

# Anisakis and anisakidosis: hosts and case reports in South America. Systematic review

Anisákidos y anisakidosis: reportes de caso y hospedadores en América del Sur. Revisión sistemática

Luis Felipe Falla-Zuñiga<sup>1</sup> 🕩 Jenniffer Alejandra Castellanos-Garzón<sup>1,2</sup> 🕩 Liliana Salazar<sup>1</sup> 🕩 María Carolina Pustovrh<sup>1</sup> 🕩

<sup>1</sup> Universidad del Valle - Faculty of Health - Department of Morphology - Cali - Colombia.

<sup>2</sup> Unidad Central del Valle - Faculty of Engineering - Tuluá - Colombia.

Corresponding author: Jenniffer Aléjandra Castellanos-Garzón. Departamento de Morfología, Facultad de Salud, Universidad del Valle. Cali. Colombia. Email: jenniffer.castellanos@correounivalle.edu.co.

#### Abstract

**Introduction:** Anisakidosis is a disease caused by the consumption of raw or undercooked seafood parasitized by nematode larvae of the family Anisakidae. Even though it is a public health issue in Europe and Asia, it is relatively unknown in South America.

**Objective:** To present case reports on anisakidosis and the intermediate hosts of *Anisakis* reported in South America.

**Materials and methods:** A systematic review was conducted in Medline, Cochrane, Embase, LILACS and Scopus using a structured search of MeSH and DeCS descriptors. The search strategy included publication period: inception of each database-September 2018; languages: English, Spanish, and Portuguese; and study types: case reports and observational cross-sectional studies. The review was complemented with an unstructured search in SciELO and Google Scholar.

**Results:** The initial search yielded 172 articles. After removing duplicates and reviewing the inclusion criteria, 69 studies were selected for full analysis: 19 case reports and 50 host records. The most reported form of anisakidosis was gastrointestinal anisakidosis with 45 cases; this infectious disease was caused by a single larva in 41 people (91.1%). Reports of 95 species of fish for human consumption parasitized by larvae of the genera *Anisakis, Contracaecum, Pseudoterranova* and *Hysterothylacium* were identified in Argentina (22 fish species), Brazil (34 species), Chile (15 species), Colombia (17 species), Ecuador (8 species), Peru (7 species), Uruguay and Venezuela (2 species each).

**Conclusion:** Anisakidosis is a latent risk in South America, so it is necessary to establish effective regulations for efficiently controlling the appearance of this parasitic disease in the region. Furthermore, the general population should receive more information about the precautions regarding saltwater fish consumption.

**Keywords:** Anisakis; Anisakiasis; South America; Zoonoses; Communicable Diseases, Emerging (MeSH).

### Resumen

**Introducción.** La anisakidosis es una parasitosis ocasionada por el consumo de pescado de mar crudo o semicrudo parasitado por larvas de nematodos de la familia Anisakidae. En Europa y Asia es un problema de salud pública; sin embargo, en América del Sur es poco conocida.

**Objetivo.** Identificar los reportes de caso de anisakidosis y los hospedadores intermediarios de anisákidos reportados en América del Sur.

**Materiales y métodos.** Se realizó una revisión sistemática en Medline, Cochrane, Embase, LILACS y Scopus mediante la búsqueda estructurada de términos MeSH y DeCS. Estrategia de búsqueda: periodo de publicación: inicio de cada base de datos-septiembre de 2018; idiomas: inglés, español y portugués; tipos de estudio: reportes de caso y estudios transversales observacionales. La revisión fue complementada con una búsqueda no estructurada en SciELO y Google Scholar.

**Resultados.** La búsqueda inicial arrojó 172 artículos. Una vez removidos los duplicados y revisados los criterios de inclusión, se seleccionaron 69 estudios para análisis completo: 19 reportes de caso y 50 registros de hospedadores. La forma de anisakidosis más reportada fue la gastrointestinal, con 45 casos, donde la parasitosis fue causada por una larva única en 41 casos (91.1%). Se identificaron reportes de 95 especies de peces para consumo humano parasitadas por los géneros Anisakis, Contracaecum, Pseudoterranova e Hysterothylacium en los siguientes países: Argentina (22 especies), Brasil (34 especies), Chile (15 especies), Colombia (17 especies), Ecuador (8 especies), Perú (7 especies), Venezuela (4 especies) y Uruguay (2 especies).

**Conclusión.** La anisakidosis es un riesgo latente para América del Sur, por lo que es necesario instaurar normativas efectivas para controlar su aparición en la región y brindar más información a la población general sobre las precauciones necesarias en relación con el consumo de pescado de agua salada.

**Palabras clave:** Anisakis; Anisakiasis; América del Sur; Zoonosis; Enfermedades Transmisibles Emergentes (DeCS).

Falla-Zuñiga LF, Castellanos-Garzón JA, Salazar L, Pustovrh MC. *Anisakis* and anisakidosis: Hosts and case reports in South America. Systematic review. Rev. Fac. Med. 2021;69(2):e79105. English. doi: https://doi.org/10.15446/revfacmed. v69n2.79105.

Falla-Zuñiga LF, Castellanos-Garzón JA, Salazar L, Pustovrh MC. [Anisákidos y anisakidosis: Reportes de caso y hospedadores en América del Sur. Revisión sistemática]. Rev. Fac. Med. 2021;69(2):e79105. English. doi: https://doi.org/10.15446/ revfacmed.v69n2.79105.

# Introduction

Anisakidosis is a parasitic disease that affects humans. Although most patients are asymptomatic, it can cause gastrointestinal symptoms and allergic or gastroallergic reactions.<sup>1</sup> This infection occurs when third-stage larvae (L3) of parasitic nematodes of the family Anisakidae are ingested through raw or undercooked fish or cephalopods.<sup>2</sup>

According to Jofré *et al.*,<sup>3</sup> the term anisakidosis was introduced by Straub in 1960, the same year in which Van Thiel and colleagues reported the first case of this disease in the Netherlands.

The symptoms of anisakidosis are explained by two pathophysiological mechanisms: an immediate hypersensitivity reaction and an inflammatory reaction.<sup>4</sup> Therefore, as mentioned above, symptoms may range from allergic reactions to gastrointestinal manifestations. This infection, which is more frequently observed in adults, is usually caused by a single larva, although there are case reports of more than one larva.<sup>4</sup>

Some of the species associated with anisakidosis are *Anisakis simplex, Anisakis physeteris* and *Pseudoterova decipiens*, as well as the genus *Hysterothylacium*.<sup>3,5,6</sup> To diagnose this disease, the symptoms of the patients and their history of consumption of raw or undercooked fish or cephalopods must be reviewed. In the presence of gastrointestinal symptoms, it is necessary to detect and identify the larvae, either in vivo by endoscopy or in situ with a biopsy.<sup>4</sup> Depending on the location of the larva, anisakidosis may be gastric, intestinal, or extraintestinal (lung, liver, and pancreas).<sup>7</sup>

On the other hand, in the presence of allergic symptoms, it is not necessary to directly visualize the larva since the symptoms are mainly attributed to the immune response of the host and the diagnosis is usually made using serological tests.<sup>1</sup> It should be noted that when the result of these tests is positive, a cross-reaction with other ascarids should be ruled out.<sup>8</sup> The allergic and gastroallergic forms are characterized by urticaria, angioedema or anaphylaxis, along with digestive symptoms.<sup>9,10</sup>

In Europe and Asia, more than 2 000 cases of anisakidosis are reported each year; for this reason, as stated by Audícana *et al.*,<sup>11</sup> authorities have extensively studied the disease and have established rules for its control and prevention. On the contrary, in South America, despite having a large fishery industry, anisakidosis is not a common disease and, therefore, its clinical manifestations are little known by the healthcare staff. This creates difficulties with the diagnosis and could lead to underreporting of the disease.<sup>12</sup> Likewise, the information available in the region about this disease is mostly found in some research papers and in a few case reports published in specialized journals,<sup>13-15</sup> but there is no updated review of the subject.

Consequently, the objective of this review was to describe intermediate hosts and identify case reports of anisakidosis published in South America, with a focus on the major marine fish species that could be involved in their transmission, given that this infection is considered a potential emerging disease in the region that needs to be known, studied, and treated.

# **Materials and methods**

A systematic review was conducted to answer two questions: What are the clinical cases of anisakidosis reported in South America? and What are the fish species reported as hosts of anisakid nematodes in South America?

### Eligibility criteria

Case reports and cross-sectional observational studies were included in the search. Inclusion criteria for case reports on anisakidosis were that one or more larvae of the family Anisakidae had been identified in the patients that had a history of fish consumption likely to be parasitized by nematode larvae from this family and that came from any South American country.

Also, to identify the hosts, studies reporting cases of fish for human consumption parasitized by nematodes of the family *Anisakidae* and captured in South American waters were included. Studies that did not have the information required to determine their eligibility and whose authors did not respond to the request for such data were excluded.

#### Search strategy

A structured search using MeSH and DeCS terms was performed in Medline, Cochrane, Embase, LILACS and Scopus databases based on the following search strategy: publication period: from the inception of each database until September 2018; languages: English, Spanish and Portuguese; type of studies: case reports and cross-sectional observational studies; search terms: "Anisakids", "Anisakiasis", "Anisakidosis", "Anisakidae", "Anisakids", "Pseudoterranova" and "Contracecum", which were combined with each of the South American country names (Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Guyana, Paraguay, Peru, Suriname, Uruguay and Venezuela).

The following is the search equation was used in MEDLINE and Cochrane (tiab means Title/Abstract): ((((anisakis [tiab] OR pseudoterrova [tiab] OR anisakidae [tiab] OR Anisakiosis [tiab] OR ANISAKIASIS [tiab] OR) AND (Argentina [tiab] OR BOLIVIA [tiab] OR BRA-ZIL [tiab] OR CHILE [tiab] OR COLOMBIA [tiab] OR ECUADOR [tiab] OR FRENCH GUIANA [tiab] OR GUYANA [tiab] OR PARAGUAY [tiab] OR PERU [tiab] OR Suriname [tiab] OR Uruguay [tiab] OR Venezuela [tiab] OR)) OR (("South America"[Mesh]) AND "Anisakis"[Mesh]).

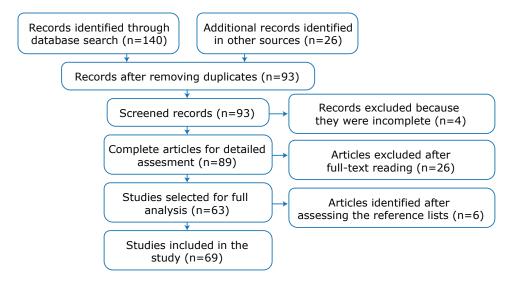
The database search was complemented by an unstructured search in SciELO and Google Scholar and with additional studies recommended by experts in the field. Similarly, the reference lists of the included narrative reviews were assessed to identify publications potentially relevant to the objective of the study, which were also included in the analysis.

The prevalence of infection in articles reporting unspecified hosts was calculated as the ratio between the number of fish parasitized by species of the family *Anisakidae* and the number of fish reviewed, if available.

#### Results

The initial search yielded 166 results (140 from the databases and 26 from SciELO, Google Scholar and

expert recommendations), of which 73 were excluded because they were duplicates, 4 because they were incomplete and 26 because they did not meet the inclusion criteria. Subsequently, the reference lists of the 63 publications found were assessed and 6 additional articles were identified, resulting in a total of 69 publications to be included in the review: 16 case reports, 3 seroprevalence studies, and 50 host records (Figure 1). The characteristics of the selected articles are described in Table 1.



**Figure 1.** Study search and selection flowchart. Source: Own elaboration.

Authors Year	Place of publication	Type of publication	Human/ Host	Sample size (n)	Relevance
Figueiredo <i>et al.</i> 2013	Brazil	RA	Humans	67	Positive anti- <i>Anisakis simplex</i> immunoglobulin E response
Jofré <i>et al.</i> <sup>3</sup> 2008	Chile	CR	Humans	1	Infection with <i>Pseudoterrova decipiens</i> after the ingestion of sushi
Castellanos <i>et al.</i> <sup>12</sup> 2017	Colombia	RA	Hosts	15	Presence of anisakid nematode larvae parasitizing <i>Mugil cephalus</i>
Cabrera & Suárez-			Hosts	2	Two probable cases of anisakidosis
Ognio <sup>13</sup> 2002	Peru	С	Humans	12	Presence of <i>Anisakis</i> larvae in <i>Coryphaena</i> hippurus
Cabrera & Trillo- Altamirano <sup>14</sup> 2004	Peru	SpC	Humans	1	Probable case of infection by a L4 <i>P. decipiens</i> larvae
Mercado <i>et al.</i> <sup>15</sup> 2001	Brazil	SC	Humans	7	Identification of seven cases of infection by L4 <i>P. decipiens</i> larvae
Torres <i>et al.</i> <sup>16</sup> 2007	Chile	RN	Humans	4	Outbreak of pseudoterranovosis in three out of four people who shared the same dish of raw fish (ceviche)
Weitzel <i>et al.</i> <sup>17</sup> 2015	Chile	LE	Humans	3	Report of three cases of infection with Pseudoterranova cattani
Mercado <i>et al.</i> <sup>18</sup> 2006	Chile	CLC	Humans	1	Patient infected with a L4 <i>Pseudoterranova</i> sp. larva
Menghi <i>et al.</i> <sup>19</sup> 2011	Argentina	CLC	Humans	1	Larva of the <i>Anisakis-Contraecum</i> complex in the stool of a girl
Mayo-Iniguez <i>et al.</i> 20 2014	Argentina	SC	Hosts	15	Molecular diagnosis of <i>Anisakis typica</i> and <i>Anisakis physeteris</i> in larvae of hosts of the Brazilian coast
Timi <i>et al.</i> <sup>21</sup> 2014	Argentina	RA	Hosts	34	Molecular diagnosis of <i>P. cattani</i> in fish from Argentine waters

# Table 1. General characteristics of articles selected for review. (continued)

Table 1. General character		1			/
Authors Year	Place of publication	Type of publication	Human/ Host	Sample size (n)	Relevance
Wadnipar-Cano <sup>22</sup> 2014	Colombia	т	Hosts	360	Parasitic infestation by anisakid nematodes in river fish in the municipality of San Marcos, Colombia.
Dias <i>et al.</i> <sup>23</sup> 2010	Brazil	OA	Hosts	100	Parasitation by <i>Anisakis</i> spp. and <i>Contraecum</i> sp. of <i>Aluterus monoceros</i> purchased in markets of the municipalities of Niteroi and Rio de Janeiro, Brazil
Cabrera <i>et al.</i> <sup>24</sup> 2003	Peru	PR	Humans	1	Case of human anisakidosis due to L3 <i>P. decipiens</i> larva.
Tanteleán & Huiza <sup>25</sup> 1993	Peru	PR	Humans	2	<i>P. decipiens</i> larvae obtained from the mouth of two people in Lima, Peru
Barriga <i>et al.</i> <sup>26</sup> 1999	Peru	PR	Humans	1	Extraction of larvae by endoscopy, subsequently identified as <i>Anisakis</i> spp., in a patient from Lima, Peru
Rosa-da Cruz <i>et al.</i> <sup>27</sup> 2010	Brazil	SC	Humans	1	First evidence of larvae similar to <i>Anisakis</i> spp. causing gastrointestinal lesions in Brazil
Patiño & Olivera <sup>28</sup> 2019	Colombia	СР	Humans	1	First case of anisakiasis in Colombia
Puccio <i>et al.</i> <sup>29</sup> 2008	Venezuela	OA	Humans	144	Report of a high percentage (45%) of children with positive skin tests for <i>A. simplex</i> extract
Verhamme & Ramboer <sup>30</sup> 1988	Chile	CR	Humans	6	Case compatible with gastrointestinal anisakidosis after eating salmon in Chile
Torres <i>et al.</i> <sup>31</sup> 2000	Chile	CR	Humans	1	Elimination of L2 anisakid larvae in a man from Santiago de Chile who had previously eaten shellfish and raw fish.
Mercado <i>et al.</i> <sup>32</sup> 1997	Chile	Ν	Humans	1	Extraction of L4 <i>P. decipiens</i> larva during gastrointestinal biopsy of the stomach in a man from southern Chile
Figueiredo <i>et al.</i> <sup>33</sup> 2015	Brazil	OA	Humans	309	Reactivity to anti- <i>Anisakis</i> in pregnant mothers in Brazil
Mancini <i>et al.</i> <sup>34</sup> 2014	Argentina	OA	Hosts	1402	Identification of <i>Contraecum</i> sp. larvae compatible with type 2 L3 larvae in 9 fish species from 19 aquatic environments in Argentina
Ulloa-Ulloa & Carrasco- Mancero <sup>35</sup> 2008	Colombia	т	Hosts	167	Identification of three fish species from Ecuador parasitized by <i>Contracecum</i> sp.
Oliverero-Verbel & Baldiris-Avila <sup>36</sup> 2008	Colombia	В	Hosts	Unknown	Identification of multiple species of fish parasitized by nematodes the family Anisakidae in Colombia
Hernández-Orts <i>et al.</i> <sup>37</sup> 2013	Argentina	RA	Hosts	542	Identification of <i>Pseudoterranova</i> sp. larvae and molecular identification of <i>P. cattani</i> in 12 fish species from Patagonia, Argentina.
Castellanos <i>et al.</i> 38	Ecuador.				Identification by taxonomic revision of 8 species of host fish of the genera <i>Anisakis</i>
2018	Colombia	OA	Hosts	438	sp. and <i>Pseudoterranova</i> sp. from Ecuador and Colombia
Luque & Alves <sup>39</sup> 2001	Brazil	OA	Hosts	115	Identification of anisakid nematodes in two fish species from Brazil
Knoff <i>et al.</i> <sup>40</sup> 2001	Brazil	OA	Hosts	217	Identification of multiple genera of the family Anisakidae parasitizing elasmobranch fish in Brazil.
Rodrigues <sup>41</sup> 2010	Brazil	т	Hosts	52	Presence of anisakids in a fish species marketed in Brazil

# Table 1. General characteristics of articles selected for review. (continued)

Table 1. General character		les selected	TOI TEVIEW. (	continueu	1)		
Authors Year	Place of publication	Type of publication	Human/ Host	Sample size (n)	Relevance		
Timi <i>et al.</i> <sup>42</sup> 2000	Argentina Uruguay	OA	Hosts	2086	Identification of four species of anisakid nematode larvae in a fish species from Argentine and Uruguayan waters.		
Chavez <i>et al.</i> <sup>43</sup> 2007	Chile	СР	Hosts	300	Larvae of <i>Anisakis</i> sp. nematodes found in samples of a fish species obtained in two localities in Chile		
Bracho-Espinoza <i>et al.</i> <sup>44</sup> 2013	Venezuela	OA	Hosts	180	Identification of anisakids in three species of fish from Venezuela		
Marigo <i>et al.</i> <sup>45</sup> 2015	Brazil	CR	Hosts	1	Identification of Pseudoterranova azarasi in cod sold for human consumption in Brazil		
de Paula Toledo Prado & Capuano <sup>46</sup> 2006	Brazil	SC	Hosts	11	Presence of nematodes larvae of the family Anisakidae in cod samples from a Brazilian locality		
Torres <i>et al.</i> <sup>47</sup> 2014	Chile	OA	Hosts	280	Identification of <i>Pseudoterranova</i> sp. in <i>Thyrsites atun</i> from Chile, and of other anisakid larvae in two other fish species.		
Di Azevedo & Iñiguez <sup>48</sup> 2018	Brazil	RA	Hosts	180	Molecular identification of <i>Hysterothylacium dearddorffoverstreetorum</i> (s.l) in three fish species from Brazil.		
Pardo <i>et al.</i> <sup>49</sup> 2007	Colombia	OA	Hosts	45	Identification of <i>Salminus affinis</i> fish from the Sinú and San Jorge rivers parasitized by <i>Contracecum</i> sp. anisakids		
Knoff <i>et al.</i> <sup>50</sup> 2013	Brazil	OA	Hosts	87	Collection of anisakid in larval stages in Lophius gastrophysus specimens from Brazil		
Torres-Frenzel <sup>51</sup> 2013	Chile	т	Hosts	78	Isolation of L3 anisakid <i>Pseudoterranova</i> sp. nematodes in ceviche servings in Chilean restaurants		
Torres <i>et al.</i> <sup>52</sup> 1993	Chile	RN	Hosts	57	Identification of nematode larvae of the family Anisakidae in five fish species in southern Chile		
Vicente <i>et al.</i> <sup>53</sup> 1989	Venezuela	OA	Hosts	136	Presence of <i>Contracecum</i> sp. (s.l) in <i>Micropogonias furnieri</i> from Venezuela		
Fernández <i>et al.</i> <sup>54</sup> 2016	Chile	BC	Host	1	Identification of larval forms of <i>Anisakis</i> sp. (Type I, L3) in the intestinal serosa of ocean sunfish from Chile		
Olivero-Verbel <i>et al.</i> <sup>55</sup> 2005	Colombia	OA	Hosts	386	Presence of L3 larvae of the family Anisakidae in mugilids from two locations on the Colombian Atlantic Coast		
Soares <i>et al.</i> <sup>56</sup>	Argentina	OA	Hosts	186	Identification of anisakid genus in a fish species from Argentine and Uruguayan		
2018	Brazil				waters.		
Saad & Luque <sup>57</sup> 2009	Brazil	RN	Hosts	36	Collection of <i>Anisakis</i> sp. and <i>Contraecum</i> sp. larvae in fish from the coastal zone of Rio de Janeiro, Brazil		
Soares <i>et al.</i> <sup>58</sup> 2014	Brazil	OA	Hosts	100	Presence of <i>Pseudoterranova</i> sp. larvae in fish from the coast of Cabo Frio, Brazil		
Paraguassú <i>et al.<sup>59</sup></i> 2002	Brazil	OA	Hosts	90	Collection of parasites of the family Anisakidae in fish from the Brazilian coast		
Farias-Rabelo <i>et al.</i> <sup>60</sup> 2017	Brazil	OA	Hosts	25	Identification of fish infested with L3 larvae of the family Anisakidae in Brazil		
Braicovich & Timi <sup>61</sup> 2008	Argentina Uruguay	OA	Hosts	177	Presence of three genera of the family Anisakidae in fish caught in fishing waters located between Argentina and Uruguay		

# Table 1. General characteristics of articles selected for review. (continued)

Table 1. General characteristics of articles selected for review. (continued)					
Authors Year	Place of publication	Type of publication	Human/ Host	Sample size (n)	Relevance
Pantoja <i>et al.</i> <sup>62</sup> 2015	Brazil	OA	Hosts	50	Molecular identification of <i>A. typica</i> and <i>Hysterothylacium</i> sp. larvae in two fish species from Rio de Janeiro, Brazil
Kuraiem <i>et al.</i> <sup>63</sup> 2016	Brazil	OA	Hosts	30	Identification of species marketed in Rio de Janeiro, Brazil, parasitized by <i>Anisakis</i> sp. and <i>Hysterothylacium</i> <i>deardorfoverstreetorum</i> larvae
Timi & Lanfranchi <sup>64</sup> 2009	Argentina	RA	Hosts	100	Identification of larvae of the family Anisakidae in a species of fish inhabiting the Argentine sea
Ramallo & Torres <sup>65</sup> 1995	Argentina	OA	Hosts	10	Isolation and morphological identification of <i>Contraecum</i> sp. in a fish species from the of Rio Hondo pond, Argentina
González <i>et al.</i> <sup>66</sup> 2006	Argentina Chile Peru	OA	Hosts	626	Presence of endoparasites of the genus <i>Anisakis</i> sp. in a species of fish present on the Pacific Coast of South America
Hamann <sup>67</sup> 1999	Argentina	OA	Hosts	237	Finding of fish from northeastern Argentina parasitized by <i>Contracecum</i> sp. larvae
Mattiucci <i>et al.</i> 68 2002	Brazil	OA	Hosts	6	Detection of <i>A. typica</i> in multiple fish on the Atlantic Coast of Brazil
Peña-Rehbein <i>et al.</i> <sup>69</sup> 2012	Chile	RN	Hosts	20	Description of the frequency and number of <i>Anisakis</i> spp. nematodes in the internal organs of <i>T. atun</i> fish from Queule, Brazil
Oliva <sup>70</sup> 1999	Chile Peru	OA	Hosts	3034	Identification of anisakid nematodes in a fish species whose specimens were captured in Chile and Peru.
Novo-Borges <i>et al.</i> <sup>71</sup> 2012	Brazil	RA	Hosts	64	Morphological and molecular identification of <i>A. typica</i> and <i>Hysterothylacium</i> sp. larvae in two fish species from Brazil
Braicovich <i>et al.</i> <sup>72</sup> 2017	Argentina	RA	Hosts	488	Identification of larvae of the family Anisakidae in fish from the coastal region of South America between Rio de Janeiro and
	Brazil				northern Argentina
Andrade-Porto <i>et al.</i> <sup>73</sup> 2015	Brazil	OA	Hosts	100	Identification of <i>Arapaima gigas</i> parasitized by L3 larvae of <i>Hysterothylacium</i> sp.
Torres <i>et al.</i> <sup>74</sup> 1998	Chile	OA	Hosts	80	Identification of <i>Hysterothylacium geschei</i> larvae in fish from Brazil
Maniscalchi Badaoui <i>et</i> <i>al.</i> <sup>75</sup> 2015	Venezuela	OA	Hosts	913	Identification of fresh fish of popular consumption in Venezuela parasitized by anisakid nematodes
Ruiz & Vallejo <sup>76</sup> 2013	Colombia	OA	Hosts	378	Identification of <i>Contracecum</i> sp. and <i>Pseudoterrova</i> sp. nematode larvae in <i>Mugil</i> cephalus from the Colombian Caribbean
Bicudo <i>et al.</i> <sup>77</sup> 2005	Brazil	OA	Hosts	80	Identification of larvae of anisakid nematodes, <i>Anisakis</i> sp. and <i>Hysterothylacium</i> sp. in fish from the coastal zone of Rio de Janeiro, Brazil
Knoff <i>et al.</i> <sup>78</sup> 2012	Brazil	OA	Hosts	60	Characterization of larvae in fish from the state of Rio de Janeiro as H. <i>deardorffoverstreetorum</i> sp. nov. larvae

RA: research article; CR: case report; C: communication; SpC: special contribution; SC: short communication; RN: research note; LE: letter to editor; CLC: clinical case; T: thesis; OA: original article; CP: case presentation; N: notes and information; B: book; CA: conference article; BC: brief communication. Source: Own elaboration.

#### Clinical cases of anisakidosis in South America

In Europe and Asia, anisakidosis is considered a public health issue due to the large number of cases reported with gastric, allergic and gastroallergic symptoms; however, in South America, it is still a little-known disease. Table 2 presents a summary of clinical cases associated with anisakid parasites in South America published between 1976 and 2018.

Table 2. Clinical cases associated with anisakid nematodes in South American countries.

No	Year	Country	Patient	Food previously ingested	Symptoms	Identified anisakid	Symptoms	Diagnostic method	Ref
1	1976	Chile	No description	Ceviche	Gastrointestinal	<i>Pseudoterranova</i> sp.	Gastrointestinal	Observation of expelled larvae	3
2	1980	Chile	No description	Chilean jack mackerel	Gastrointestinal	Anisakis sp.	Gastrointestinal	Gastric endoscopy	3
3	1980	Chile	35-year- old woman	No data	Esophageal pain and productive cough	Anisakis sp.	Gastrointestinal	Observation of expelled larvae	32
4	1985	Chile- Belgium	62-year- old man	Raw salmon	Abdominal cramps, small bowel obstruction, slow intestinal transit with area of edematous gastric folds, and transition from a slightly dilated jejunum to a normal ileum	No data	Intestinal	X-ray	30
5	1993	Peru	Men without age report	No data	Oropharyngeal pain	Pseudoterranova decipiens	Oropharynx	Observation of the larva in the oral cavity	25
6	1995	Paraguay- Chile	Man without age report	Ceviche	Cough	Anisakis sp.	Oropharynx	Identification of expelled larvae	16
7	1997	Chile	45-year- old man	Smoked fish	Acute epigastric pain and empty stomach feeling for three days	<i>P. decipiens</i> L4 larva	Gastric	Gastric endoscopy	32
8	1997	Peru	42-year- old woman	No data	No data	Anisakis simplex	Oropharynx	Observation of larvae	14
9	1997	Peru	22-year- old man	Mahi-mahi ceviche (Coryphaena hippurus)	Abdominal pain that decreased progressively and disappeared spontaneously after two days	Anisakis physeteris	Gastrointestinal	Epidemiological history	13
10	1997	Chile	10-year- old child	No data	Cough	<i>P. decipiens</i> L4 larva	Oropharynx	Morphological identification of the larva	15
11	1997	Chile	51-year- old woman	No data	Cough	<i>P. decipiens</i> L4 larva	Oropharynx	Morphological identification of the larva	15
12	1998	Chile	30-year- old woman	No data	Cough	<i>P. decipiens</i> L4 Iarva	Oropharynx	Morphological identification of the larva	15
13	1998	Chile	22-year- old woman	Chilean common hake slightly fried	Nausea, nasal pruritus, productive cough, and pharyngeal pain that persisted for one week after larva expulsion and was associated with a local allergic reaction	<i>P. decipiens</i> L4 Iarva	Gastroallergic	Morphological identification of the larva	15
14	1998	Peru	36-year- old man	Mahi-mahi ceviche ( <i>C.</i> <i>hippurus)</i>	Abdominal discomfort eight hours after eating the ceviche that progressed to epigastric pain of increasing intensity within the first 24 hours. Symptoms ceased spontaneously 72 hours later.	A. physeteris	Gastric	Epidemiological history	13
15	1998	Peru	38-year- old woman	No data	Retrosternal burning sensation and sporadic colicky pain in right iliac fossa for one week	<i>Anisakis</i> spp.	Gastric	Direct observation of the larvae extracted by endoscopy.	26

#### Table 2. Clinical cases associated with anisakid nematodes in South American countries. (continued)

ιαυι	ez. Ci		sassociated		nematodes in South Ar		es. (continueu)		
No	Year	Country	Patient	Food previously ingested	Symptoms	Identified anisakid	Symptoms	Diagnostic method	Ref
16	1999	Chile	55-year- old woman	Red cusk-eel ( <i>Genypterus</i> <i>chilensis)</i> sushi	Cough	<i>P. decipiens</i> L4 Iarva	Oropharynx	Morphological identification of the larva	15
17	1999	Chile	37-year- old man	Ceviche	Nausea and pain in the pharynx	<i>P. decipiens</i> L4 larva	Oropharynx	Morphological identification of the larva	15
18	1999	Chile	26-year- old woman	No data	Productive cough	<i>P. decipiens</i> L4 larva	Oropharynx	Morphological identification of the larva	15
19	2000	Chile	38-year- old man	Mariscal (seafood and raw fish)	Persistent and disabling pain in the epigastrium that evolved into regurgitation of gastric contents (where the larva was found) and decreased in intensity over time until it disappeared after 15 days. Endoscopy of the upper gastrointestinal tract showed mild gastritis without ulcers or presence of other larvae.	<i>Anisakis</i> sp.	Gastric	Observation of the larva in the oral cavity	31
20	2001		40-year- old man	Pomfret ( <i>Brama</i> <i>australis</i> ceviche	Foreign body sensation in the teeth (the patient found a larva when she was washing her teeth)	Anisakid nematode	Oropharynx	Identification of larvae after clarification process with lactophenol.	16
21	2001	Peru	42-year- old woman	No data	No data	P. decipiens L4	Oropharynx	Observation of larvae extracted from the oral cavity, initially classified as <i>Toxocara</i> sp.	14
22	2002	Peru	17-year- old woman	Chilean jack mackerel ( <i>Trachurus</i> <i>murphyi</i> ), "bonito" ( <i>Sarda</i> <i>chiliensis</i> ) and tuna ( <i>Thunnus</i> sp.) ceviche	Nausea and epigastric pain four hours after ingestion and nausea of greater intensity after an hour with sore throat and vomiting of food contents	P. decipiens	Gastric	Processing of the larva expelled in vomit	24
23	2002	Chile	54-year- old woman	Ceviche	Coughing and tingling in the pharynx	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
24	2002	Chile	19-year- old woman	Hake ceviche	No data	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
25	2002	Chile	43-year- old woman	Fried fish	Tingling in the pharynx	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
26	2002	Chile	50-year- old man	Hake ceviche	Coughing and tingling in the pharynx	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
27	2002	Chile	33-year- old woman	Hake ceviche	No data	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
28	2003	Chile	15-year- old child	Chilean jack mackerel ceviche	Severe cough	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
29	2003	Chile	26-year- old woman	Chilean common hake slightly fried	Heartburn and vomiting with larva presence	<i>Pseudoterranova</i> sp.	Gastric	Morphological identification of the larva	16

# Table 2. Clinical cases associated with anisakid nematodes in South American countries. (continued)

No	Year	Country	Patient	Food previously ingested	Symptoms	Identified anisakid	Symptoms	Diagnostic method	Ref
30	2003	Chile	47-year- old woman	Fried pomfret ( <i>B australis)</i>	Sensation of asphyxia and increased production of oropharyngeal secretions after larval removal.	<i>Pseudoterranova</i> sp.	Gastroallergic	Morphological identification of the larva	16
31	2003	Chile	6-year-old child	Hake ceviche	Tingling in the pharynx with removal of two larvae through the oral cavity	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
32	2004	Chile	24-year- old woman	Hake ceviche	Severe cough with subsequent elimination of larvae through the oral cavity.	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
33	2004	Chile	Man without age report	Hake ceviche	No data	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
34	2004	Chile	Woman without age report	Hake ceviche	No data	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
35	2005	Chile	1-year-old girl	No data	Severe diarrhea, loss of appetite, and elimination of larva through the anus	<i>Pseudoterranova</i> sp.	Intestinal	Morphological identification of the larva	16
36	2005	Chile	Man without age report	Corvina ( <i>Cilus gilberti)</i> ceviche	Foreign body sensation in the mouth, tingling in the larynx and severe cough before removing the larva (the patient found the larva when brushing his teeth).	<i>Pseudoterranova</i> sp.	Oropharynx	Morphological identification of the larva	16
37	2006	Chile	60-year- old woman	Ceviche	Heartburn, malaise, persistent nausea, abdominal distension, and flatulence for five days	<i>Pseudoterranova</i> sp.	Gastrointestinal	Morphological identification of the larva	18
38	2007	Chile	30-year- old woman	Raw salmon sushi	Coughing and sneezing	P. decipiens	Oropharynx	Morphological identification of the larva	3
39	2010	Brazil	73-year- old man	Raw seafood	Epigastric pain, sensation of fullness and early satiety. Improvement of symptoms after endoscopic removal of the larva. The patient died at 20 days from unknown causes	Family Anisakidae	Gastrointestinal	Direct observation of the larvae extracted by endoscopy.	27
40	2011	Argentina	9-year-old girl	No data	Nonspecific gastrointestinal symptoms for 9 days	<i>Anisakis - Contraecum</i> complex	Gastrointestinal	Morphological identification of the larva through photographs	19
41					Three patients				17
42	2012-	Chile	Two women and two men	Ceviche	regurgitated the larva after eating ceviche. One of them had a tingling sensation in the pharynx, nausea and cough before	Pseudoterranova	Oropharyny	Morphological and molecular identification (DNA extraction	17
43	2014	CITIC	between the ages of 22 and 59		the larva was expelled; two patients had no symptoms, and there is no data on symptoms of	cattani	Oropharynx	with DNeasy Blood and Tissue Kit) of the expelled larvae	17
44					the other patient.				17
45	2018	Colombia	52-year- old woman	Fish-based raw food	Severe epigastric pain accompanied by nausea, loss of appetite, vomiting, diarrhea and urticaria for two days	A. simplex	Gastrointestinal	Morphological identification of larvae after processing with glutaraldehyde	28

Source: Own elaboration.

# Intermediate fish hosts of anisakids reported in South America

Anisakid nematodes are parasites present in marine mammals and fish species for human consumption. For

example, in South America, fish parasitized by these species have been identified in Argentina, Brazil, Chile, Colombia, Ecuador, Peru and Venezuela. Table 3 lists the fish species identified as hosts of parasites of the family *Anisakidae* in the region.

Scientific nameCommon nameCommon nameContract originStandlor originStandlor (%)RefAcanthisius patachonicusNDArgentinaPseudoterranova catani16257Ageneiosus caucanus"Doncella"ColombiaContracecum sp.4542.221Aluterus monoceros"Pez puerco"BrazilAnisakis simplex80100123Aluterus monoceros"Pez puerco"BrazilAnisakis simplex81301023Aphos porosusNDChileContracecum sp.100166333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333333	Host		Country of		Consulad	Prevalence	
Acathisius patachonicus     ND     Argenting     Feeddaterranova cattanin     45     57.8     4       Ageneiosus caucanus     "Doncella"     Colombia     Contraccum sp.     460     23.3     2       Aluterus monoceros     "Pez puerco"     Brazil     Anisakis sp.     100     16     2       Aphos porosus     "Per puerco"     Brazil     Anisakis simplex     8     13     9       Austromenidia laticlavia     ND     Chile     Contracecum sp.     300     00     8       Austromenidia laticlavia     ND     Chile     Contracecum sp.     300     100     9       Austromenidia laticlavia     ND     Celadot:     Anisakis shyseteris     6     50     3       Austromenidia laticlavia     ND     Argentina     Colorabia     Contracecum sp.     ND     ND     9       Austromenidia laticlavia     Bullet tuna     Ecadot:     Anisakis physeteris     600     3.3     9       Aust stazard     "Migara     Colombia     Contracecum sp.     600     6.7     3     9<	Scientific name	Common name	Country of origin	Genus and/or species	Sampled fish (n)		Ref
Acanthisus patachonicus ND Argentina Fermion (Contracecum sp.) (45) (57.8) (41)   Ageneiosus caucanus "Doncella" Colombia Contracecum sp. (60) 2.2 32   Aluterus monoceros "Pez pueco" Brazil Anisakis sp. 100 1 23   Aphos porosus "Per pueco" Brazil Anisakis simplex 88 133 92   Arapalma gigas "Pirarucú" Brazil Hysterothylacium sp. 100 98 30   Austromenidal abticlavian ND Chile Contracecum sp. 100 98 30   Austromenidal abticlavian ND Chile Contracecum sp. 100 98 30   Austrochei Bullet tuna Ecuador. Anisakis hysteris 60 ND 30   Auxis thazard "figate tuna Brazil Contracecum sp. 600 3.3 30   Caquetala kraussii "Mojarra amarilla" Colombia Contracecum sp. 600 3.3 30   Caranx Hipos Chilea jack Golombia Family Anisakis physeteris ND ND 30 30   Caranx Lipos Chilea jack Golombia Family Anisakis physeteris 160 <td< td=""><td></td><td></td><td></td><td>Pseudoterranova cattani</td><td>16</td><td>25</td><td>37</td></td<>				Pseudoterranova cattani	16	25	37
Ageneiosus caucausus     "Doncella"     Colombia     Contracecum sp.     60     22.3       Aluterus monoceros     "Pez puerco"     Brazil     Anisakis sp.     100     1     23       Aluterus monoceros     "Pez puerco"     Brazil     Anisakis sp.     100     16     33       Aphos porosus     ND     Chile     Anisakis simplex     88     133     28       Arapalma gigas     "Pirarucú"     Brazil     Hysterothylacium sp.     100     98     20       Austromenidia laticlavian     ND     Chile     Contracecum sp.     300     100     20       Austrochei     Bullet tuna     Ecuador.     Anisakis hysteeris     60     ND     20       Auxis thazard     "indjara"     Colombia     Contracecum sp.     600     3.3     3       Caquetaia kraussii     "Mojarra"     Colombia     Contracecum sp.     600     3.3     3       Caranx Hipos     Crevalle jack     Brazil     Contracecum sp.     600     6.7     3       Caranx Lipos     ND     Bra	Acanthisius patachonicus	ND	Argentina	r seuuolen anova callani	45	57.8	21
Additional contract of a strain strain strain strain strain strain strain				Pseudoterranova sp.	45	42.2	21
Aluterus monoceros     "Pez puerco"     Brazil     Industry for accum sp.     100     16     3       Aphos porosus     ND     Chile     Anisakis simplex     8     13     2       Arapaima gigas     "Piraucú"     Brazil     Hysterothylacium sp.     100     98     7       Austromenidia laticiavian     ND     Chile     Contracecum sp.     ND     100     34       Ausis rochei     Bullet tuna     Ecuador.     Anisakis physeteris     6     50     3       Auxis torochi     Bullet tuna     Ecuador.     Anisakis physeteris     ND     ND     3       Auxis thazard     Prigate tuna     Brazil     Contracecum sp.     60     9.5     2       Caquetaia kraussii     "Mojarra amarilla"     Colombia     Contracecum sp.     60     3.3     3       Caranx Hipos     Crevalle jack     Brazil     Colombia     Family Anisakidae     12     8.5     3.8     3       Caranx Lipos     Colombia     Family Anisaki sp.     55     18.2     3     3     3 <td>Ageneiosus caucanus</td> <td>"Doncella"</td> <td>Colombia</td> <td>Contracecum sp.</td> <td>60</td> <td>23.3</td> <td>22</td>	Ageneiosus caucanus	"Doncella"	Colombia	Contracecum sp.	60	23.3	22
Aphos porosus     ND     Chile     Anisakis simplex     8     110     16     29       Arpaima gigas     "Pirarucú"     Brazil     Hysterothylacium sp.     100     98     20       Austromenidia laticlavian     ND     Chile     Contracecum sp.     300     100     20       Austromenidia laticlavian     ND     Argentina     Contracecum sp.     ND     100     20       Auxis rochei     Bullet tuna     Ecuador.     Anisakis physeteris     ND     ND     20       Auxis thazard     "Migiara amarilia"     Colombia     Contracecum sp.     ND     ND     20       Caquetaia kraussii     "Migiara amarilia"     Colombia     Contracecum sp.     60     95     22       Caranx Hipos     Chilean jack     Rolo     Fazil     Contracecum sp.     60     6.7     30     32     32       Caranx latus     Modereralova desceric     Fazil     Anisakis thylacium sp.     55     1.8.2     32     32     32     32     32     32     32     32     32<	Aluterus monoceros	"Pez puerco"	Brazil	Anisakis sp.	100	1	23
Aphos porosus ND Chile Principal defermancy and definitions 10 10 10   Arapaima gigas "Pirarucú" Brazil Hysterothylacium sp. 100 98 73   Austromenidia laticlavian ND Chile Contracecum sp. 300 100 98   Auxis rochei Bullet tuna Ecuador. Anisakis physeteris 6 500 30   Auxis thazard Frigate tuna Brazil Aniskis physeteris 600 900 30   Caquetaia kraussii "Mojarra amarila" Colombia Contracecum sp. 600 93.3 30   Caquetaia kraussii "Mojarra amarila" Colombia Contracecum sp. 600 3.3 30   Caranx Hipos Chilean jack mackerel Brazil Contracecum sp. 600 6.7 30   Caranx Latus Chilean jack mackerel Brazil Pseudoterranova sp. 600 6.7 30   Caranx Latus ND Brazil Famila Aniskis sp. 55 18.2 30   Carcharhinus brachyurus ND Brazil Contracecum sp. 55 32.2 30   Carcharhinus signatus ND Brazil Contracecum sp. 55 32.2 30 </td <td>Alaterus monoceros</td> <td>rez puerco</td> <td>DIGZII</td> <td>Contracecum sp.</td> <td>100</td> <td>16</td> <td>23</td>	Alaterus monoceros	rez puerco	DIGZII	Contracecum sp.	100	16	23
Arapaima gigas"Pirarucú"BrazilHysterothylacium sp.100987573Austromenidia laticlavianNDChileContracecum sp.301009873Austromenidia laticlavianNDArgentinaContracecum sp.3010034Auxis rocheiBullet tunaEcuador.Anisakis physteris6650039Auxis thazardFrigate tunaBrazilAnisakis physterisNDND10029Caquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.60095022Caranx HiposCrevalle jackBrazilContracecum sp.6003.399Caranx HiposCrevalle jackBrazilContracecum sp.6006.739Caranx hiposCrevalle jackBrazilPseudoterranova sp.6006.739Caranx hiposNDBrazilPseudoterranova sp.6006.739Caranx huposNDBrazilContracecum sp.6006.739Caranx huposNDBrazilContracecum sp.551.839Caranx huposNDBrazilContracecum sp.6006.739Caranx huposNDBrazilContracecum sp.553.639Caranx huposNDBrazilContracecum sp.553.639Caranx huposNDBrazilContracecum sp.556.040Carcharhinus signatus	Anhos norosus	ND	Chile	Anisakis simplex	8	13	52
Austromenidia laticilaria Austromenidia laticilaria NDNDChile Contracecum sp.3010030Austromenidia laticilaria Auxis rocheiNDArgentina Ecuador.Contracecum sp.NDND30Auxis rocheiBullet tunaEcuador.Anisakis physeterisNDND30Auxis thazardFrigate tuna amarilia"BrazilAnisakis physeterisNDND30Caquetaia kraussii"Mojarra amarilia"ColombiaContracecum sp.60952Caranx HiposCrevalle jack mackereiBrazilContracecum sp.603.3330Caranx HiposChilean jack mackereiColombiaFamily Anisakidae128.53303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030303030	Aprios por osus	ND	Cline	Pseudoterranova decipiens	8	75	52
Astyanax eigenmanniorumNDArgentinaContracecum sp.ND10014Auxis rocheiBullet tunaEcuador.Anisakis physeteris65030Auxis thazard $Frigate tuna$ $Brazil$ $Anisakis physeteris$ NDND1020Caquetaia kraussii $"monilla"$ Colombia $Contracecum sp.$ 60952Caranx HiposCrevalle jackBrazilContracecum sp.603.3.320Caranx HiposCrevalle jackBrazilContracecum sp.606.7.73Caranx HiposColombiaFamily Anisakidae128.5.51.83Caranx HiposNDBrazilSeudoterranova sp.606.7.733Caranx hiposNDBrazilSeudoterranova sp.551.8.233Caranx hiposNDBrazilContracecum sp.553.6.133 <tr< td=""><td>Arapaima gigas</td><td>"Pirarucú"</td><td>Brazil</td><td>Hysterothylacium sp.</td><td>100</td><td>98</td><td>73</td></tr<>	Arapaima gigas	"Pirarucú"	Brazil	Hysterothylacium sp.	100	98	73
Advainax elignmentationHoFrigenentationFrigenentationContracecum sp.RoRoSoAuxis rocheiBullet tunaEcuador.Anisakis physeterisNDNDRoAuxis thazardFrigate tunaBrazilAnisakis physeterisNDNDRoRoCaquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.60952Caquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.603.39Caranx HiposChilea jack mackerelBrazilContracecum sp.606.730Caranx HiposChilea jack mackerelBrazilPseudoterranova sp.606.730Caranx IatusChilea jack mackerelBrazilPseudoterranova sp.606.730Caranx latusNDBrazilPseudoterranova sp.551.8.220Carcharhinus brachyurusNDBrazilContracecum sp.553.2.730Carcharhinus signatusNDBrazilContracecum sp.553.2.730Cauque mauleanum"Cauque del maule"Anisakis sp.556090Contracecum sp.556090203030Cauque mauleanumGauge del maule"Anisakis sphyseteris1478.9Centropomus undecimalisNDColombiaAnisakis physeteris124.3.3Contracecum sp.ColombiaGaniskis physeteris124.3	Austromenidia laticlavian	ND	Chile	Contracecum sp.	30	10	52
AussidenceEducationEducationAnisakis physeterisOSOSOAuxis thazardFrigate tuna amarilla"BrazilAnisakis physeterisNDND20Caquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.609522Caquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.603.330Caranx HiposCrevalle jackBrazilContracecum sp.606.730Caranx HiposColombiaFamily Anisakidae128.5536Caranx latusMorse-eye jackBrazilPseudoterranova sp.606.730Parant latusNDBrazilContracecum sp.551.8.230Carcharhinus brachyurusNDBrazilContracecum sp.553.6.630Carcharhinus signatusNDBrazilContracecum sp.553.2.730Cauque mauleanum"Cauque del maule"ColombiaContracecum sp.553.6.730Carcharhinus signatusNDBrazilContracecum sp.553.6.73030Cauque mauleanumCauque del maule"ColombiaAnisakis spinplex92252Contraceum sp.551.6.1Anisakis spinplex92252Contraceum sp.553.07303030Cauque mauleanumCauque del maule"Anisakis physeteris124.330 <t< td=""><td>Astyanax eigenmanniorum</td><td>ND</td><td>Argentina</td><td>Contracecum sp.</td><td>ND</td><td>100</td><td>34</td></t<>	Astyanax eigenmanniorum	ND	Argentina	Contracecum sp.	ND	100	34
Auxis thazardFrigate tunaBrazilAnisakis typicationsNotNotNotCaquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.609522Caquetaia kraussii"Mojarra amarilla"ColombiaContracecum sp.603.330Caranx HiposCrevalle jackBrazilContracecum sp.603.330Caranx HiposChilean jack mackerelColombiaFamily Anisakidae128.553.6Caranx HiposChilean jack mackerelBrazilPseudoterranova sp.606.730Caranx IatusHorse-eye jack Pseudoterranova sp.606.7303030Carcharhinus brachyurusNDBrazilContracecum sp.551.8.230Carcharhinus signatusNDBrazilContracecum sp.553.6.73030Cauque mauleanum maule""Gauque del maule"Anisakis sp.5530.73030Carthopomus armatusNDColombiaAnisakis shyseteris124230Cortropomus armatusNDColombiaFamily Anisakidae234.330Cortyphaena hippurusMoColombiaAnisakis physeteris124230Cortyphaena hippurusNDColombiaAnisakis physeteris124.330Cottoperca gobioNDAnisakis physeteris12712Cottoperca gobioNDArgentiaAni	Auxis rochei	Bullet tuna	Ecuador.	Anisakis physeteris	6	50	38
Anisakis typicalNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDNDND<	Auvis thazard	Frigate tuna	Brazil	Anisakis physeteris	ND	ND	20
Caduetala kraussinamarilla"ColombiaContracecum sp.60959595Caranx HiposCrevalle jackBrazilContracecum sp.603.339Caranx HiposChilean jack mackerelColombiaFamily Anisakidae128.536Crevalle jackBrazilPseudoterranova sp.606.739Caranx latusMasserepe jackBrazilSeudoterranova sp.606.739Carcharhinus brachyurusNDBrazilContracecum sp.551.8.239Carcharhinus signatusNDBrazilContracecum sp.553.2.739Cauque mauleanum"Cauque del maule"ChileAnisakis sp.556040Cantropomus armatusNDScalueAnisakis simplex92232Cartoropomus undecimalisNDColombiaAnisakis physeteris11478.930Cortopomus armatusArmed snookColombiaAnisakis physeteris124230Cortopomus undecimalisNDColombiaFamily Anisakidae234.330Cortopomus undecimalisNDColombiaFamily Anisakis physeteris124230Cortopomus undecimalisNDColombiaFamily Anisakis physeteris124230Cortopomus undecimalisNDColombiaFamily Anisakis physeteris124330Cortopomus undecimalisNDColombiaAnisakis physeteris<	Auxis ulazalu		DI dZII	Anisakis typica	ND	ND	20
Caranx HiposChilean jack mackerelColombiaFamily Anisakidae128.53.6Crevalle jackBrazilPseudoterranova sp.6006.73Caranx latusAnsakis sp.551.83Parama latusHorse-eye jackContracecum sp.553.63Carcharhinus brachyurusNDBrazilContracecum sp.553.6.73Carcharhinus signatusNDBrazilContracecum sp.553.6.73Cauque mauleanum"Cauque del male"ColombiaAnisakis sp.56004Centropomus armatusArmed snoskColombiaAnisakis physeteris12423ContrapeunaMonoColombiaFamily Anisakidae124.13Contracecum sp.MonoGolombiaAnisakis physeteris124.23Contropomus armatusNDColombiaFamily Anisakidae2.34.33Corphaena hippurusNDColombiaFamily Anisakidae2.34.33Corphaena hippurusNDColombiaFamily Anisakidae2.34.33Cotoperca gobioNDArgentinaPeru1243Cotoperca gobioNDArgentinaPseudoterranova sp.812.533Cotoperca gobioNDArgentinaPseudoterranova sp.5343Cotoperca gobioNDArgentinaArisakis sp.53	Caquetaia kraussii		Colombia	Contracecum sp.	60	95	22
Caranx hiposmackerelColombiaFamily Anisakidae128.53.6Crevalle jackBrazilPseudoterranova sp.606.739Caranx latusAnisakis sp.551.839Caranx latusHorse-eye jackBrazilContracecum sp.551.8.239Horse-eye jackBrazilContracecum sp.553.639Pseudoterranova sp.553.63939Carcharhinus brachyurusNDBrazilContracecum sp.72.5Carcharhinus signatusNDBrazilContracecum sp.72.5Cauque mauleanum"Cauque del maule"Anisakis sp.56040Cauque mauleanumCauque del maule"Anisakis simplex92252Contropomus armatusArmed snookColombiaAnisakis physeteris1124238Cortyphaena hippurusNDColombiaFamily Anisakidae234.336Coryphaena hippurusMDColombiaFamily Anisakidae234.336Coryphaena hippurusNDPeruAnisakis physeteris1258.3314Cotoperca gobioNDArgentinaPseudoterranova sp.812.537.7Cotoperca gobioNDArgentinaPseudoterranova sp.12413Cotoperca gobioNDArgentinaPseudoterranova sp.12413Cotoperca gobioNDArgentina <t< td=""><td></td><td>Crevalle jack</td><td>Brazil</td><td>Contracecum sp.</td><td>60</td><td>3.3</td><td>39</td></t<>		Crevalle jack	Brazil	Contracecum sp.	60	3.3	39
Anisakis sp.551.89Caranx latusHorse-eye jackFrazilContracecum sp.5518.29Horse-eye jackFrazilContracecum sp.553.69Kacharhinus brachyurusNDBrazilContracecum sp.72.59Carcharhinus signatusNDBrazilAnisakis sp.56009Cauque mauleanumMaule"ChileAnisakis simplex9225Centropomus armatusArmed snookColombiaAnisakis physeteris11478.99Coryphaena hippurusNDColombiaFamily Anisakidae234.39Coryphaena hippurusNDScuadorAnisakis physeteris124.39Cotoperca gobioNDArgentinaAregentina121213Cotoperca gobioNDArgentinaSeudoterranova sp.12413Cotoperca gobioNDArgentinaSeudoterranova sp.12413Cotoperca gobioNDArgentinaSeudoterranova sp.12413Cotoperca gobioNDArgentinaSeudoterranova sp.12413Cotoperca gobioNDArgentinaSeudoterranova sp.1312.530.7613Cotoperca gobioNDBrazilContracecum sp.124.31313Cotoperca gobioNDArgentinaSeudoterranova sp.1312.513.614 <td>Caranx Hipos</td> <td>5</td> <td>Colombia</td> <td>Family Anisakidae</td> <td>12</td> <td>8.5</td> <td>36</td>	Caranx Hipos	5	Colombia	Family Anisakidae	12	8.5	36
Caranx latus     Horse-eye jack     Brazil     Contracecum sp.     55     1.8     39       Caranx latus     Horse-eye jack     Brazil     Contracecum sp.     55     3.6     39       Carcharhinus brachyurus     ND     Brazil     Contracecum sp.     55     32.7     39       Carcharhinus brachyurus     ND     Brazil     Contracecum sp.     7     2.5     40       Carcharhinus signatus     ND     Brazil     Contracecum sp.     55     60     40       Carcharhinus signatus     ND     Brazil     Contracecum sp.     55     40     40       Carcharhinus signatus     ND     Brazil     Anisakis sp.     5     40     40       Carcharhinus signatus     ND     Colombia     Anisakis sp.     5     40     40       Cauque mauleanum     "Cauque del maule"     Chile     Anisakis physeteris     12     42     38       Centropomus undecimalis     ND     Colombia     Family Anisakidae     23     4.33     36       Coryphaena hippurus     Morad		Crevalle jack	Brazil	Pseudoterranova sp.	60	6.7	39
Caranx latusHorse-eye jack $-$ BrazilBrazil $-$ Contracecum sp.553.639Carcharhinus brachyurusNDBrazilContracecum sp.72.532.739Carcharhinus brachyurusNDBrazilContracecum sp.72.54040Carcharhinus signatusNDBrazilContracecum sp.56040Cauque mauleanum"Cauque del maule"Anisakis simplex92252Centropomus armatusArmed snookColombiaAnisakis simplex92252Centropomus undecimalisNDColombiaAnisakis physeteris1124238Corphaena hippurusCommon 			Brazil	Anisakis sp.	55	1.8	39
Hysterothylacium sp.553.639Carcharhinus brachyurusNDBrazilContracecum sp.5532.799Carcharhinus signatusNDBrazilContracecum sp.72.540Carcharhinus signatusNDBrazilAnisakis sp.56040Cauque mauleanum"Cauque del maule"ChileAnisakis simplex92252Centropomus armatusArmed snookColombiaAnisakis physeteris1478.976Centropomus undecimalisNDColombiaFamily Anisakidae234.336Coryphaena hippurusCommon olophinfishEcuadorAnisakis physeteris124238Cottoperca gobioNDArgentinPeru1258.3314Cottoperca gobioNDArgentinPseudoterranova sp.12439Cottoperca popioNDArgentinPseudoterranova sp.530.7631Cottoperca popioNDArgentinPseudoterranova sp.530.7631Cottoperca popioNDArgentinPseudoterranova sp.3230.7631Cottoperca popioNDArgentinPseudoterranova sp.3230.7631Cottoperca popioNDArgentinPseudoterranova sp.3230.7631Cottoperca popioNDArgentinPseudoterranova sp.5230.7631Cottoperca popioNDArgentinCont	Carapylatus	Horse overjack		Contracecum sp.	55	18.2	39
Carcharhinus brachyurusNDBrazilContracecum sp.500500600400Carcharhinus signatusNDBrazilAnisakis sp.5600400Carcharhinus signatusNDBrazilAnisakis sp.5400400Cauque mauleanum"Cauque del maule"ChileAnisakis simplex92.252Hysterothylacium geschei11478.974Centropomus armatusArmed snookColombiaAnisakis physeteris1124238Centropomus undecimalisNDColombiaFamily Anisakidae2.34.336Coryphaena hippurusCommon dolphinfishEcuadorAnisakis physeteris1258.3314PeruPeruHysterothylacium sp.1258.3314Cottoperca gobioNDArgentinPseudoterranova sp.812.537Cottoperca gobioNDArgentinContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.530.7641Cottoperca gobioNDBrazilContracecum sp.52	Caranx latus	Horse-eye Jack		Hysterothylacium sp.	55	3.6	39
Carcharhinus signatusNDBrazilAnisakis sp. Contracecum sp.56040Cauque mauleanum"Cauque del maule"Anisaki simplex92252Cantropomus armatusArmed snookColombiaAnisakis simplex1478.974Centropomus armatusArmed snookColombiaAnisakis physeteris124238Centropomus undecimalisNDColombiaFamily Anisakidae234.336Corpphaena hippurusCommon olphinfishEcuadorAnisakis physeteris1258.3331Corpphaena hippurusPeruPeruInsakis physeteris1258.3331Cottoperca gobioNDArgentinaPseudoterranova sp.812.530.7631Cotnos pp.NDBrazilContracecum sp.5267.341				Pseudoterranova sp.	55	32.7	39
Carcharhinus signatusNDBrazilHindukts sp.Is of the constraint of th	Carcharhinus brachyurus	ND	Brazil	Contracecum sp.	7	2.5	40
Cauque mauleanum"Cauque del maule"ChileContracecum sp.54040Cauque mauleanum"Cauque del maule" $Anisaki s simplex$ 92252Centropomus armatusArmed snookColombiaAnisakis physeteris1478.974Centropomus undecimalisNDColombiaAnisakis physeteris124238Centropomus undecimalisNDColombiaFamily Anisakidae234.336Coryphaena hippurusCommon colphinfishEcuadorAnisakis physeteris1258.3314PeruPeruInisakis physeteris1258.331413Cottoperca gobioNDArgentinaPseudoterranova sp.812.530.7641Cynoscion spp.NDBrazilContracecum sp.5267.341	Carcharbinus signatus	ND	Brazil	Anisakis sp.	5	60	40
Cauque mauleanum maule"Cauque del maule"Amsakri simplexIIICauque mauleanum maule"Armed snookColombiaAnisakis simplex1478.974Centropomus armatusArmed snookColombiaAnisakis physeteris124238Centropomus undecimalisNDColombiaFamily Anisakidae234.336Coryphaena hippurusCommon olphinfishEcuadorAnisakis physeteris693038Coryphaena hippurusPeruPeruHysterothylacium sp.1258.3314Cottoperca gobioNDArgentinaPseudoterranova sp.812.530.76Coryposcion spp.NDBrazilContracecum sp.5267.341	Carchanninus Signatus	ND	DI dZII	Contracecum sp.	5	40	40
InduceInduceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstanceInstance <td>Cauque mauleanum</td> <td></td> <td>Chile</td> <td>Anisakis simplex</td> <td>9</td> <td>22</td> <td>52</td>	Cauque mauleanum		Chile	Anisakis simplex	9	22	52
Centropomus undecimalisNDColombiaFamily Anisakidae234.336Common dolphinfishEcuador $A_{AB}$ Anisakis physeteris693038Coryphaena hippurus $Peru$ $Anisakis physeteris$ 1258.3314 $Peru$ $Peru$ $Hysterothylacium sp.$ 12713Cottoperca gobioNDArgentina $Pseudoterranova sp.$ 812.537Cynoscion spp.NDBrazil $Contracecum sp.$ 5267.341	Cauque mauleanum	maule"	Cline	Hysterothylacium geschei	14	78.9	74
$ \begin{array}{c} \mbox{Correl} \mbox{Constraints} & \mbox{MD} & \mbox{Constraints} & Const$	Centropomus armatus	Armed snook	Colombia	Anisakis physeteris	12	42	38
$ \begin{array}{c} \mbox{dolphinfish} & \mbox{Ecuador} & \mbox{Anisakis physeteris} & \mbox{for anisotropy base} & \mbox{Anisakis physeteris} & \mbox{for anisotropy base} &$	Centropomus undecimalis	ND	Colombia	Family Anisakidae	23	4.3	36
Coryphaena hippurus     "Dorado"     Peru     Inc     Inc <td></td> <td></td> <td>Ecuador</td> <td></td> <td>69</td> <td>30</td> <td>38</td>			Ecuador		69	30	38
ModelHysterothylacium sp.12413Cottoperca gobioNDArgentinaPseudoterranova sp.812.537Cynoscion spp.NDFazilContracecum sp.5230.7641	Coryphaena hippurus			Anisakis physeteris	12	58.33	14
Cottoperca gobio     ND     Argentina     Pseudoterranova sp.     8     12.5     37       Anisakis sp.     52     30.76     41       Cynoscion spp.     ND     Brazil     Contracecum sp.     52     67.3     41		"Dorado"	Peru		12	7	13
Anisakis sp.     52     30.76     41       Cynoscion spp.     ND     Brazil     Contracecum sp.     52     67.3     41				Hysterothylacium sp.	12	4	13
Cynoscion spp.NDBrazilContracecum sp.5267.341	Cottoperca gobio	ND	Argentina	Pseudoterranova sp.	8	12.5	37
				Anisakis sp.	52	30.76	41
Pseudoterranova sp. 52 3.84 <sup>41</sup>	Cynoscion spp.	ND	Brazil	Contracecum sp.	52	67.3	41
				Pseudoterranova sp.	52	3.84	41

Host		Country of		Sampled	Prevalence of	
Scientific name	Common name	origin	Genus and/or species	fish (n)	infection (%)	Ref
Dipturus trachyderma	ND	Brazil	Anisakis sp.	8	25	40
, ,			Contracecum sp.	8	62.5	40
		Argentina Uruguay	Anisakis simplex	2 086	5.85	42
Engraulis anchoíta	"Anchoíta"	Argentina Uruguay	Contracecum sp.	2 086	39.7	42
		Argentina Uruguay	Pseudoterranova sp.	2 086	0.34	42
Engraulis ringens	Peruvian anchoveta	Chile	Anisakis sp.	598	3	43
Eugerres plumieri	"Mojarra"	Venezuela	Contracaecum spp.	90	97	44
Eugenes plannen	riojuria	Venezuela	Pseudoterranova spp.	90	3	44
<i>Gadus</i> sp.	Cod	Brazil	Pseudoterranova azarasi	ND	ND	45
Gadus macrocephalus	Zarbo cod	Brazil	Family Anisakidae	11	64	46
			Anisakis sp.	37	8.1	40
Galoeorhinus vitaminicus	ND	Brazil	Contracecum sp.	37	5.4	40
			Pseudoterranova sp.	37	5.4	40
	Pink cusk-eel	Argentina	Pseudoterranova sp.	44	2.3	37
Convertorius bloos doo		Chile	Anisakis sp. (Type I)	81	7.4	47
Genypterus blacodes			Anisakis sp. (Type II)	81	4.9	47
			Pseudoterranova spp.	81	53.1	47
			Anisakis typica	60	1.7	48
Genypterus brasiliensis	ND	Brazil	<i>Hysterothylacium deardorffoverstreetorum</i> (s.l)	60	5	48
Genypterus chilensis	ND	Chile	Pseudoterranova decipiens	18	50	47
			Anisakis sp.	7	14.3	40
Heptranchias perlo	ND	Brazil	Contracecum sp.	7	28.6	40
			Anisakis sp.	1	100	40
Hexanchus griseus	ND	Brazil	Contracecum sp.	1	100	40
Hexanematichthys sp.	"Bagre pintado"	Ecuador	Contracecum sp.	64	12.5	35
		Argentina	Contracecum sp.	10	70	34
		5		227	100	36
Hoplias malabaricus	Wolf fish	Colombia	Contracecum sp.	60	93.3	22
			Contracaecum spp.	45	100	49
Hoplias microlepis	"Trahira"	Ecuador	Contracecum sp.	64	25	35
Katsuwonus pelamis	Skip jack tuna	Ecuador	Anisakis physeteris	100	31	38
Larimus argenteus	Silver drum	Colombia	Anisakis physeteris	2	100	38
	Blackfin	30.0.11010	Anisakis simplex	87	1.14	50
Lophius gastrophysus	goosefish	Brazil	<i>Hysterothylacium</i> sp.	87	12.6	50
			Anisakis physeteris	3	33	52
			Anisakis simplex	3	100	52
			A MOURIS SIMPLEX	4	100	47
Macruronus magellanicus	ND	Chile	Hysterothylacium sp.	3	33	52
					25	47
			Pseudoterranova decipiens	4		
			,	3	33	52

Host					Prevalence	
Scientific name	Common name	Country of origin	Genus and/or species	Sampled fish (n)	of infection (%)	Ref
			Anisakis sp. (Type I)	104	8.7	47
Merluccius australis	ND	Chile	Pseudoterranova spp.	104	4.8	47
				78	21.8	51
		Ecuador	Anisakis pegreffi	62	92	38
Merluccius gayi			Anisakis physeteris	62	92	38
	Hake	Chile	Anisakis simplex	17	5.9	47
		Colombia	Family Anisakidae	126	26.19	36
M I · I II ·		Chile	Pseudoterranova decipiens	17	23.5	47 37
Merluccius hubbsi	Argentine hake	Argentina	Pseudoterranova sp.	79		53
		Venezuela	Contracecum sp. (s. l)	136	0.7	55
Micropogonias furnieri	"Corvina rubia"	Brazil	<i>Hysterothylacium deardorffoverstreetorum (s.l)</i>	60	3.3	48
Mola mola	Ocean sunfish	Chile	Anisakis sp.	1	100	54
	Flathead gray mullet	Colombia	Anisakis physeteris	12	33	38
Musiloonholus		Peru	Anisakis simplex	ND	ND	14
Mugil cephalus	"Lisa"	Colombia	Anisakis sp.	15	33	12
		Ecuador	Contracecum sp.	64	14	35
	"Lebranche"	Colombia	Family Anisakidae	19	100	36
	White mullet	Colombia	Anisakis physeteris	16	94	38
		Venezuela	Anisakis spp.	143	47.16	75
			Contracaecum spp.	143		75
Mugil curema	"Lisa"			90		44
			Pseudoterranova spp.	143		75
				90		44
	White mullet	Colombia	Pseudoterranova decipiens	16		38
			<i>Contracaecum</i> spp.	378		76 76
Mugil incilis	"Lisa"	Colombia	Pseudoterranova spp.	378		36
			Family Anisakidae	355		55
			Family Anisakidae	386 65		75
			<i>Anisakis</i> spp. <i>Contracaecum</i> spp.	65		75
Mugil liza	"Lebranche"	Venezuela	Contracaecum spp.	90		44
inugii nza	Lebranche	venezuela	contracaecum spp.	65		75
			Pseudoterranova spp.	90		44
<i>Mugil</i> spp.	Mugil	Colombia	Family Anisakidae	5		36
		ee.onbiu	Hysterothylacium	J	100	
Mullus argentinae	ND	Argentina	dearddorffoverstreetorum (s.l)	60	48.3	48
			Pseudoterranova sp.	2	100	37
			Anisakis sp.	37	5.4	40
Mustelus canis	ND	Brazil	Contracecum sp.	37	2.7	40
			Pseudoterranova sp.	37	10.8	40
Mustelus schmitti	ND	Brazil	Contracecum sp.	35	5.7	40
			Pseudoterranova sp.	35	3.8     0.7     3.3     100     33     ND     33     14     100     94     47.16     12.74     97     40.1     3     94     100     80.5     83.9     64.51     5.61     84.39     97     100     3     100     3     100     3     97     100     3     97     100     3     100     3     100     3     100     5.4     2.7     10.8	40

Host					Prevalence	
Scientific name	Common name	Country of origin	Genus and/or species	Sampled fish (n)	of infection (%)	Ref
Nemadactylus bergi	ND	Argentina	Pseudoterranova sp.	32	3.1	37
Odontesthes bonariensis	ND	Argentina	Contracecum sp.	ND	5	34
Oligosarcus jenynsii	ND	Argentina	Contracecum sp.	ND	54	34
Oreochromis niloticus	Nile tilapia	Ecuador	Contracecum sp.	64	14.06	35
Pagrus pagrus	Red porgy	Argentina	Anisakis simplex	38	23.8	56
		Brazil	Anisakis simplex (s.l)	148	85.13	56
			<i>Anisakis</i> sp.	36	5.56	57
				100	40	58
				ND	7.7	59
			Contracecum sp.	38	5.3	56
				36	8.33	57
				100	3	58
		Argentina		38	100	56
		<u>J</u>	<i>Hysterothylacium</i> sp.	36	13.89	57
				100	90	58
		Brazil		ND	93.3	59
				148	4.05	56
			Pseudoterranova sp.	100	6.6	58
Paralichthys isosceles	ND	Argentina	Pseudoterranova cattani	15	26.7	37
		Brazil	Hysterothylacium deardorffoverstreetorum sp. nov.	60	ND	78
Paralichthys microps	ND	Chile	Anisakis simplex	10	10	47
			Pseudoterranova decipiens	10	70	47
Pellona castelnaeana	ND	Brazil	Family Anisakidae	10	100	60
Percophis brasiliensis	Brazilian flathead	Argentina Uruguay	Anisakis simplex	177	5.08	61
		Brazil	Anisakis sp.	178	14.6	72
		Argentina Uruguay	Contracecum sp.	177	5.08	61
		Brazil	Contracecum sp.	178	6.74	72
			Hysterothylacium aducum	178	0.56	72
			<i>Hysterothylacium fortalezae</i>	178	2.8	72
		Argentina Uruguay	Hysterothylacium sp.	177	74.01	61
		Brazil	Hysterothylacium sp.	178	97.19	72
		Argentina	Pseudoterranova cattani	8	25	37
Pimelodus albicans	ND	Argentina	Contracecum sp.	ND	100	34
Pinguipes brasilianus	ND	Brazil	Hysterothylacium sp.	25	100	62
Plagioscion magdalenae	"Pacora"	Colombia	Contracecum sp.	60	46.7	22
Plagioscion squamosissimus	ND	Brazil	Family Anisakidae	5	100	60
Paralonchurus peruanus	"Suco"	Peru	Anisakis simplex	ND	ND	14
			Anisakis sp.	30	20	63
Priacanthus arenatus	ND	Brazil	Hysterothylacium deardorffoverstreetorum	30	66.7	63

Host		Country of		Sampled	Prevalence	
Scientific name	Common name	origin	Genus and/or species	fish (n)	of infection (%)	Ref
Prionotus nudigula	ND	Argentina	Pseudoterranova cattani	32	100	37
Prionotus punctatus	"Cabrinha"	Brazil	Anisakis sp.	80	17.5	77
			Hysterothylacium sp.	80	97.5	77
Pseudopercis númida	Namorado	Brazil	Anisakis typica	25	4	62
r seudopercis numida	sandperch	DIAZII	Hysterothylacium sp.	25	88	62
Pseudopercis semifasciata	Sea salmon	Argentina	Anisakis simplex	100	45	64
			Contracecum sp.	100	26	64
			Hysterothylacium aducum	100	43	64
			Hysterothylacium sp.	100	79	64
			Pseudoterranova sp.	100	35	64
			Pseudoterranova sp.	131	25.8	64
Pseudoplatystoma magdaleniatum	"Bagre pintado"	Colombia	Contracecum sp.	60	95	22
Rhamdia quelen	ND	Argentina	Contracecum sp.	ND	100	34
Salminus affinis	"Rubio"	Colombia	Contracecum sp.	45	95	49
Salminus maxillosus	"Dorado"	Argentina	Contracecum sp.	10	80	65
Sarda chiliensis	"Bonito"	Peru	Anisakis physeteris	ND	ND	14
Sciades herzbergii	Pemecou sea catfish	Colombia	Family Anisakidae	64	1.6	36
Sciaena deliciosa	"Lorna"	Peru	Anisakis simplex	ND	ND	14
			Anisakis physeteris	ND	ND	14
Scomber japonicus	Chub mackerel	Peru	Anisakis simplex	ND	ND	14
	"Bonito"	i ci u	Pseudoterranova decipiens	ND	ND	14
	"Estornino"	Argentina	Pseudoterranova sp.	13	7.7	37
Scyliorhinus haeckelii	ND	Brazil	Contracecum sp.	9	22.2	40
Sebastes capensis	ND	Chile	Anisakis simplex	487	36.3	66
			Pseudoterranova decipiens	487	14.6	66
Serrasalmus spilopleura	Dark-banded piranha	Argentina	Contracecum sp.	237	91.98	67
Sorubim cuspicaudus	"Blanquillo"	Colombia	Contracecum sp.	60	100	22
Sphyrna zygaena	ND	Brazil	<i>Contracecum</i> sp.	16	12.5	40
		2.02.0	Anisakis sp.	14	7.1	40
Squalus megalops	ND	Brazil	Pseudoterranova sp.	14	14.3	40
Squatina sp.	ND	Brazil	Anisakis sp.	20	3.8	40
Thunnus thynnus	ND	Brazil	Anisakis typica	ND	ND	68
Thyrsites atun	Sea pike	Chile	Anisakis sp.	ND	ND	69
			Anisakis sp. (Type I)	95	7.4	47
			Pseudoterranova spp.	95	5.3	47
Trachurus murphyi	Chilean jack mackerel	Peru	Anisakis physeteris	ND	ND	14
		Chile	Anisakis physeteris	2 200	6.1	70
		Peru	Anisakis simplex	ND	ND	14
		Chile	Anisakis simplex	16	12.5	47
			Anisakis simplex	2 200	36.9	70
		Peru	Anisakis simplex	843	1.8	70
		Chile	Hysterothylacium sp.	2 200	7.3	70
		Peru	<i>Pseudoterranova decipiens</i>	2 200 ND	ND	14
		reiu	Pseudoterranova decipiens	16		47
		Chile			31.3	70
Trichiurus lepturus	Atlantic	Brazil	<i>Pseudoterranova decipiens</i> <i>Anisakis typica</i>	2 200 64	0.045 57	71
·	cutlassfish					27
<i>Xystreurys rasile</i> ID: no data.	ND	Argentina	Pseudoterranova sp.	29	3.4	37

ND: no data. Source: Own elaboration. The present systematic review also made it possible to establish, for the first time, the geographic distri-

bution of the different species of anisakid nematodes reported in fish marketed in South America (Figure 2).



**Figure 2.** Geographical distribution of anisakid nematodes reported in fish for human consumption in South America. Source: Own elaboration.

#### Discussion

The presence of nematodes of the family *Anisakidae* in fish for human consumption in South American countries, both in the Atlantic and Pacific Oceans, is significant. However, cases of anisakidosis reported in humans are few and mostly localized in Chile and Peru, leading to the idea that this infection is present in the region and may be a probable zoonosis that is not being detected due to a lack of awareness among healthcare professionals.

The recent increase in the availability of Mediterranean and Eastern dishes made from raw or undercooked fish and cephalopods in South America has increased the risk of infection and hence the number of cases of anisakidosis. This may also explain why most of the known cases in the region are found in Chile and Peru, countries where the gastronomic culture involves the consumption of marinated or salted seafood, mainly ceviche.

According to the findings of the present study, it is noteworthy that the genera *Anisakis* and *Pseudoterranova* 

were identified in the registry of parasitized fish for consumption since they are the parasites with the greatest impact on human health in countries where anisakidosis is highly prevalent, such as Spain and Japan;<sup>11</sup> however, this is not reflected in the clinical cases described.

Similarly, this registry included fish parasitized by the genera *Contracaecum* and *Hysterothylacium*, of which, at the time of writing this review, no infections had been reported in humans in South America. In contrast, there have been case reports of infection with *Hysterothylacium aduncum* (two in South Korea and two in Japan) and *Contracecum osculatum* (one in Japan) from across the world,<sup>7</sup> indicating either a possible emerging disease or a severe underreporting of this infection in the region.

Reports of anisakidosis in South America are relatively new, except for the first case reported in Chile in 1976, according to Jofré *et al.*,<sup>3</sup> as they were published in the last 25 years. Moreover, in the region, there is evidence of an increase in the registration of cases between 1997 and 2005, followed by a large number of studies, of which the most recent was published in 2018.<sup>28</sup>

As mentioned above, most cases of anisakidosis were found in Chile  $(n=35)^{3,13,18-23}$  and Peru (n=8),  $^{13,14,24-26}$ where consumption of raw fish in the form of ceviche is common in coastal areas. However, due to the long periods between cases, healthcare personnel may not issue a disease warning if they are considered rare cases. In addition, in Brazil, Argentina and Colombia, only one report was found in each country.<sup>19,27,28</sup>

It should be noted that the report made public in Colombia was based on studies conducted by Castellanos *et al.*,<sup>38</sup> Castellanos-Garzón<sup>79</sup> and Castellanos *et al.*,<sup>80</sup> all published in 2018, who identified for the first time the presence of *A. physeteris* and *P. decipiens* in fish for human consumption marketed in the country, specifically in the port of Buenaventura. As a result, a warning about the possibility of anisakidosis being an emerging disease was released.

Gastrointestinal anisakidosis was the most frequent in South America, with a total of 45 cases.<sup>3,13-19,24-28,30-32</sup> The oropharyngeal, gastric, intestinal and gastrointestinal presentations were also observed, the oropharyngeal being the most common with 29 cases. In addition, 4 asymptomatic cases with expulsion of larvae through the oral cavity were found, as well as 12 cases with clinical symptoms that varied depending on the section of the gastrointestinal tract affected by the larvae; regarding the latter, cough, pain and tingling in the pharynx, acute epigastric pain (pyrosis), nausea and abdominal cramps were the most frequent symptoms. In general, patients' condition improved in the following 36 hours to 15 days after ingestion.

In 41 (91.1%) cases of gastrointestinal anisakidosis, the infection was caused by a single larva in the late stages (L3 and L4), which was identified taxonomically. At this point it is important to note that diagnosing anisakidosis can be problematic because larval identification only allows for genus identification, and species can only be determined morphologically in adult parasites found in marine mammals. Molecular biology techniques that are costly and not available in the region are required to identify the larval stage of the species.

Despite the above, Weitzel *et al.*, <sup>17</sup> using the DNeasy Blood and Tissue Kit molecular diagnostic technique, identified *Pseudoterranova cattani* as the causal agent of gastrointestinal anisakidosis in the oropharyngeal form in five patients from Chile. Furthermore, Castellanos *et al.*<sup>36</sup> described the species *A. physeteris and P. decipiens* in fish from the Pacific Ocean in Colombia and Ecuador using molecular biology and the multiplex PCR technique.

Similarly, three studies were found in the literature, two from Brazil and one from Venezuela, in which an allergic response to *A. simplex* allergens was reported. <sup>1,29,33</sup> This is evidence that people who have come into contact with *Anisakis* spp. and have formed antibodies against it are present in the region and are more likely to have an allergic reaction, and even an anaphylactic response, in a posterior contact through a type I hypersensitivity reaction.

Based on the information obtained from the clinical case reports, it was established that 95 species of anisakid intermediate fish hosts have been reported in South America, which should be studied in depth as they are the direct source of infection in humans. These species were found in Argentina, Brazil, Chile, Colombia, Ecuador, Peru and Venezuela<sup>11-13.15,19-22,33-77</sup>, as described below.

In Argentina, 22 species of fish for human consumption were identified. They were parasitized with species of the genera *Contracaecum*, *Pseudoterranova*, and *Hysterothylacium* and the species *Hysterothylacium aducum*, *A. simplex* and *P. cattani*, with a variable prevalence of up to 100% in several hosts.<sup>34,37,56</sup>

The largest number of host fish species was identified in Brazil, 34 in total, which were parasitized by *A. typica*, *A. physeteris*, *A. simplex*, *P. azarasi*, *Contracaecum* sp., *Anisakis* sp., *Pseudoterranova* sp., *Hysterothylacium* sp., *H. fortalezae* and *H. deardorffoverstreetorum*.<sup>20,23,39-41,45,46,48,50,56-60,62,63,68,71-73,77,78</sup>

In Chile, 15 hosts were identified and there were reports of infection with the genera *Anisakis*, *Pseudoterova* and *Hysterothylacium* and their species *A. simplex*, *A. physeteris*, *P. decipiens* and *H. geschei*.<sup>43,47,51,52,54,66,69,70,74</sup>

In Colombia, reports were found for 17 host species, including marine and inland water fish, in which the presence of the genera *Anisakis*, *Pseudoterranova* and *Contracaecum* and the species *A. physeteris* and *P. decipiens* was established by taxonomic identification.<sup>12,22,36,38,49,55,76</sup>

In Ecuador, 8 host fish species were identified: Ulloa-Ulloa & Carrasco<sup>38</sup> reported the presence of *Contracaecum* sp. in flathead grey mullet (*Mugil cephalus*), "trahira" (*Hoplias microlepis*), "bagre pintado" (*Hexanematichthy* sp.) and tilapia (*Oreochromis niloticus*), while Castellanos *et al.*, <sup>38</sup> in a study carried out on the country's Pacific coast, identified the species *A. physeteris* and *A. pegreffi* in hake (*Merluccius gayi*); these authors also compiled other reports of infection by *A. physeteris* in bullet tuna (*Auxis rochei*), "bonito" (*Katsuwonus pelamis*) and "dorado" (*Coryphaena hippurus*).

In Peru, which was the country with the second highest number of reports of clinical cases of anisakidosis, 7 species of anisakid fish hosts were identified. *Anisakis* sp, *A. physeteris*, *P. decipiens*, *A. simplex*, *Pseudoterranova* sp., *Contracaecum* sp. and *Hysterothylacium* sp. were isolated.<sup>13,14,70</sup>

In Venezuela, Vicente *et al.*, <sup>53</sup> Bracho-Espinosa *et al.*<sup>44</sup> and Maniscalchi-Badaoui *et al.*<sup>75</sup> reported that two species of fish of the family *Mugilidae* were hosts of the

genera Anisakis, Pseudoterranova and Contracaecum. In addition, reports of infestation by Contracaecum sp. and Pseudoterranova sp. in the fish Eugerres plumieri and Micropogonias furnier were found in this country.<sup>44,53,75</sup>

Finally, in Uruguay, only two hosts with infection by *A. simplex*, *Contracaecum* sp., *Hysterothylacium* sp. and *Pseudoterranova* sp. were reported.<sup>42,61</sup>

Mullets, especially those of the species *M. cephalus*, rank first among all fish reported to have been infected by nematodes of the family *Anisakidae*, with a prevalence of infection of up to 100%. The high percentage of parasitization in mullets can be attributed to their geographical distribution, as they are found in both Atlantic and Pacific coastal waters, being species of economic and commercial importance.

Among the species with a high number of reports are also Brazilian flathead, which was parasitized by *Hysterothylacium* sp. (74.01%), *A. simplex* (51.41%), *P. cattani* (25%) and *Contracaecum* sp. (6.74%) and *Hoplias malabaricus*, which were all parasitized by *Contraecum* sp. A remarkable finding was that in about 45% (n=43) of the hosts, multiple infection by two or more species of parasites of the family *Anisakidae* was reported.

Based on the above, it is possible to establish that legislation on handling, processing, early evisceration and freezing at standard temperatures (-20°C) of fish is necessary in South America to adequately control the number of infected animals and thus prevent anisakidosis. At the same time, the study of this infectious disease should be promoted in healthcare institutions, with a focus on disease prevention and early management to effectively monitor the emergence of new cases and reduce underreporting.

#### Conclusions

The present review established the current panorama of the reported intermediate hosts for anisakids and clinical case reports of anisakidosis in South America. It was determined that this infectious disease is a latent risk for the region, so it is necessary to establish effective regulations to control its occurrence and provide more information to the general population on the necessary precautions regarding the consumption of saltwater fish.

#### **Conflicts of interest**

None stated by the authors.

#### Funding

None stated by the authors.

### Acknowledgments

To Call 567 for the training of national PhDs of the Department of Science Technology and Innovation of Colombia (COLCIENCIAS). To María Teresa Alarcón, for her guidance to develop the search algorithm. To Carlos Quilindo for the design of the distribution map of the species.

# References

1. Figueiredo I, Vericimo MA, Cardoso LR, São Clemente SC, do Nascimento ER, Teixeira GA. Cross-sectional study of serum reactivity to Anisakis simplex in healthy adults in Niterói, Brazil. Acta Parasitol. 2013;58(3):399-404. https://doi.org/f5cz.

- Hochberg NS, Hamer DH. Anisakidosis: Perils of the deep. Clin Infect Dis. 2010;51(7):806-12. https://doi.org/fj653q.
- Jofré L, Neira P, Noemí I, Cerva JL. Pseudoterranovosis y sushi. Rev. Chil. Infectol. 2008;25(3):200-5. https://doi.org/ccnkwv.
- Amo-Peláez M, Muñoz-Codoceo C, Martínez-Montiel P, Sánchez-Gómez F, Castellano G, Solís-Herruzo JA. Anisakiasis múltiple. Rev Esp Enfermedades Dig (Madrid). 2008;100(9):581-2.
- Cabrera R. Anisakiasis outbreak by Anisakis simplex larvae associated to Peruvian food in Spain. Rev Esp Enfermedades Dig (Madrid). 2010;102(10):610-1.
- Picó-Durán G, Pulleiro-Potel L, Abollo E, Pascual S, Muñoz P. Molecular identification of *Anisakis* and *Hysterothylacium* larvae in commercial cephalopods from the Spanish Mediterranean coast. Vet Parasitol. 2016;220:47-53. https://doi.org/f8g6gr.
- Takahashi S, Ishikura H, Kikuchi K. Anisakidosis: Global Point of View. In: Ishikura H, Aikawa M, Itakura H, Kikuchi K, editors. Host Response to International Parasitic Zoonoses. Springer; 1998. p. 109-120.
- Cuéllar C, Daschner A, Valls A, De Frutos C, Fernández-Fígares V, Anadón AM, et al. Ani s 1 and Ani s 7 recombinant allergens are able to differentiate distinct Anisakis simplex-associated allergic clinical disorders. Arch Dermatol Res. 2012;304(4):283-8. https://doi.org/fxpv8t.
- Minciullo PL, Cascio A, David A, Pernice LM, Calapai G, Gangemi S. Anaphylaxis caused by helminths: review of the literature. Eur Rev Med Pharmacol Sci. 2012;16(11):1513-8.
- Baird FJ, Gasser RB, Jabbar A, Lopata AL. Foodborne anisakiasis and allergy. Mol Cell Probes. 2014;28(4):167-74. https://doi.org/f5596k.
- Audícana MT, Ansotegui IJ, Fernández-de Corres L, Kennedy MW. Anisakis simplex: dangerous--dead and alive? Trends Parasitol. 2002;18(1):20-5. https://doi.org/brfz86.
- Castellanos JA, Tangua AR, Salazar L. Anisakidae nematodes isolated from the flathead grey mullet fish (*Mugil cephalus*) of Buenaventura, Colombia. Int J Parasitol Parasites Wildl. 2017;6(3):265-70. https://doi.org/gbw9md.
- Cabrera R, Suárez-Ognio L. Probable emergencia de anisakiosis por larvas de *Anisakis physeteris* durante el fenómeno El Niño 1997-98 en la costa peruana. Parasitol Latinoam. 2002;57(3-4):166-70. https://doi.org/cm4rfs.
- Cabrera R, Trillo-Altamirano MP. Anisakidosis: ¿Una zoonosis parasitaria marina desconocida o emergente en el Perú? Rev. Gastroenterol. Perú. 2004;24(4):335-42.
- Mercado R, Torres P, Muñoz V, Apt W. Human infection by Pseudoterranova decipiens (Nematoda, Anisakidae) in Chile: report of seven cases. Mem Inst Oswaldo Cruz. 2001;96(5):653-5. https://doi.org/d3v5wt.
- Torres P, Jercic MI, Weitz JC, Dobrew EK, Mercado RA. Human Pseudoterranovosis, an Emerging Infection in Chile. J Parasitol. 2007;93(2):440-3. https://doi.org/bwchzd.
- Weitzel T, Sugiyama H, Yamasaki H, Ramirez C, Rosas R, Mercado R. Human infections with Pseudoterranova cattani nematodes, Chile. Emerg Infect Dis. 2015;21(10):1874-5. https://doi.org/f5c3.
- Mercado R, Torres P, Gil LC, Goldin L. Anisakiasis en un paciente portadora de una pequeña hernia hiatal. Caso clínico. Rev Med Chil. 2006;134(12):1562-4. https://doi.org/dzgt29.
- Menghi CI, Comunale E, Gatta CL. Anisakiosis: primer diagnóstico en Buenos Aires, Argentina. Rev Soc Venez Microbiol. 2011;31(1):71-3.
- Mayo-Iñiguez A, Portes-Santos C, Paulo-Vicente AC. Genetic characterization of *Anisakis typica* and *Anisakis physeteris* from marine mammals and fish from the Atlantic Ocean off Brazil. Vet Parasitol. 2009;165(3-4):350-6. https://doi.org/dz7n6r.

- Timi JT, Paoletti M, Cimmaruta R, Lanfranchi AL, Alarcos AJ, Garbin L, *et al.* Molecular identification, morphological characterization and new insights into the ecology of larval *Pseudoterranova cattani* in fishes from the Argentine coast with its differentiation from the Antarctic species, P. *decipiens* sp. E (Nematoda: Anisakidae). Vet Parasitol. 2014;199(1-2):59-72. https://doi.org/f5phjc.
- 22. Wadnipar-Cano LM. Evaluación de la infección parasitaria por nemátodos anisákidos en peces de interés comercial en el municipio de San Marcos (Sucre). Manizales: Facultad de Ciencias Contables, Económicas y Administrativas, Universidad de Manizales; 2013.
- Dias Fde JE, Clemente SC de S, Knoff M. Larvae of Anisakidae nematodes and Trypanorhyncha cestodes of public health importance in Aluterus monoceros (Linnaeus, 1758) in Rio de Janeiro State, Brazil. Rev Bras Parasitol Vet. 2010;19(2):94-7.
- Cabrera R, Luna-Pineda MA, Suarez-Ognio L. Nuevo caso de infección humana por una larva de *Pseudoterranova decipiens* (Nematoda, Anisakidae) en el Perú. Rev Gastroenterol. Perú. 2003;23(3):217-20.
- Tanteleán M, Huiza A. Larva de nemátodos con importancia médica hallados en peces del mar de Perú y dos primeros casos de infección humana. Rev Peru Med Trop. 1993;7(1):61-5.
- Barriga JA, Salazar F, Barriga E. Anisakiasis: presentación de un caso y revisión de la literatura. Rev Gastroenterol Perú. 1999;19(4):317-23.
- Rosa-da Cruz A, de Sousa-Souto P, Bucalen-Ferrari CK, Marques-Allegretti S, Arrais-Silva WW. Endoscopic imaging of the first clinical case of anisakidosis in Brazil. Sci Parasitol. 2010;11(2):97-100.
- Patiño JA, Olivera MJ. Anisakiasis gastro-alérgica, primera descripción de un caso en Colombia y revisión de literatura. Biomédica. 2019;39(2):1-8. https://doi.org/f5k5.
- Puccio F, Cifarelli D, Blanco F, López E, Sarmiento L, Ordaz R, et al. Reactividad alérgica a Anisakis simplex y su asociación con asma bronquial en niños escolares del estado Nueva Esparta, Venezuela. Bol Mal Salud Amb. 2008;48:145-52.
- 30. Verhamme MA, Ramboer CH. Anisakiasis caused by herring in vinegar: a little known medical problem. Gut. 1988;29(6):843-7. https://doi.org/cg92n7.
- Torres M, Canales M, Concha M, Cofre X, Tellez P. Un caso de anisakiosis en un adulto. Parasitol. día. 2000;24(3-4):1-4. https://doi.org/dnw77b.
- Mercado R, Torres P, Maira J. Human case of gastric infection by a fourth larval stage of *Pseudoterranova decipiens* (Nematoda, Anisakidae). Rev Saude Publica. 1997;31(2):178-81. https://doi.org/ft8h7g.
- Figueiredo I, Vericimo M, Terra L, Ferreira T, Sao Clemente SC, Teixeira G. Association between immunoreactivity to *Ani-sakis* spp. antigens and high-risk pregnancy. Acta Parasitol. 2015;60(4):609-13. https://doi.org/f7s3bd.
- Mancini MA, Biole FG, Salinas VH, Guagliardo SE, Tanzola RD, Morra G. Prevalence, Intensity and ecological aspects of Contracaecum sp. (Nematode:Anisakidae) in freshwater of Argentina. Neotrop Helminthol. 2014;8(1):111-22.
- 35. Ulloa-Ulloa LA, Carrasco-Mancero W. Determinación de parásitos en pescados más comercializados en los mercados 10 de noviembre y 24 de mayo (Bellavista), ubicado en el cantón Guaranda, provincia Bolívar [tesis]. Granada, Ecuador: Universidad Estatal de Bolívar; 2008.
- Olivero Verbel J, Baldiris Avila R. Parásitos en peces colombianos: Están enfermando nuestros ecosistemas?. Cartagena: Editorial Universidad de Cartagena; 2008.
- 37. Hernández-Orts JS, Aznar FJ, Blasco-costa I, García NA, Víllora-montero M, Crespo EA, *et al.* Description, micro-

habitat selection and infection patterns of sealworm larvae (*Pseudoterranova decipiens* species complex, nematoda: Ascaridoidea) in fishes from Patagonia, Argentina. Parasites Vectors. 2013;6(252):1-15. https://doi.org/f5mp.

- Castellanos JA, Santana-piñeros AM, Mercado R, Peña S, Pustovrh C, Cruz-Quintana Y. Presence of anisakid larvae in commercial fishes landed in the Pacific coast of Ecuador and Colombia. Infect. 2018;22(4):206-12. https://doi.org/f5mq.
- Luque JL, Alves DR. Ecologia das comunidades de metazoários parasitos, do xaréu, Caranx hippos (Linnaeus) e do xerelete, Caranx latus Agassiz (Osteichthyes, Carangidae) do litoral do estado do Rio de Janeiro, Brasil. Rev. Bras. Zool. 2001;18(2):399-410. https://doi.org/d27mm8.
- 40. Knoff M, de São Clemente SC, Pinto RM, Gomes DC. Nematodes of elasmobranch fishes from the southern coast of Brazil. Mem Inst Oswaldo Cruz. 2001;96(1):81-7. https://doi.org/c56q64.
- Rodrigues MV. Presença do parasita anisaquideo em pescada (*Cynoscion* spp.) como ponto critico de controle na cadeia productiva do pescado comercializado na Baixada Santista. Sao Paulo: Instituto Biologico Sao Paulo; 2010.
- 42. Timi JT, Sardella NH, Navone GT. Parasitic nematodes of *Engraulis anchoita* Hubbs et Marini, 1935 (Pisces, Engraulidae) off the Argentine and Uruguayan coasts, South West Atlantic. Acta Parasitologica. 2000;46(3):186-93.
- 43. Chavez RA, Valdivia IM, Oliva ME. Local variability in metazoan parasites of the pelagic fish species, *Engraulis ringens*: Implications for fish stock assessment using parasites as biological tags. J Helminthol. 2007;81(2):113-6. https://doi.org/cj3fzf.
- Bracho-Espinoza H, Molina JD, Pirona M, Milagro C. Nematodos de la Familia Anisakidae en productos de la pesca, faja costera Médano Blanco, estado Falcón, Venezuela. Revista Científica FCV-LUZ. 2013;23(2):163-7.
- Marigo J, Taniwaki SA, Pinto PL, Soares RM, Catao-Dias JL. Molecular identification of *Pseudoterranova azarasi* larvae in cod (*Gadus* sp.) sold for human consumption in Brazil. Rev Inst Med Trop Sao Paulo. 2015;57(6):537-9. https://doi.org/f75zxg.
- de Paula Toledo Prado S, Capuano DM. Relato de nematóides da família Anisakidae em bacalhau comercializado em Ribeirão Preto, SP. Rev Soc Bras Med Trop. 2006;39(6):580-1. https://doi.org/bn7xjt.
- 47. Torres P, Puga S, Castillo L, Lamilla J, Miranda JC. Helmintos, myxozoos y microsporidios en músculos de peces comercializados frescos y su importancia como riesgo potencial para la salud humana en la ciudad de Valdivia, Chile. Arch Med Vet. 2014;46(1):83-92. https://doi.org/f5mt.
- Di Azevedo MIN, Iñiguez AM. Nematode parasites of commercially important fish from the southeast coast of Brazil: Morphological and genetic insight. Int J Food Microbiol. 2018;267:29-41. https://doi.org/gczdfk.
- Pardo S, Mejía K, Navarro Y, Atencio V. Prevalencia y abundancia de *Contracaecum* sp. en rubio (*Salminus affinis*) en el río Sinú y San Jorge: Descripción morfológica. Rev MVZ Córdoba. 2007;12(1):887-96.
- 50. Knoff M, de São Clemente SC, da Fonseca MCG, Felizardo NN, de Lima FC, Pinto RM, et al. Anisakidae nematodes in the blackfin goosefish, Lophius gastrophysus Miranda-Ribeiro, 1915 purchased in the State of Rio de Janeiro, Brazil. Acta Scientiarum. Biological Sciences. 2013;35(1):129-33. https://doi.org/f5mv.
- Torres-Frenzel P. Parasitos anisakidos en cebiche de merluza, comercializado en las localidades de Valdivia y Niebla, Chile. Valdivia: Universidad Austral de Chile; 2013.
- 52. Torres P, Contreras A, Revenga J, Fritz N. Helmith Parasites in Fishes From Valdivia and Tornagaleones River Estuaries in the South of Chile. Mem Inst Oswaldo Cruz. 1993;88(3):491-2. https://doi.org/bptm4h.

- Vicente JJ, Pinto RM, Aguilera O. On *Dichelyne (Cucullanellus)* elongatus (Tornquist, 1931) Petter, 1974: South American correlated species (Nematoda, Cucullanidae) and some other helminths of *Micropogonias furnieri* (Desmarest, 1823) (Pisces, Sciaenidae). Mem Inst Oswaldo Cruz. 1989;84(3):357-61. https://doi.org/czrvj6.
- Fernández I, Oyarzún C, Valenzuela A, Burgos C, Guaquín V, Campos V. Parásitos del pez luna *Mola mola* (Pisces: Molidae). Primer registro en aguas de la costa centro sur de Chile. Gayana (Concepción). 2016;80(2):192-7. https://doi.org/f5r8.
- 55. Olivero-Verbel J, Baldiris-Avila R, Arroyo-Salgado B. Nematode infection in *Mugil incilis* (Lisa) from Cartagena Bay and Totumo Marsh, north of Colombia. J Parasitol. 2005;91(5):1109-12. https://doi.org/bd2gwd.
- 56. Soares IA, Lanfranchi AL, Luque JL, Haimovici M, Timi JT. Are different parasite guilds of *Pagrus pagrus* equally suitable sources of information on host zoogeography? Parasitol Res. 2018;117(6):1865-75. https://doi.org/gdkjjx.
- 57. Saad CDR, Luque JL. Larvas de Anisakidae na musculatura do pargo, Pagrus pagrus, no Estado do Rio de Janeiro, Brasil. Rev Bras Parasitol Vet. 2009;18(Suppl 1):71-3. https://doi.org/c6cznm.
- Soares IA, Vieira FM, Luque JL. Parasite community of *Pagrus pagrus* (Sparidae) from Rio de Janeiro, Brazil: evidence of temporal stability. Rev Bras Parasitol Vet. 2014;23(2):216-23. https://doi.org/f5sj.
- Paraguassú AR, Luque JL, Alves DR. Community ecology of the metazoan parasites of red porgy, *Pagrus pagrus* (L., 1758) (Osteichthyes, Sparidae), from the coastal zone, state of Rio de Janeiro, Brazil. Acta Sci Maringá. 2002;24(2):461-7.
- 60. Farias-Rabelo NL, Muniz-e Silva TC, Ferreira-Araujo LR, da Silva-Pinheiro RE, Machado-da Rocha CA. Detection of Anisakidae larvae parasitizing *Plagioscion squamosissimus* and *Pellona castelnaeana* in the state of Pará, Brazil. Acta Sci Biol Sci. 2017;39(3):389-95. https://doi.org/f5sn.
- 61. Braicovich PE, Timi JT. Parasites as biological tags for stock discrimination of the Brazilian flathead *Percophis brasiliensis* in the south-west Atlantic. J Fish Biol. 2008;73(3):557-71. https://doi.org/d7mtcb.
- Pantoja CS, Borges JN, Santos CP, Luque JL. Molecular and Morphological Characterization of Anisakid Nematode Larvae from the Sandperches Pseudopercis numida and Pinguipes brasilianus (Perciformes: Pinguipedidae) off Brazil. J Parasitol. 2015;101(4):492-9. https://doi.org/f5sq.
- Kuraiem BP, Knoff M, Felizardo NN, Gomes DC, Clemente SS. Nematode larvae infecting *Priacanthus arenatus* Cuvier, 1829 (Pisces: Teleostei) in Brazil. An Acad Bras Cienc. 2016;88(2):857-63. https://doi.org/f5sr.
- 64. Timi JT, Lanfranchi AL. The metazoan parasite communities of the Argentinean sandperch *Pseudopercis semifasciata* (Pisces: Perciformes) and their use to elucidate the stock structure of the host. Parasitology. 2009;136(10):1209-19. https://doi.org/bn5b5z.
- Ramallo G, Torres P. Infección por larvas de Contracaecum sp. (Nematoda, Anisakidae) en Salminus maxillosus (Pisces, Characidae) en el embalse de Termas de Río Hondo, Argentina. Bol Chil Parasitol. 1995;50(1-2):21-3.
- 66. González MT, Barrientos C, Moreno CA. Biogeographical patterns in endoparasite communities of a marine fish (Sebastes capensis Gmelin) with extended range in the Southern Hemisphere. J Biogeogr. 2006;33(6):1086-95. https://doi.org/dk4n8x.
- 67. Hamann MI. Aspectos ecológicos de la relación parasitaria entre larvas de *Contracaecum* sp. (Nematoda, Anisakidae) y

*Serrasalmus spilopleura* Kner, 1860 (Pisces, Characidae) en poblaciones naturales del nordeste argentino. Bol Chil Parasitol. 1999;54(3-4):1-8. https://doi.org/ck78md.

- Mattiucci S, Paggi L, Nascetti G, Portes-Santos C, Costa G, Di Beneditto AP, *et al.* Genetic markers in the study of *Anisakis typica* (Diesing, 1860): Larval identification and genetic relationships with other species of Anisakis Dujardin, 1845 (Nematoda: Anisakidae). Syst Parasitol. 2002;51(3):159-70. https://doi.org/dtccwv.
- Peña-Rehbein P, De los Ríos-Escalante P. Use of negative binomial distribution to describe the presence of Anisakis in *Thyrsites atun.* Rev Bras Parasitol Vet. 2012;21(1):78-80. https://doi.org/f5sw.
- Oliva ME. Metazoan parasites of the jack mackerel *Trachurus* murphyi (Teleostei, Carangidae) in a latitudinal gradient from South America (Chile and Peru). Parasite. 1999;6(3):223-30. https://doi.org/f5sx.
- Novo-Borges J, Gulla-Cunha LF, Carneiro-Santos HL, Monteiro-Neto C, Portes-Santos C. Morphological and molecular diagnosis of anisakid nematode larvae from cutlassfish (*Trichiurus lepturus*) off the coast of Rio de Janeiro, Brazil. PLoS One. 2012;7(7):e40447. https://doi.org/f5tz.
- Braicovich PE, Pantoja C, Pereira AN, Luque JL, Timi JT. Parasites of the Brazilian flathead *Percophis brasiliensis* reflect West Atlantic biogeographic regions. Parasitology. 2017;144(2):169-78. https://doi.org/f9v5zp.
- 73. Andrade-Porto SM, Cárdenas MQ, Martins ML, Oliveira JKQ, Pereira JN, Araujo CSO De, *et al.* First record of larvae of *Hysterothylacium* (Nematoda: Anisakidae) with zoonotic potential in the pirarucu *Arapaima gigas* (Osteichthyes: Arapaimidae) from South America. Braz J Biol. 2015;75(4):790-5. https://doi.org/f8jbw7.
- 74. Torres P, Andrade P, Silva R. On a New Species of Hysterothylacium (Nematoda: Anisakidae) from Cauque mauleanum (Pisces: Atherinidae) by Brightfield and Scanning Electron Microscopy. Mem Inst Oswaldo Cruz. 1998;93(6):745-52. https://doi.org/ddz8nm.
- Maniscalchi-Badaoui MT, Lemus-Espinoza D, Marcano Y, Nounou E, Zacarías M, Narváez N. Larvas Anisakidae en peces del género *Mugil* comercializados en mercados de la región costera nor-oriental e insular de Venezuela. Saber. 2015;27(1):30-8.
- Ruiz L, Vallejo A. Parámetros de infección por nematodos de la familia Anisakidae que parasitan la lisa (*Mugil incilis*) en la Bahía de Cartagena (Caribe colombiano). Intropica. 2013;8(53):53-60.
- 77. Bicudo AJA, Tavares LER, Luque JL. Anisakidae larvae (Nematoda: Ascaridoidea) parasites of the bluewing searobin Prionotus punctatus (Bloch, 1793) (Osteichthyes: Triglidae) from the coastal zone of the State of Rio de Janeiro, Brazil. Rev Bras Parasitol Vet. 2005;14(3):109-18.
- Knoff M, Felizardo NN, Iñiguez AM, Maldonado A, Torres EJL, Pinto RM, et al. Genetic and morphological characterisation of a new species of the genus *Hysterothylacium* (Nematoda) from *Paralichthys isosceles* Jordan, 1890 (Pisces : Teleostei) of the Neotropical Region, state of Rio de Janeiro, Brazil. Mem Inst Oswaldo Cruz. 2012;107(2):186-93. https://doi.org/f5t6.
- Castellanos-Garzón JA. Nematodos anisákidos en peces de consumo y prevalencia de anticuerpos anti-Anisakis en una población de Colombia. Cali: Universidad del Valle; 2018.
- Castellanos JA, Tangua AR, Mercado R, Salazar L. First reporting of *Anisakis* sp. in the Armed Snook fish (*Centropomus armatus*) caught and commercialized in Buenaventura, Colombia. Infectio. 2018;22(3):136-40. https://doi.org/f5t7..