Nitrogen uptake by ornamental bromeliad during atmospheric and tank developmental stages

Captación de nitrógeno por las bromelias ornamentales durante las etapas de desarrollo atmosférico y de tanque

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ABSTRACT

The aim of this study was to evaluate the growth of atmospheric (tankless-bromeliad) and tank (tank-bromeliad) developmental stages of silver vase bromeliad (Aechmea fasciata) as a function of nutrition with different sources and concentrations of nitrogen. The experiment consisted of the fertilization of atmospheric and tank stages of silver vase bromeliad with 50 mL of Hoagland and Arnon n. 1 (1950) solution, supplemented with 0, 15, 30 or 45 mM N of urea, ammonium nitrate or glutamine as nitrogen sources. After 210 days of experimentation, tank and atmospheric stages of silver vase bromeliad did not show significant differences for the variables plant height, rosette and stem diameter, leaf and total fresh mass, and root, leaf and total dry matter masses. Both stages have grown better with 15 mM N of ammonium nitrate and urea, while 45 mM of all N sources were deleterious. Glutamine in the tank occasioned the presence of a film and dark spots lowing the ornamental quality. The agronomic nitrogen use efficiency showed that tank stage use N more efficiently, when fertilized with inorganic form, while atmospheric stage is indifferent for N source.

Keywords: urea, nitrate, glutamine, Bromeliaceae, floriculture, mineral nutrition, Aechmea fasciata.

RESUMEN

El objetivo de este estudio fue evaluar el crecimiento de bromelia fasciata (Aechmea fasciata) en etapa atmosférica (periodo en ausencia de tanque y utilización de agua o humedad disponible en el aire) y etapa con presencia de tanque (estructuras desplegadas para captar el agua de lluvia o condensación) en función de la nutrición con diferentes fuentes y concentraciones de nitrógeno. El experimento consistió en fertilizar ambas etapas de la planta con 50 ml de Hoagland y Arnon n. 1 solución (1950), suplementada en concentraciones de 0, 15, 30 y 45 mM N, con urea, nitrato de amonio y glutamina como fuentes nitrogenadas. Después de 210 días del inicio del experimento, no se observaron diferencias significativas para las variables de altura de planta; diámetro de roseta; diámetro del tallo; masa fresca total de hojas y raíces; masas secas de raíces, hojas y materia seca total. Ambas etapas se han desarrollado mejor con 15 mM N de nitrato de amonio y urea, mientras que 45 mM de todas las fuentes de N fueron perjudiciales. La glutamina en el tanque, producto de la fertilización, ocasionó la presencia de una membrana y manchas oscuras que disminuyeron la calidad ornamental. La eficiencia del uso agronómico de nitrógeno mostró que la etapa de tanque usa N de modo más eficiente, cuando se fertiliza con manera inorgánica, mientras que la etapa atmosférica es indiferente para la fuente de N.

Palabras clave: urea, nitrato, glutamina, Bromeliaceae, floricultura, nutrición mineral, Aechmea fasciata.

Introduction

Silver vase bromeliad (Aechmea fasciata (Lindl.) Baker) is the most commercialized bromeliad in floriculture markets, followed by Guzmania (var. Compacta, Empire, Magenta, Cherry and Denise), Vriesea sp. var. Charlote, V. splendens and Neoregelia carolinae as potted

plants for indoor environments, and *Vriesea regina*, *Aechmea blanchetiana*, *Neoregelia compacta* and *Alcantarea imperialis* for landscape use (Vitari, 1994). Silver vase bromeliad is a native bromeliad from Espírito Santo and Rio de Janeiro State, Brazil. The specie has beautiful foliage and inflorescence, and can reach 40 cm height; its coriaceous leaves are arranged as an open rosette forming a tank

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(cistern), with transverse silver bands on leaf blades (Reitz 1983, Wanderley, 1999).

Leaves constitute a major part of bromeliads and are considered the main organs for nutrients and water uptake and organic compounds storage, and can be divided into apical and basal regions (Takahashi *et al.*, 2007). The basal region of the tank is in contact with the solution present into the tank and can uptake water and nutrients by structures termed trichomes or scales that are common on the leaf surface (Benzing, 1990, Takahashi *et al.*, 2007).

The first stage that seedlings of bromeliads develop into so-called atmospherics and presents narrow leaves, which latter change into plants forming the tank by overlapping the broad leaf bases (Meisner et al., 2013). The stages atmospheric and tank show considerable morphoanatomical and physiological differences, with small juveniles having morphological characteristics of atmospherics (non-impounding rosettes of small and linear leaves), whereas larger conspecifics leaves form a tank (overlap broad leaves basally forming water-filled chambers) (Zotz et al., 2004). In relation to water and nutrients supply the small atmospheric plants lack a tank and are "pulse supplied" compared to larger more "continuous supplied" bromeliads with tanks (Zotz and Hietz, 2001). Bromeliads tends to transfer water and nutrient uptake from root towards to tank during plant development from atmospheric to tank stage, even though there is differences in the balance of roots vs. tank uptake among bromeliad cultivars (Vanhoutte et al., 2017).

Bromeliads have a long cultivation cycle as compared to other ornamental horticultural crops. The cultivation cycle of silver vase bromeliad from seeds takes 2 years and 2 months (780 days), divided into 5 phases: sowing (240 days), 34 cells trays (120 days), pot n. 11 (120 days), pot n. 15 (180 days), pot n. 17 (120 days) and another 120 days to flourish and be marketed (Sanches, 2009). Consequently, studies on mineral nutrition of bromeliads are essential to accelerate plant development, reduce production costs such as time in greenhouse, irrigation, man labor, phytosanitary treatments, among others.

Therefore, we aimed to study the effect of nitrogen doses and sources on growth and development of silver vase bromeliad during the atmospheric and tank stages. This knowledge will contribute to a more comprehensive understanding of how to manage the fertilization of bromeliads in horticultural conditions, improving plant growth, and reducing N losses.

Material and methods

The experiment was carried out in a greenhouse with irradiance of 500 µmol m⁻² s⁻¹, daily average temperature of 27.5 °C and watered using microsprinklers irrigation (Modular NaanDanJain® Microaspersor, flow rate 141 L h⁻¹) on two periods during 15 min daily. The analysis of irrigation water showed: pH - 7.7; K - 0.07 mM; Ca - 0.320 mM; Mg - 0.060 mM; Cl - 0.960 mM; Na - 0.100 mM; CO₃ - 0.000 mM; HCO₃ - 0.420 mM; SAR (Sodium Absorption Ratio) - 0.23 and EC - 0.080 dS m⁻¹.

Silver vase bromeliads with atmospheric (without tank, 7.4 leaves, 2.76 mm of stem diameter, 6.73 cm of plant height, 3.02 cm of root length, 0.06522 g of total dry mass, n = 10), and tank (11.6 leaves, 6.18 mm stem diameter, 9.97 cm of plant height, 6.97 cm root length, 0.30504 g of total dry mass, n = 10) stages were used in experiment (Figure 1).

The plants were transplanted into black polypropylene pots with a volumetric capacity of 0.45 L. The substrate used was *Pinus* bark substrate (composted; granulation 6-10 mm; N - 0.50%, P_2O_5 - 0.1%, K_2O - Undetected; Ca - 0.3%, Mg - 0.1%, S - 0.3%, O.M. - 26.0% and C - 14.0 % and 129, 6, 3.68, 52, 13 mg kg⁻¹ of Na, Cu, Fe, Mn and Zn, respectively; C/N - 28/1 and pH - 3.5).

The plants were fertilized three times a week with 50 mL of solution of ion balanced Hoagland and Arnon n. 1 (1950) modified solution with 0, 15, 30 or 45 mM N of urea, ammonium nitrate or glutamine as nitrogen sources.



Figure 1. Tank (A) and atmospheric (B) stages of silver vase plant.

After 210 days of experimentation the variables plant height and rosette diameter, stem diameter, width of leaf and number of leaves were evaluated. Plants were sectioned into roots, stem and leaves and weighted for fresh mass and dried in air-forced oven at 60 °C until constant weight was reached for dry mass.

Nitrogen Agronomic Efficiency index was calculated as follows:

Nitrogen Agronomic Efficiency (NAE, g g⁻¹) = (Total plant dry mass with fertilization - total plant dry mass without fertilization) / (Applied Nitrogen) (adapted from Fageria, 1998);

The experimental design was randomized blocks and consisted of three blocks and twenty four treatments in a factorial scheme of 2 x 3 x 4 (plant stages x nitrogen sources x nitrogen doses). Each plot was composed of five plants, making three hundred and sixty plants, of which 180 tank and 180 atmospheric plant stage. Data were submitted to analysis of variance (F test) and the means were compared by the Tukey test ($p \le 0.05$) using the SISVAR statistical program (Ferreira, 2000).

Results and discussion

Tank and atmospheric stages of silver vase bromeliad did not show significant differences $(p \le 0.05)$ for the variables plant height, rosette and stem diameter, leaf and total fresh mass, and root, leaf and total dry matter masses (Table 1). Despite the tank plants were almost five times heavier (0.305 g DW) than atmospheric plants (0.065 g DW) at the begin of experimentation, in the end both stages had the same weight $(p \le 0.05)$. This

response may be related to the rapid growth of the atmospheric stage plants to establish the tank, while the tank stage plants invested in the developmental structures (e.g. trichomes and scales) in detriment to plant growth. Tank establishment in Guzmania monostachia was positively associated with the emergence of morphological compartmentalization along the leaf blade that facilitates CO₂ uptake by the photosynthesizing cells improving carbon assimilation when stomata are closed (Rodrigues et al., 2016). Atmospheric stages of bromeliads Guzmania monostachia, Guzmania lingulata, and Werauhia sanguinolenta showed photosynthetically active chlorenchyma cells embedded into an upper and lower hydranchyma, which decreases in size as plant stage the tank (Beltran et al., 2013). Water and nutrient uptake by two Vriesea cultivars was shown to be dependent of the growth stage; total uptake of water and nutrients (roots + trichomes) per gram of fresh weight of both cultivars was higher in young plants (Vanhoutte et al., 2017). In addition, the roots of atmospheric stage of *Vriesea* gigantea has an important role on nutrient uptake, while the plants develop the tank on the base of the leaves undertake the uptake function, as the roots decrease its capacity for nitrogen uptake (Takahashi, 2014). On the other hand, for Meisner et al., (2013), the changes from atmospheric to tank stage are gradual and primarily related to size and not to the abrupt switch in habit or stage

Higher concentrations of glutamine, urea and nitrate were detrimental for silver vase bromeliad growth and development, showing the best results at 15 mM N for all N sources. Epiphytic species are exposed to a constant nutritional stress in their natural habitat, thus these plants have a

Table 1. Biometric and biomass variables of silver vase bromeliad during tank and atmospheric stages.

	Height (cm)	Rosette diameter (cm)	Stem diameter (mm)	Stem fresh mass (g)	Root fresh mass (g)	Leaf width (mm)	Number of leaves (un)
Tank	21.8 a	32.43 a	15.24 a	3.46 a	1.64 a	45.48 a	9.1 b
Atmospheric	22.5 a	33.68 a	15.39 a	2.59 b	0.84 b	43.67 b	9.8 a
	Stem dry mass (g)	Root dry mass (g)	Leaf dry mass (g)	Total fresh mass (g)	Leaf dry mass (g)	Total dry mass (g)	
Tank	0.37 a	0.32 a	51.45 a	56.55 a	5.42 a	6.12 a	
Atmospheric	0.32 b	0.30 a	51.24 a	54.67 a	5.69 a	6.31 a	

Means followed by the same letter within a column are not differ significantly from each other at a 5% probability level by the Tukey test.

great efficiency for nitrogen uptake and reach the maximum of its growth potential with low levels of N (Santos et al., 2012). Aechmea blanchetiana seedlings growing in vitro in Murashige and Skoog (MS) (1962) medium modified with different nitrogen concentrations, decreased plant height linearly as N concentration increased, supporting the hypothesis of low nutrient requirement and slow growth of bromeliads in nature (Kanashiro et al., 2007). Fertilization with high concentrations of glutamine showed to be deleterious, mainly for tank plants; however, phytotoxic and detrimental effects of glutamine fertilization in plants were not found in literature. Silver vase bromeliad submitted to high concentrations of glutamine showed a darkening solution into the tank and the formation of a dark film adhered to the central leaves of the tank, which was not observed in urea and ammonium nitrate treatments. The dark solution into the tank and the film adhered to leaves had an impact on ornamental characteristics of the leaves, showing a "dirty" appearance. In addition, the 45 mM glutamine treatment showed the presence of fly larvae (Tipulomorpha infraorder, family Limonidae) inside the tank of all plants. Probably, the glutamine fermentation inside the tank, became a propitious site for attraction, emergence and development of the Diptera larvae.

Stem, root, leaf and total fresh mass variables of atmospheric and tank stages had the lowest mass as N concentrations increase in urea and ammonium nitrate treatments. Whereas for glutamine bromeliads with tank presented a deleterious effect at 45 mM N, while bromeliads without tank did not showed those characteristics. As regards for dry mass, roots were more responsive to the fertilization with high dose of N (45 mM) for all sources, when compared to leaves. Leaves and total dry masses also showed a decrease in the concentration of 45 mM N in all the sources, except for glutamine in atmospheric stage plants (Table 2). Nitrogen isotopic study showed that N source shift as plants age and grow, though small bromeliads depend upon atmospheric inputs, and tank bromeliads reflect the isotopic composition of the leaf litter (Reich et al., 2003). Neoregelia cruenta require 300 days to show the effects of urea fertilization, which is due to the time of the seedlings demand to use nitrogen for amino acid and protein synthesis and as the seedlings

reach full vegetative development, they initiate to respond better to urea fertilization in an increasing linear mode (Ferreira *et al.*, 2007). *Aechmea blanchetiana* seedlings fertilized with potassium nitrate, had the lowest values of leaf length, length of the largest root, number of leaves, dry and fresh masses and percentage of live plants on higher concentrations (Tavares *et al.*, 2008). Studies by Hamasaki *et al.*, (2005) showed that concentrations of 32 mM glutamine was inhibitory and lead to a strong reduction on the frequency of regeneration in pineapple, even though, glutamine is a fast and easy source of nitrogen, and glutamine and glutamate are the main endogenous amino acids involved in plant metabolism.

The chemical analysis of the leaves of silver vase bromeliad (Table 3) showed that there was N uptake for all nitrogen sources used in the experiment, with an increase of N contents in the leaves of all treatments. Epiphytic tank bromeliads tend to prefer an organic nitrogen source, while terrestrial such as pineapple would prefer inorganic nitrogen (Enders and Mercier, 2001, Romero et al., 2006). Atmospheric stage of *V. gigantea* were able to uptake and assimilate inorganic nitrogen rapidly in the 1st hour of the experimental time, while the tank stages absorbed nitrate slowly at the end of the experiment (Takahashi, 2014). Small plants of Vriesea sanguinolenta responded more strongly to increases of water and NPK fertilizer (18-14-18, formulated with ammonium and nitrate), than larger plants (Laube and Zotz, 2003). Although silver vase bromeliad is an epiphytic bromeliad, our data did not shown a preference for organic or inorganic N source; besides glutamine had the highest concentrations in leaves of atmospheric and tank stages, it showed to be deleterious in relation

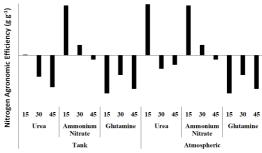


Figure 2. Agronomic efficiency of nitrogen use of tank and atmospheric stages of *Aechmea fasciata* submitted to 15, 30 or 45 mM of urea, ammonium nitrate or glutamine.

Table 2. Biometric and biomass variables of silver vase bromeliad during tank and atmospheric
stages submitted to treatments with glutamine, ammonium nitrate and
urea at concentrations of 0, 15, 30 or 45 mM N.

Glutam		(mM) 0	(cm)	(cm)	(mm)	(mm)	(un)	(g)	(g)	(g)	(g)	(g)	(g)	(g)	(g)
Glutam			24.0 a	32.5 a	15.92 a	48.23 a	10.53 a	2.97 a	0.93 a	55.76 a	59.66 a	0.34 a	0.37 a	5.10 a	6.54 a
Glutam		15	22.5 ab	34.5 a	15.21 a	44.80 ab	9.60 a	2.78 a	1.10 a	56.57 a	60.45 a	0.34 ab	0.30 ab	6.02 a	6.65 a
	Glutamine	30	19.8 b	32.4 a	13.18 a	38.76 b	9.67 a	1.67 b	0.76 a	40.09 a	42.52 a	0.21 b	0.24 b	4.50 a	4.95 a
		45	21.5 b	32.4 a	14.17 a	38.83 b	10.53 a	2.36 ab		44.71 a	47.69 a	0.30 ab		5.10 a	5.66 a
sinc		0	25.6 a	37.4 a	18.47 a	52.80 a	10.07 a	3.61 a	0.94 ab	66.35 a	70.90 a	0.43 a	0.45 a	7.64 a	8.53 a
Ammor	ium	15	24.0 ab	36.0 a	17.02 ab	48.07 ab	9.80 a	2.88 ab	1.28 a	62.41 a	66.57 a	0.34 ab	0.35 ab		7.30 ab
Atmospheric Aumor nitrate		30	22.6 ab	32.0 a	15.99 ab	43.24 bc	9.73 a	2.93 ab	0.77 ab	54.50 ab		0.37 ab	0.31 b		6.97 ab
		45	20.8 b	32.8 a	14.46 b	52.80 c	9.67 a	2.18 b	0.49 b	42.06 b	44.73 b	0.28 b	0.25 b	4.99 b	5.52 b
		0	23.5 ab	33.9 a	16.11 a	46.49 a	9.60 a	2.96 a	1.06 a	51.14 ab	55.15 ab	0.32 a	0.32 ab	5.32 ab	5.96 ab
T	Urea	15	24.2 a	37.2 a	15.50 ab	46.45 a	9.47 a	2.73 ab	0.87 a	58.46 a	62.06 a	0.33 a	0.35 a	6.75 a	7.44 a
Urea		30	21.4 ab	31.6 a	13.81 ab	40.21 ab	9.33 a	2.21 ab	0.53 a	41.65 b	44.40 ab	0.28 a	0.22 b	4.66 b	5.16 b
		45	20.4 b	31.3 a	13.09 b	37.81 b	9.60 a	1.83 b	0.71 a	41.20 b	43.75 b	0.25 a	0.24 b	4.62 b	5.11 b
		0	24.2 a	33.3 a	17.12 a	52.71 a	8.87 a	4.18 a	2.75 a	62.50 a	69.43 a	0.43 a	0.35 a	5.95 a	6.73 a
CI.		15	22.9 a	32.0 a	14.93 ab	45.75 ab	8.67 a	2.98 a	1.52 b	49.29 ab	53.79 ab	0.33 ab	0.30 ab	4.99 a	5.63 ab
Glutam	ne	30	19.2 b	28.4 ab	13.21 bc	38.97 b	8.93 a	2.29 a	1.52 b	39.55 bc	43.11 bc	0.29 ab	0.29 ab	5.01 ab	5.59 ab
		45	15.7 с	23.1 b	12.06 c	31.42 c	9.13 a	1.63 a	0.81 b	44.71 c	31.23 c	0.22 b	0.22 b	3.34 b	3.79 b
		0	23.2 ab	32.1 b	17.02 a	49.42 ab	9.40 a	4.12 a	2.47 a	56.70 a	63.29 ab	0.39 a	0.45 a	5.62 a	6.46 a
≟ Ammor	ium	15	24.1 a	38.4 a	17.67 a	53.44 a	9.40 a	3.79 a	1.78 ab	69.39 ab	74.96 a	0.43 a	0.36 ab	7.12 a	7.91 a
Ammor nitrate		30	23.5 a	35.7 ab	16.33 a	48.07 ab	9.53 a	3.56 a	1.37 b	55.73 ab	60.67 ab	0.42 a	0.31 b	6.33 a	7.05 a
		45	20.0 b	34.4 ab	15.83 a	42.39 b	9.53 a	3.24 a	1.08 b	47.07 b	51.39 b	0.44 a	0.26 b	5.39 a	6.09 a
		0	24.8 a	35.9 a	18.49 a	54.21 a	9.33 a	4.80 ab	2.39 a	63.27 a	70.47 a	0.49 a	0.37 a	6.17 a	7.04 a
**		15	23.5 ab	35.1 a	16.2 ab	48.04 ab	9.27 a	6.27 a	1.78 a	64.64 ab	72.69 a	0.46 a	0.31 a	6.29 a	7.05 a
Urea	Urea	30	21.2 bc	33.0 ab	14.16 b	45.00 b	9.20 a	3.04 bc	1.67 a	46.90 bc	51.61 b	0.36 ab	0.37ab	5.06 ab	5.79 ab
		45	19.2 c	27.8 b	11.61 c	36.30 c	9.33 a	1.65 c	0.73 b	33.56 с	35.94 b	0.23 b	0.22 b	3.82 b	4.26 b

Means followed by the same letters within a column, from each fertilizer source, are not differ significantly from each other at a 5% probability level by Tukey test, being H, height; RD, rosette diameter; SD, stem diameter; LF, leaf width; NL, number of leaves; FSM, fresh stem mass; FRM, fresh root mass; FLM, fresh leaf mass; TFM, total fresh mass; SDM, stem dry mass; RDM, dry mass of the roots; LDM, leaf dry mass and TDM, total dry mass.

to plant quality. Nutrients Ca, S, P, Cu, Fe and B contents did not change with higher doses of N for atmospheric or tank plants. The K, Mg and Mn contents decrease as N levels on treatments increases for both stages of bromeliads. Nitrogen fertilization may result in a dilution effect; in another word, when mineral-element concentrations in shoots decrease as shoot-dry-matter accumulation increase (Riedell, 2010). Such effect may be the main reason why plants did not increase nutrients contents in leaves, besides the fast and vigorous growth of atmospheric and tank stages of *A. fasciata* under nitrogen fertilization. Differently,

Vriesea sanguinolenta showed marked increases in N and P content in all plant size classes, pointing to both "luxury" consumption and storage of N and P, such increase in P concentration, reflecting the switch from vegetative growth to storage and reproductive stage (Zotz et al., 2004).

Conclusions

The results corroborate the data observed for plant growth and development, once the best results were obtained at low N concentrations; moreover, the results also showed that tank bromeliads become

Table 3. Macro and micronutrients content averages of in the leaves of silver vase bromeliad during tank and atmospheric stages submitted to treatments with glutamine, ammonium nitrate and urea at concentrations 0, 15, 30 or 45 mM N.

	Treatment		N	P	K	Ca	Mg	S	В	Cu	Fe	Mn	Zn
N source (mM)		g kg-1						mg kg ⁻¹					
		0	13	2.1	55	5	3.0	1.6	39	3	179	248	23
	Urea	15	25	3.2	53	5	2.7	1.6	36	3	177	169	23
		30	31	3.3	48	5	2.5	1.6	34	2	161	127	20
		45	35	3.0	46	4	1.8	1.6	30	2	163	115	18
		0	14	3.1	59	6	2.9	1.6	33	2	135	238	22
놙		15	23	3.1	55	4	2.9	1.6	32	2	122	154	25
Tank	Ammonium nitrate	30	25	3.1	48	4	2.2	1.5	29	2	117	185	21
		45	30	2.8	42	4	2.2	1.5	26	2	144	156	19
	GI	0	15	2.9	60	5	2.8	1.6	28	2	119	247	19
		15	35	3.1	42	6	2.7	1.6	32	2	198	165	22
	Glutamine	30	29	3.0	48	5	2.2	1.6	30	2	153	142	17
		45	28	2.9	39	5	1.9	1.7	29	2	201	123	14
		0	12	2.7	59	5	3.5	1.6	28	2	102	196	26
	**	15	26	2.6	43	4	2.3	1.5	29	2	128	125	19
	Urea	30	32	2.7	49	5	2.7	1.6	30	2	162	161	31
		45	23	3.0	52	4	2.3	1.6	31	3	164	155	18
ric		0	10	3.0	51	5	3.4	1.6	31	3	104	177	30
bhe		15	21	2.8	54	4	2.8	1.5	30	3	107	162	24
uos	Ammonium nitrate	30	24	2.7	46	4	2.3	1.5	31	3	114	135	25
Atmospheric		45	32	2.7	48	4	2.3	1.5	33	3	152	148	26
		0	11	3.0	58	6	3.5	1.5	33	5	121	261	28
	Glutamine	15	27	3.3	55	5	2.8	1.5	34	4	172	183	24
		30	32	3.1	49	5	2.5	1.5	34	3	126	162	24
		45	34	3.0	46	6	2.3	1.7	32	3	149	167	23
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more efficiently with inorganic N, while atmospheric bromeliads are indifferent about N source. The agronomic efficiency of nitrogen use shows that the tank stage was more efficient on concentration of 15 mM of ammonium nitrate, and 15 mM urea and 30 mM ammonium nitrate doses similarly resulted on positive efficiency. The atmospheric stage was more efficient on concentrations of 15 mM of urea and ammonium nitrate, and the dose of 30 mM of ammonium nitrate was also efficiently. Glutamine was a deleterious nitrogen source for bromeliads at all concentrations.

Acknowledgments

The authors would like to thanks Dr. Luis Filipe Mucci SUCEN for helping to identify the larvae on Bromeliads. The authors also like to thank CNPq/PIBIC (Conselho Nacional de Desenvolvimento Científico e Tecnológico - Programa Institucional de Bolsas de Iniciação Científica) for the scholarship to D.B. Müller and (306140/2012-8) granted for research productivity for A.R. Tavares.

Literature Cited

Beltrán, J.D.; Lasso, E.; Madriñán, S.; Virgo, A.; Winter, K. 2013. Juvenile tank-bromeliads lacking tanks: do they engage in CAM photosynthesis. *Photosynthetica*, 51: 55-62. Benzing, D.H.

1990. Vascular epiphytes. Cambridge University Press. New York, USA. 354 p.

Endres, L.; Mercier, H.

2001. Influence of nitrogen forms of the growth and nitrogen metabolism of bromeliads. *Journal of Plant Nutrition*, 24: 29-42.

Fageria, N.K.

1998. Otimização da eficiência nutricional na produção das culturas. *Revista Brasileira de Engenharia Agrícola e Ambiental*, 2: 6-16.

Ferreira, D.F.

2000. SISVAR - Sistemas de análises de variância para dados balanceados: programa de análises estatísticas e planejamento de experimentos. Versão 4.3. UFLA. Lavras, Brasil.

Ferreira, C.A.; Paiva, P.D.O.; Rodrigues, T.M.; Ramos, T.D.P.; Carvalho, J.G.; Paiva, R.

2007. Desenvolvimento de mudas de bromélia (Neoregelia cruenta (R. Graham) L.B. Smith) cultivadas em diferentes substratos e adubações foliares. Ciência e Agrotecnologia, 31: 666-671.

Hamasaki, R.M.; Purgatto, E.; Mercier, H.

2005. Glutamine enhance competence for organogenesis in pineapple leaves cultivated *in vitro*. *Brazilian Journal of Plant Physiology*, 17: 383-389.

Hoagland, D.R.; Arnon, D.I.

1950. The water culture method for growing plants without soils. California Agricultural Experimental Station. Berkeley, USA. 347 p.

Kanashiro, S.; Ribeiro, R.C.S.; Gonçalves, A.N.; Dias, C.T.S.; Jocys, T.

2007. Efeitos de diferentes concentrações de nitrogênio no crescimento de *Aechmea blanchetiana* (Baker) L.B. Sm. cultivada *in vitro. Hoehnea*, 34: 59-66.

Laube, S.; Zotz, G.

2003. Which abiotic factors limit vegetative growth in a vascular epiphyte? *Functional Ecology*, 17: 598-604.

Meisner, K.; Winkler, U.; Zotz, G.

2013. Heteroblasty in bromeliads-anatomical, morphological and physiological changes in ontogeny are not related to the change from atmospheric to tank form. *Functional Plant Biology*, 40: 251-262.

Murashige, T.; Skoog, F.

1962. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia plantarum*, 15: 473-497.

Reich, A.; Ewel, J.J.; Nadkarin, N.M.; Dawson, T.; Evans, R.D. 2003. Nitrogen isotope ratios shift with plant size in tropical bromeliads. *Oecologia*, 137: 587-590.

Reitz, R.

1983. Bromeliáceas e a malária - bromélia endêmica. Flora Ilustr. Catarinense. Itajaí, Brasil. 559.

Riedell, W.E.

2010. Mineral-nutrient synergism and dilution responses to nitrogen fertilizer in field-grown maize. *Journal of Plant Nutrition and Soil Science*, 173: 869-874.

Rodrigues, M.A.; Hamachi, L.; Mioto, P.T.; Purgatto, E.; Mercier, H.

2016. Implications of leaf ontogeny on drought-induced gradients of CAM expression and ABA levels in rosettes of the epiphytic tank bromeliad *Guzmania monostachia*. *Plant Physiology and Biochemistry*, 108: 400-411.

Romero, G. Q.; Mazzafera, P.; Vasconcellos-Neto, J.; Trivelin, P. C. O. 2006. Bromeliad-living spiders improve host plant nutrition and growth. *Ecology*, 87: 803-808.

Sanches, L.V.C.

2009. Desenvolvimento de Aechmea fasciata (Bromeliaceae) em função de diferentes saturações por bases no substrato e modos de aplicação da fertirrigação. UNESP-FCA. Botucatu, Brasil, 124 p.

Santos, F.H.D.S.; Almeida, E.F.A.; Frazão, J.E.M.; dos Santos, A.C.P.

2012. Nitrogen nutrition of bromeliads. *Ornamental Horticulture*, 18: 39-46.

Takahashi, C.A.

2014. Assimilação do nitrogênio em folhas de Vriesea gigantea (Bromeliaceae) durante a transição ontogenética do hábito atmosférico para o epífito com tanque. USP. São Paulo, Brasil.

Takahashi, C.A.; Ceccantini, G.C.T.; Mercier, H.

2007. Differential capacity of nitrogen assimilation between apical and basal leaf portions of a tank epiphytic bromeliad. Brazilian Journal of Plant Physiology, 19: 119-126.

Tavares, A.R.; Giampaoli, P.; Kanashiro, S.; Aguiar, F.F.A.; Chu, E.P.

2008. Efeito da adubação foliar com KNO₃ na aclimatização de bromélia cultivada in vitro. Horticultura Brasileira, 26: 175-179.

Vanhoutte, B.; Schenkels, L.; Ceusters, J.; De Proft, M.P. 2017. Water and nutrient uptake in *Vriesea* cultivars: Trichomes vs. Roots. *Environmental and Experimental Botany*, 136: 21-30.

Vitari, M.

1994. Bromélia: produção e proteção. *Ecologia e Desenvolvimento*, 3: 15-17.

Wanderley, M.G.L.

1999. Bromélias Brasileiras: aquarelas de Margaret Mee. Instituto de Botânica. São Paulo, Brasil. 159 p.

Zotz, G.; Hietz, P.

2001. The physiological ecology of vascular epiphytes: current knowledge, open questions. *Journal of Experimental Botany*, 52: 2067-2078.

Zotz, G.; Enslin, A.; Hartung, W.; Ziegler, H.

2004. Physiological and anatomical changes during the early ontogeny of the heteroblastic bromeliad, *Vriesea sanguinolenta*, do not concur with the morphological change from atmospheric to tank form. *Plant, Cell & Environment*, 27: 1341-1350.