalence of 5.4%, however no data on the prevalence of these mutations stratified by sex was reported.³⁰
- vi) In the case of Oceania, a study conducted in 173 individuals from Australia reported a prevalence of mutations of 12.1%.⁴³

Finally, the most frequent mutation in this gene was the substitution of valine by glutamate in codon 600 (p.Val600Glu). On the other hand, it is worth noting that Samowitz *et al.*⁷⁶ reported that individuals with mutations in *BRAF* have more aggressive CRC phenotypes and show a poor response to treatment with cetuximab or panitumumab. Thus, and given that most of the studies included in this review only studied the p.Val600Glu genetic variant, further research on other possible hotspot regions in this gene associated with CRC is required.

Regarding studies conducted before 2012, the following data on the prevalence of mutations in the *KRAS*, *PIK3CA* and *BRAF* genes was found: Segura-Urbe *et al.*⁷⁷ and Vaughn *et al.*⁷⁸ reported a prevalence of *KRAS* mutations of 32.4% and 42.4% in 37 colorectal tumors in Mexican individuals and in 2 121 colorectal adenocarcinomas in people living in the United States, respectively. Herreros-Villanueva *et al.*⁷⁹ and Velho *et al.*⁸⁰ reported a prevalence of *PIK3CA* mutations of 8.22% and 7.1% in CRC specimens of 73 Spanish individuals and in 103 CRC carcinomas collected from Portuguese individuals, respectively. Finally, Nicolantonio *et al.*⁸¹, in a study conducted in 113 patients with metastatic CRC from Italy and Switzerland, reported a prevalence of *BRAF* mutations of 14%.

After comparing these data with those reported in the studies included in this systematic review, it is possible to say that the frequencies of mutations in *KRAS*, *PIK3CA* and *BRAF* genes in CRC patients have not changed much in recent years. However, systematic reviews that include a greater publication period are necessary to evaluate increasing or decreasing trends in the prevalence of mutations in these genes in CRC patients.

The differences between the general prevalence of mutations and their prevalence in men and women in the same geographical area could be attributed to both differences in the sample sizes of the studies and the sensitivity of the molecular techniques used, which have been shown to influence the frequency in which these mutations are detected.^{81,82} Other factors that might influence said frequency include the quality and quantity of DNA obtained, tumor heterogeneity and possible environmental exposures unknown to the researchers or that they cannot control.^{83,84}

Although in most of the studies included in our review there was not a significant association between the sex of the patient and the prevalence of mutations in *KRAS*, *PIK3CA*, *BRAF* and *AXIN2*, said association was evaluated since it has been described that associations between genetic mutations and the sex of the individual could provide information about the pathogenesis of different diseases,⁸⁵ and because of the differences, both environmental and genetic, that have been described between men and women around the world in terms of susceptibility to and incidence of cancer.⁸⁶⁻⁸⁸ Therefore, it can be assumed that the differences in the prevalence of mutations in these genes between males and females might condition a higher frequency of CRC in individuals of a specific sex.

The incidence and mortality rates of CRC have increased in recent years,⁸⁹ therefore, there is a higher need to identify and implement diagnostic strategies for the early detection of this cancer, including the analysis of molecular and clinical biomarkers. In this regard, it has been suggested that *AXIN2* gene could be considered a molecular biomarker of CRC, since the presence of mutations in this gene and of tooth agenesis have been proposed as predictive factors of CRC.^{13,16}

In this regard, the present systematic review aimed to report the prevalence of mutations in *AXIN2* in individuals with primary and/or metastatic CRC around the world. However, only one study was retrieved. In addition, due to the knowledge of the possible associations of *AXIN2* gene with both CRC phenotypes, we sought to identify if some of the most prevalent genetic mutations in CRC, such as the ones in *KRAS*, *PIK3CA* and *BRAF* genes, were also associated with tooth agenesis; nevertheless, no studies reporting such an association were identified.

AXIN2 is known for its tumor suppressing activity by negatively regulating the Wnt pathway through the intracellular degradation of β -catenin.^{90,91} In mice, *AXIN2* is expressed during odontogenesis in dental mesenchyme, enamel knot, dental papilla and mesenchymal odontoblast.¹⁶ It is reasonable to hypothesize that an impairment in this gene could affect the development of molars and incisors, leading to tooth agenesis.⁹² In addition, there is evidence that the expression of *AXIN2* in colorectal tissue can lead to carcinomas.¹⁶ Wu *et al.*⁹³ reported that mutations in *AXIN2* could influence the expression of the protein it codes for, which can play a critical role in carcinogenesis, a similar claim to that of Rosales-Reynoso *et al.*,¹⁷ who state that these mutations act as a genetic risk factor for the development of CRC. Similarly, Marvin *et al.*⁹⁴ reported that the presence of nonsense mutation p.Tyr663X (c.1989G>A) in *AXIN2* results in protein truncation in individuals with oligodontia and gastrointestinal neoplasms. Therefore, tooth agenesis and the presence of variants in *AXIN2* could be used as clinical and molecular markers for the development of CRC.

The present study highlights the importance of researching the distribution of mutations in *KRAS*, *PIK3CA*, *BRAF* and *AXIN2* genes in people with CRC from different regions of the world to determine the impact of these variants in the early diagnosis of this type of cancer, as well as in its prognosis in terms of survival and efficacy of therapies. Studying the presence of genetic mutations in heterogeneous populations is of great importance, given the possible association between a higher probability of mortality due to CRC and the ethnic and sexual differences in the presence of certain genetic mutations. In this regard, ethnicity has been shown to be associated with a higher risk of cancer^{93,95-97} and with a worse prognosis in the presence of *KRAS*,⁹⁸ *PIK3CA*²⁹ and *BRAF*⁹⁹ mutations.

Conducting additional studies on the association between genetic mutations, including those in *KRAS*, *PIK3CA*, *BRAF* and *AXIN2* genes, and the development of CRC in diverse populations is recommended in order to contribute to the knowledge on the genome of

this type of cancer genome. Likewise, conducting studies that analyze the association between the presence of mutations in *AXIN2* and dental agenesis as a clinical marker for the early diagnosis of CRC is also recommended.

Conclusions

Our findings suggest that worldwide there is a diverse distribution of *KRAS*, *PIK3CA* and *BRAF* mutations in individuals with CRC, being *KRAS* mutations the most prevalent. Moreover, according to the evidence here retrieved there is no association between tooth agenesis and *KRAS*, *PIK3CA* and *BRAF* germline gene mutations in these patients. *AXIN2* is the unique gene in which an association with both phenotypes (i.e., primary and metastatic) has been well established, but population studies on the prevalence of *AXIN2* mutations are limited.

Conflicts of interest

None stated by the authors.

Funding

None stated by the authors.

Acknowledgements

None stated by the authors.

References

1. Favoriti P, Carbone G, Greco M, Pirozzi F, Pirozzi RE, Corcione F. Worldwide burden of colorectal cancer: a review. *Updates Surg.* 2016;68(1):7-11. <https://doi.org/gfxqf9>.
2. Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. *Gut.* 2017;66(4):683-691. <https://doi.org/f9tq5n>.
3. Bartsch H, Dally H, Popanda O, Risch A, Schmezer P. Genetic risk profiles for cancer susceptibility and therapy response. *Recent Results Cancer Res.* 2007;174:19-36. <https://doi.org/fk598d>.
4. Kolligs FT. Diagnostics and Epidemiology of Colorectal Cancer. *Visc Med.* 2016;32(3):158-64. <https://doi.org/f9g5n6>.
5. Peyssonnaud C, Eychène A. The Raf/MEK/ERK pathway: new concepts of activation. *Biol Cell.* 2001;93(1-2):53-62. <https://doi.org/bfq2jp>.
6. Calistri D, Rengucci C, Seymour I, Lattuneddu A, Polifemo AM, Monti F, *et al.* Mutation analysis of p53, K-ras, and BRAF genes in colorectal cancer progression. *J Cell Physiol.* 2005;204(2):484-8. <https://doi.org/dmw4n9>.
7. Zhang J, Zheng J, Yang Y, Lu J, Gao J, Lu T, *et al.* Molecular spectrum of KRAS, NRAS, BRAF and PIK3CA mutations in Chinese colorectal cancer patients: analysis of 1,110 cases. *Sci Rep.* 2015;5:18678. <https://doi.org/f9dsbb>.
8. Roy S, Majumdar APN. Cancer Stem Cells in Colorectal Cancer: Genetic and Epigenetic Changes. *J Stem Cell Res Ther.* 2012;Suppl 7(6):10342. <https://doi.org/j7m3>.
9. Rentsch M, Schiergens T, Khandoga A, Werner J. Surgery for Colorectal Cancer - Trends, Developments, and Future Perspectives. *Visc Med.* 2016;32(3):184-91. <https://doi.org/f988w4>.
10. Ciardiello F, Tortora G. EGFR antagonists in cancer treatment. *N Engl J Med.* 2008;358(11):1160-74. <https://doi.org/cxrg3b>.
11. Di Nicolantonio F, Martini M, Molinari F, Sartore-Bianchi A, Arena S, Saletti P, *et al.* Wild-type BRAF is required for response to panitumumab or cetuximab in metastatic colorectal cancer. *J Clin Oncol.* 2008;26(35):5705-12. <https://doi.org/fhk5p>.

12. Abdul-Jalil KI, Sheehan KM, Toomey S, Schmid J, Prehn J, O'Grady A, *et al.* The frequencies and clinical implications of mutations in 33 kinase-related genes in locally advanced rectal cancer: a pilot study. *Ann Surg Oncol.* 2014;21(8):2642-9. <https://doi.org/f59qtn>.
13. Callahan N, Modesto A, Meira R, Seymen F, Patir A, Vieira AR. Axis Inhibition Protein 2 (AXIN2) Polymorphisms and Tooth Agnesis. *Arch Oral Biol.* 2009;54(1):45-9. <https://doi.org/dr8xp>.
14. Kantaputra PN, Kaewgahya M, Hatsadaloi A, Vogel P, Kawasaki K, Ohazama A, *et al.* GREMLIN 2 Mutations and Dental Anomalies. *J Dent Res.* 2015;94(12):1646-52. <https://doi.org/f72b8h>.
15. Williams MA, Letra A. The Changing Landscape in the Genetic Etiology of Human Tooth Agnesis. *Genes (Basel).* 2018;9(5):255. <https://doi.org/j7m8>.
16. Lammi L, Arte S, Somer M, Jarvinen H, Lahermo P, Thesleff I, *et al.* Mutations in AXIN2 cause familial tooth agnesis and predispose to colorectal cancer. *Am J Hum Genet.* 2004;74(5):1043-50. <https://doi.org/dm48>.
17. Rosales-Reynoso MA, Arredondo-Valdez AR, Wence-Chavez LI, Barros-Nunez P, Gallegos-Arreola MP, Flores-Martinez SE, *et al.* AXIN2 Polymorphisms and Their Association with Colorectal Cancer in Mexican Patients. *Genet Test Mol Biomarkers.* 2016;20(8):438-44. <https://doi.org/j7nb>.
18. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* 2009;6(7):e1000097. <https://doi.org/bq3jpc>.
19. Berra S, Elorza-Ricart JM, Estrada MD, Sánchez E. Instrumento para la lectura crítica y la evaluación de estudios epidemiológicos transversales. *Gac Sanit.* 2008;22(5):492-7.
20. Gavin PG, Colangelo LH, Fumagalli D, Tanaka N, Remillard MY, Yothers G, *et al.* Mutation profiling and microsatellite instability in stage II and III colon cancer: an assessment of their prognostic and oxaliplatin predictive value. *Clin Cancer Res.* 2012;18(23):6531-41. <https://doi.org/ggsmtb>.
21. Palomba G, Colombino M, Contu A, Massidda B, Baldino G, Pazzola A, *et al.* Prevalence of KRAS, BRAF, and PIK3CA somatic mutations in patients with colorectal carcinoma may vary in the same population: clues from Sardinia. *J Transl Med.* 2012;10:178. <https://doi.org/f97t57>.
22. Mao C, Zhou J, Yang Z, Huang Y, Wu X, Shen H, *et al.* KRAS, BRAF and PIK3CA mutations and the loss of PTEN expression in Chinese patients with colorectal cancer. *PLoS One.* 2012;7(5):e36653. <https://doi.org/j7nd>.
23. Liao X, Morikawa T, Lochhead P, Imamura Y, Kuchiba A, Yamauchi M, *et al.* Prognostic role of PIK3CA mutation in colorectal cancer: cohort study and literature review. *Clin Cancer Res.* 2012;18(8):2257-68. <https://doi.org/gk7r4j>.
24. Watanabe T, Yoshino T, Uetake H, Yamazaki K, Ishiguro M, Kurokawa T, *et al.* KRAS mutational status in Japanese patients with colorectal cancer: results from a nationwide, multicenter, cross-sectional study. *Jpn J Clin Oncol.* 2013;43(7):706-12. <https://doi.org/f43kj6>.
25. Shen Y, Wang J, Han X, Yang H, Wang S, Lin D, *et al.* Effectors of Epidermal Growth Factor Receptor Pathway: The Genetic Profiling of KRAS, BRAF, PIK3CA, NRAS Mutations in Colorectal Cancer Characteristics and Personalized Medicine. *PLoS One.* 2013;8(12):e81628. <https://doi.org/gfw2qn>.
26. Patil H, Korde R, Kapat A. KRAS gene mutations in correlation with clinicopathological features of colorectal carcinomas in Indian patient cohort. *Med Oncol.* 2013;30(3):617. <https://doi.org/f47n29>.
27. Chang YS, Chang SJ, Yeh KT, Lin TH, Chang JG. RAS, BRAF, and TP53 gene mutations in Taiwanese colorectal cancer patients. *Onkologie.* 2013;36(12):719-24. <https://doi.org/j7nf>.
28. Rosty C, Young JP, Walsh MD, Clendenning M, Sanderson K, Walters RJ, *et al.* PIK3CA activating mutation in colorectal carcinoma: associations with molecular features and survival. *PLoS One.* 2013;8(6):e65479. <https://doi.org/ghb4x8>.
29. Kang M, Shen XJ, Kim S, Araujo-Perez F, Galanko JA, Martin CF, *et al.* Somatic gene mutations in African Americans may predict worse outcomes in colorectal cancer. *Cancer Biomark.* 2013;13(5):359-66. <https://doi.org/j7nk>.
30. Marchoudi N, Amrani-Hassani-Joutei H, Jouali F, Fekkak J, Rhaissi H. Distribution of KRAS and BRAF mutations in Moroccan patients with advanced colorectal cancer. *Pathol Biol (Paris).* 2013;61(6):273-6. <https://doi.org/f5m6c9>.
31. Samadder NJ, Vierkant RA, Tillmans LS, Wang AH, Weisenberger DJ, Laird PW, *et al.* Associations between colorectal cancer molecular markers and pathways with clinicopathologic features in older women. *Gastroenterology.* 2013;145(2):348-56.e1-2. <https://doi.org/f2kqp2>.
32. Baskin Y, Dagdeviren YK, Calibasi G, Canda AE, Sarioglu S, Ellidokuz H, *et al.* KRAS mutation profile differences between rectosigmoid localized adenocarcinomas and colon adenocarcinomas. *J Gastrointest Oncol.* 2014;5(4):265-9. <https://doi.org/j7nm>.
33. Imamura Y, Lochhead P, Yamauchi M, Kuchiba A, Qian ZR, Liao X, *et al.* Analyses of clinicopathological, molecular, and prognostic associations of KRAS codon 61 and codon 146 mutations in colorectal cancer: cohort study and literature review. *Mol Cancer.* 2014;13:135. <https://doi.org/f57kgf>.
34. Bader T, Ismail A. Higher prevalence of KRAS mutations in colorectal cancer in Saudi Arabia: Propensity for lung metastasis. *Alex J Med.* 2014;50(3):203-9. <https://doi.org/j7qk>.
35. Chen J, Guo F, Shi X, Zhang L, Zhang A, Jin H, *et al.* BRAF V600E mutation and KRAS codon 13 mutations predict poor survival in Chinese colorectal cancer patients. *BMC Cancer.* 2014;14:802. <https://doi.org/f6p4jw>.

36. Bisht S, Ahmad F, Sawaimoon S, Bhatia S, Das BR. Molecular spectrum of KRAS, BRAF, and PIK3CA gene mutation: determination of frequency, distribution pattern in Indian colorectal carcinoma. *Med Oncol*. 2014;31(9):124. <https://doi.org/f6h8mh>.
37. Russo AL, Borger DR, Szymonifka J, Ryan DP, Wo JY, Blaszkowsky LS, *et al*. Mutational analysis and clinical correlation of metastatic colorectal cancer. *Cancer*. 2014;120(10):1482-90. <https://doi.org/j7qp>.
38. Siraj AK, Bu R, Prabhakaran S, Bavi P, Beg S, Al Hazmi M, *et al*. A very low incidence of BRAF mutations in Middle Eastern colorectal carcinoma. *Mol Cancer*. 2014;13:168. <https://doi.org/f59nrk>.
39. Ye JX, Liu Y, Qin Y, Zhong HH, Yi WN, Shi XY. KRAS and BRAF gene mutations and DNA mismatch repair status in Chinese colorectal carcinoma patients. *World J Gastroenterol*. 2015;21(5):1595-605. <https://doi.org/f62q96>.
40. Phipps AI, Ahnen DJ, Cheng I, Newcomb PA, Win AK, Burnett T. PIK3CA Somatic Mutation Status in Relation to Patient and Tumor Factors in Racial/Ethnic Minorities with Colorectal Cancer. *Cancer Epidemiol Biomarkers Prev*. 2015;24(7):1046-51. <https://doi.org/ghb4x2>.
41. Foltran L, De Maglio G, Pella N, Ermacora P, Aprile G, Masiero E, *et al*. Prognostic role of KRAS, NRAS, BRAF and PIK3CA mutations in advanced colorectal cancer. *Future Oncol*. 2015;11(4):629-40. <https://doi.org/f63jmd>.
42. Allard MA, Saffroy R, de la Maisonneuve PB, Ricca L, Bosselut N, Hamelin J, *et al*. Colorectal liver metastases are more often super wild type. Toward treatment based on metastatic site genotyping? *Target Oncol*. 2015;10(3):415-21. <https://doi.org/j7qq>.
43. Vatandoust S, Price TJ, Ullah S, Roy AC, Beeke C, Young JP, *et al*. Metastatic Colorectal Cancer in Young Adults: A Study From the South Australian Population-Based Registry. *Clin Colorectal Cancer*. 2016;15(1):32-6. <https://doi.org/f8b6pb>.
44. Watson R, Liu TC, Ruzinova MB. High frequency of KRAS mutation in early onset colorectal adenocarcinoma: implications for pathogenesis. *Hum Pathol*. 2016;56:163-70. <https://doi.org/gfxr5w>.
45. Al-Shamsi HO, Jones J, Fahmawi Y, Dahbour I, Tabash A, Abdel-Wahab R, *et al*. Molecular spectrum of KRAS, NRAS, BRAF, PIK3CA, TP53, and APC somatic gene mutations in Arab patients with colorectal cancer: determination of frequency and distribution pattern. *J Gastrointest Oncol*. 2016;7(6):882-902. <https://doi.org/gh92vq>.
46. Molaei M, Kishani-Farahani R, Maftouh M, Taleghani MY, Vahdatinia M, Khatami F, *et al*. Lack of BRAFV600E mutation in stage I and II of colorectal cancer. *Gastroenterol Hepatol Bed Bench*. 2016;9(2):94-9.
47. Jauhri M, Bhatnagar A, Gupta S, Bp M, Minhas S, Shokeen Y, *et al*. Prevalence and coexistence of KRAS, BRAF, PIK3CA, NRAS, TP53, and APC mutations in Indian colorectal cancer patients: Next-generation sequencing-based cohort study. *Tumour Biol*. 2017;39(2):1010428317692265. <https://doi.org/f9q7f4>.
48. Chang SC, Lan YT, Lin PC, Yang SH, Lin CH, Liang WY, *et al*. Patterns of germline and somatic mutations in 16 genes associated with mismatch repair function or containing tandem repeat sequences. *Cancer Med*. 2020;9(2):476-86. <https://doi.org/j7rc>.
49. Loeb KR, Loeb LA. Significance of multiple mutations in cancer. *Carcinogenesis*. 2000;21(3):379-85. <https://doi.org/d857gf>.
50. Housman G, Byler S, Heerboth S, Lapinska K, Longacre M, Snyder N, *et al*. Drug resistance in cancer: an overview. *Cancers (Basel)*. 2014;6(3):1769-92. <https://doi.org/gcx56j>.
51. Adjiri A. DNA Mutations May Not Be the Cause of Cancer. *Oncol Ther*. 2017;5(1):85-101. <https://doi.org/gg2jrw>.
52. Ihle NT, Byers LA, Kim ES, Saintigny P, Lee JJ, Blumenschein GR, *et al*. Effect of KRAS oncogene substitutions on protein behavior: implications for signaling and clinical outcome. *J Natl Cancer Inst*. 2012;104(3):228-39. <https://doi.org/fzh8zm>.
53. Arrington AK, Heinrich EL, Lee W, Duldulao M, Patel S, Sanchez J, *et al*. Prognostic and Predictive Roles of KRAS Mutation in Colorectal Cancer. *Int J Mol Sci*. 2012;13(10):12153-68. <https://doi.org/f95z3g>.
54. McGrath JP, Capon DJ, Smith DH, Chen EY, Seeburg PH, Goeddel DV, *et al*. Structure and organization of the human Ki-ras proto-oncogene and a related processed pseudogene. *Nature*. 1983;304(5926):501-6. <https://doi.org/dqxvhv>.
55. Rosty C, Young JP, Walsh MD, Clendenning M, Walters RJ, Pearson S, *et al*. Colorectal carcinomas with KRAS mutation are associated with distinctive morphological and molecular features. *Mod Pathol*. 2013;26(6):825-34. <https://doi.org/ghb4x9>.
56. Kimura K, Nagasaka T, Hoshizima N, Sasamoto H, Notohara K, Takeda M, *et al*. No duplicate KRAS mutation is identified on the same allele in gastric or colorectal cancer cells with multiple KRAS mutations. *J Int Med Res*. 2007;35(4):450-7. <https://doi.org/f3vthg>.
57. Takahashi N, Yamada Y, Taniguchi H, Akiyoshi K, Honma Y, Iwasa S, *et al*. Mutations in NRAS codon 61, KRAS codon 146, and BRAF V600E as prognostic factors in patients who received anti-EGFR antibody for metastatic colorectal cancer. *Journal of Clinical Oncology*. 2012;30(Suppl 15):e14126-e. <https://doi.org/j7rd>.
58. Edkins S, O'Meara S, Parker A, Stevens C, Reis M, Jones S, *et al*. Recurrent KRAS codon 146 mutations in human colorectal cancer. *Cancer Biol Ther*. 2006;5(8):928-32. <https://doi.org/dp5vkc>.
59. Poehlmann A, Kuester D, Meyer F, Lippert H, Roessner A, Schneider-Stock R. K-ras mutation detection in colorectal cancer using the Pyrosequencing technique. *Pathol Res Pract*. 2007;203(7):489-97. <https://doi.org/cgcmtm>.

60. Karapetis CS, Khambata-Ford S, Jonker DJ, O'Callaghan CJ, Tu D, Tebbutt NC, *et al.* K-ras mutations and benefit from cetuximab in advanced colorectal cancer. *N Engl J Med.* 2008;359(17):1757-65. <https://doi.org/bmmxxk>.
61. Amado RG, Wolf M, Peeters M, Van Cutsem E, Siena S, Freeman DJ, *et al.* Wild-type KRAS is required for panitumumab efficacy in patients with metastatic colorectal cancer. *J Clin Oncol.* 2008;26(10):1626-34. <https://doi.org/c6qdf3>.
62. Clevers H, Nusse R. Wnt/beta-catenin signaling and disease. *Cell.* 2012;149(6):1192-205. <https://doi.org/gf3r2k>.
63. Peeters M, Douillard JY, Van Cutsem E, Siena S, Zhang K, Williams R, *et al.* Mutant KRAS codon 12 and 13 alleles in patients with metastatic colorectal cancer: assessment as prognostic and predictive biomarkers of response to panitumumab. *J Clin Oncol.* 2013;31(6):759-65. <https://doi.org/j7rf>.
64. Peeters M, Oliner KS, Parker A, Siena S, Van Cutsem E, Huang J, *et al.* Massively parallel tumor multigene sequencing to evaluate response to panitumumab in a randomized phase III study of metastatic colorectal cancer. *Clin Cancer Res.* 2013;19(7):1902-12. <https://doi.org/j7rg>.
65. Neumann J, Zeindl-Eberhart E, Kirchner T, Jung A. Frequency and type of KRAS mutations in routine diagnostic analysis of metastatic colorectal cancer. *Pathol Res Pract.* 2009;205(12):858-62. <https://doi.org/ch5s82>.
66. Abubaker J, Bavi P, Al-Haqawi W, Sultana M, Al-Harbi S, Al-Sanea N, *et al.* Prognostic significance of alterations in KRAS isoforms KRAS-4A/4B and KRAS mutations in colorectal carcinoma. *J Pathol.* 2009;219(4):435-45. <https://doi.org/btwj9g>.
67. Bader AG, Kang S, Vogt PK. Cancer-specific mutations in PIK3CA are oncogenic in vivo. *Proc Natl Acad Sci U S A.* 2006;103(5):1475-9. <https://doi.org/d4vxjr>.
68. Samuels Y, Ericson K. Oncogenic PI3K and its role in cancer. *Curr Opin Oncol.* 2006;18(1):77-82. <https://doi.org/cmr8j3>.
69. Manning BD, Cantley LC. AKT/PKB signaling: navigating downstream. *Cell.* 2007;129(7):1261-74. <https://doi.org/bmgbc4>.
70. Samuels Y, Waldman T. Oncogenic mutations of PIK3CA in human cancers. *Curr Top Microbiol Immunol.* 2010;347:21-41. <https://doi.org/dngh7p>.
71. Hsieh LL, Er TK, Chen CC, Hsieh JS, Chang JG, Liu TC. Characteristics and prevalence of KRAS, BRAF, and PIK3CA mutations in colorectal cancer by high-resolution melting analysis in Taiwanese population. *Clin Chim Acta.* 2012;413(19-20):1605-11. <https://doi.org/j7rh>.
72. Cathomas G. PIK3CA in Colorectal Cancer. *Front Oncol.* 2014;4:35. <https://doi.org/gprrq4>.
73. De Roock W, Claes B, Bernasconi D, De Schutter J, Biesmans B, Fountzilas G, *et al.* Effects of KRAS, BRAF, NRAS, and PIK3CA mutations on the efficacy of cetuximab plus chemotherapy in chemotherapy-refractory metastatic colorectal cancer: a retrospective consortium analysis. *Lancet Oncol.* 2010;11(8):753-62. <https://doi.org/ff8zv5>.
74. Soeda H, Shimodaira H, Gamoh M, Ando H, Isobe H, Suto T, *et al.* Phase II trial of cetuximab plus irinotecan for oxaliplatin- and irinotecan-based chemotherapy-refractory patients with advanced and/or metastatic colorectal cancer: evaluation of efficacy and safety based on KRAS mutation status (T-CORE0801). *Oncology.* 2014;87(1):7-20. <https://doi.org/fs8w84>.
75. Cantwell-Dorris ER, O'Leary JJ, Sheils OM. BRAFV600E: implications for carcinogenesis and molecular therapy. *Mol Cancer Ther.* 2011;10(3):385-94. <https://doi.org/b8d8qv>.
76. Samowitz WS, Sweeney C, Herrick J, Albertsen H, Levin TR, Murtaugh MA, *et al.* Poor survival associated with the BRAF V600E mutation in microsatellite-stable colon cancers. *Cancer Res.* 2005;65(14):6063-9. <https://doi.org/b7t4np>.
77. Segura-Urbe J, Santiago-Payán H, Quintero A. Transitions and transversions in Ki-ras gene in colorectal cancers in Mexican patients. *Tumori.* 2003;89(3):259-62. <https://doi.org/j7rs>.
78. Vaughn CP, ZoBell SD, Furtado LV, Baker CL, Samowitz WS. Frequency of KRAS, BRAF, and NRAS mutations in colorectal cancer. *Genes, Chromosomes and Cancer.* 2011;50(5):307-12. <https://doi.org/bsrv3x>.
79. Herreros-Villanueva M, Gomez-Manero N, Muniz P, Garcia-Giron C, Coma del Corral MJ. PIK3CA mutations in KRAS and BRAF wild type colorectal cancer patients. A study of Spanish population. *Mol Biol Rep.* 2011;38(2):1347-51. <https://doi.org/c7nv5n>.
80. Velho S, Oliveira C, Ferreira A, Ferreira AC, Suriano G, Schwartz S, *et al.* The prevalence of PIK3CA mutations in gastric and colon cancer. *Eur J Cancer.* 2005;41(11):1649-54. <https://doi.org/fp7qms>.
81. Pinto P, Rocha P, Veiga I, Guedes J, Pinheiro M, Peixoto A, *et al.* Comparison of methodologies for KRAS mutation detection in metastatic colorectal cancer. *Cancer Genet.* 2011;204(8):439-46. <https://doi.org/bskwcx>.
82. Ogino S, Kawasaki T, Brahmandam M, Yan L, Cantor M, Namgyal C, *et al.* Sensitive sequencing method for KRAS mutation detection by Pyrosequencing. *J Mol Diagn.* 2005;7(3):413-21. <https://doi.org/d75tss>.
83. Gil-Ferreira C, Aran V, Zalcborg-Renault I, Victorino AP, Salem JH, Bonamino MH, *et al.* KRAS mutations: variable incidences in a Brazilian cohort of 8,234 metastatic colorectal cancer patients. *BMC Gastroenterol.* 2014;14:73. <https://doi.org/gbf2f6>.
84. Frayling IM. Methods of molecular analysis: mutation detection in solid tumours. *Mol Pathol.* 2002;55(2):73-9. <https://doi.org/b5c8vm>.

85. Arnold K. Journal to encourage analysis by sex/ethnicity. *J Natl Cancer Inst.* 2000;92(19):1561. <https://doi.org/b4qqx5>.
86. Edgren G, Liang L, Adami HO, Chang ET. Enigmatic sex disparities in cancer incidence. *Eur J Epidemiol.* 2012;27(3):187-96. <https://doi.org/fzxcsz>.
87. Zahm SH, Fraumeni JF. Racial, ethnic, and gender variations in cancer risk: considerations for future epidemiologic research. *Environ Health Perspect.* 1995;103(Suppl 8):283-6. <https://doi.org/c26cpb>.
88. Cook MB, Dawsey SM, Freedman ND, Inskip PD, Wichner SM, Quraishi SM, *et al.* Sex disparities in cancer incidence by period and age. *Cancer Epidemiol Biomarkers Prev.* 2009;18(4):1174-82. <https://doi.org/d6hm48>.
89. Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. *Gut.* 2017;66(4):683-91. <https://doi.org/f9tq5n>.
90. Gunes EG, Pinarbasi E, Pinarbasi H, Silig Y. Strong association between lung cancer and the AXIN2 polymorphism. *Mol Med Rep.* 2009;2(6):1029-35. <https://doi.org/dr6xzf>.
91. Jho EH, Zhang T, Domon C, Joo CK, Freund JN, Costantini F. Wnt/ β -Catenin/Tcf Signaling Induces the Transcription of Axin2, a Negative Regulator of the Signaling Pathway. *Mol Cell Biol.* 2002;22(4):1172-83. <https://doi.org/bbfkrh>.
92. Arzoo PS, Klar J, Bergendal B, Norderyd J, Dahl N. WNT10A mutations account for (1/4) of population-based isolated oligodontia and show phenotypic correlations. *Am J Med Genet A.* 2014;164A(2):353-9. <https://doi.org/f24ps6>.
93. Wu Z, Sun Y, Tang S, Liu C, Zhu S, Wei L, *et al.* AXIN2 rs2240308 polymorphism contributes to increased cancer risk: evidence based on a meta-analysis. *Cancer Cell Int.* 2015;15:68. <https://doi.org/j7rx>.
94. Marvin ML, Mazzoni SM, Herron CM, Edwards S, Gruber SB, Petty EM. AXIN2-associated autosomal dominant ectodermal dysplasia and neoplastic syndrome. *Am J Med Genet A.* 2011;155A(4):898-902. <https://doi.org/cw2d4z>.
95. Zhong AY, Pan X, Shi MH, Xu HJ. -148 C/T polymorphism of Axin2 contributes to a decreased risk of cancer: evidence from a meta-analysis. *Onco Targets Ther.* 2015;8:1957-66. <https://doi.org/j7rz>.
96. Pinarbasi E, Gunes EG, Pinarbasi H, Donmez G, Silig Y. AXIN2 polymorphism and its association with prostate cancer in a Turkish population. *Med Oncol.* 2011;28(4):1373-8. <https://doi.org/b7mw8t>.
97. Gong J, Jiang Y, Hao N, Zhu B, Li Y. Quantitative assessment of the association between AXIN2 rs2240308 polymorphism and cancer risk. *Sci Rep.* 2015;5:10111. <https://doi.org/j7r2>.
98. Staudacher JJ, Yazici C, Bul V, Zeidan J, Khalid A, Xia Y, *et al.* Increased Frequency of KRAS Mutations in African Americans Compared with Caucasians in Sporadic Colorectal Cancer. *Clin Transl Gastroenterol.* 2017;8(10):e124. <https://doi.org/gb4kdp>.
99. Won DD, Lee JI, Lee IK, Oh ST, Jung ES, Lee SH. The prognostic significance of KRAS and BRAF mutation status in Korean colorectal cancer patients. *BMC Cancer.* 2017;17(1):403. <https://doi.org/gbj6gp>.

Appendix 1. Search equations:

MEDLINE (via PubMed)

("Prevalence" AND ("Mutation" OR "Polymorphism") AND ("Genes" OR "RAS" OR "Kras") AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")
("Prevalence" AND ("Mutation" OR "Polymorphism") AND ("Genes" OR "PIK3CA") AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")
("Prevalence" AND ("Mutation" OR "Polymorphism") AND ("Genes" OR "BRAF") AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")
("Prevalence" AND ("Mutation" OR "Polymorphism") AND ("Genes" OR "AXIN2") AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")

Embase (via Ovid)

((("Mutation" OR Polymorphism*) AND "Prevalence" AND ("RAS" OR "Kras") OR "PIK-3CA" OR "BRAF" OR "AXIN2" AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")

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("Prevalence" AND ("Mutation" OR "Polymorphism") AND ("Genes") AND ("Colorectal Neoplasms" OR "Colorectal Cancer") AND "dental agenesis")