

Effect of de Carbon/Nitrogen ratio on the production of microalgae-based carotenoids

INGENIERÍA QUÍMICA

Efecto de la relación carbono/Nitrógeno en la producción de carotenoids en microalgas

Andrés Fernando Barajas-Solano^{1§}, Estefanía Guarín-Villegas¹, Linda Maciel Remolina-Páez¹,
Johanna Patricia Bermúdez-Castro¹, Sandra Oriana Mogollón-Londoño¹, Jefferson Eduardo
Contreras-Ropero¹, Janet Bibiana García-Martínez¹

¹*Universidad Francisco de Paula Santander, Faculty of Agricultural and Environmental Sciences,
Department of Environmental Sciences, Cúcuta, Colombia*

[§]*andresfernandobs@ufps.edu.co, estegu-96@hotmail.com, linda_1918@live.com, johannap97@gmail.com,
sandraorianaml@ufps.edu.co, jeffersoneduardocr@ufps.edu.co, janetbibianagm@ufps.edu.co*

(Recibido: 27 de Junio de 2019 - Aceptado: 24 de Julio de 2019)

Abstract

This study investigates the effect of C/N ratio on the production of biomass and total carotenoids on a *Scenedesmus* sp. Initially, three different carbon sources (sodium carbonate, sodium bicarbonate and sodium acetate) were tested under different concentrations of a nitrogen source (sodium nitrate) in 250 mL tubular air-lift reactors. The reactors were operated at 25 °C for 40 days. in light:dark cycle of 12:12, under a continuous flow of air. Results showed that by the adjustment of the concentration of the carbon and nitrogen source, it is possible to increase the concentration of biomass up to 0.8 g/L. However, by the regulation on the concentration of sodium carbonate and sodium nitrate, the final content of total carotenoids was increased two times (from 0.3 to 0.66 % w/w). Results from this study shows that an specific ratio between the carbon source employed and the concentration of the nitrogen source shows that an outstanding increase on the final biomass and the concentration of total carotenoids that can be produced. Finally, the effect of well-known strategies such as light, salinity and pH, coupled with C/N ratio must be studied to achieve a proper method to stress the cell culture and enhance the synthesis of carotenoids in *Scenedesmus* sp.

Keywords: *Biomass production, Photobioreactor, Scenedesmus sp, Sodium carbonate, Sodium nitrate.*

Resumen

El presente trabajo de investigación tiene como objetivo determinar el efecto de la relación Carbono/Nitrógeno en la producción de biomasa y carotenoides totales en una cepa de *Scenedesmus* sp. Inicialmente, se evaluaron tres fuentes de carbono diferentes (carbonato de sodio, bicarbonato de sodio y acetato de sodio) bajo diferentes concentraciones de una fuente de nitrógeno (nitrato de sodio) en reactores tubulares de 250 ml. Los reactores fueron operados a 25°C durante 40 días en un ciclo de luz:oscuridad de 12:12 horas y un flujo continuo de aire. De acuerdo con los resultados se encontró que mediante el ajuste de la concentración de la fuente de carbono y nitrógeno, es posible aumentar la concentración de biomasa hasta 0.8 g/L. Por otra parte, mediante la regulación de la concentración de carbonato de sodio y nitrato de sodio, el contenido final carotenoides totales se incrementó dos veces (de 0.3 a 0.66% p/p). Los resultados de este estudio muestran que, al ajustar las concentraciones de la fuente de carbono y de nitrógeno es posible obtener un aumento interesante en la biomasa final y la concentración de carotenoides totales. Finalmente, es importante resaltar que se debe estudiar el efecto de otras estrategias como la luz, la salinidad y el pH, junto con la relación C/N para obtener un método adecuado que lleve a las células hacia un estrés metabólico y mejore así la síntesis de carotenoides en *Scenedesmus* sp.

Palabras clave: Carbonato de sodio, Fotobioreactor, Nitrato de sodio, Producción de biomasa, *Scenedesmus* sp.

1. Introduction

Microalgae are a novel source of different metabolites and products for different industries such as pharmaceutical, animal feed, nutraceuticals, cosmetics, biofuels as well as a large number of products such as polyunsaturated fatty acids, antioxidants, dyes, fertilizers, soil conditioners, bioflocculants, biodegradable polymers, and polysaccharides.

The production of microalgae for obtaining high value products can be a promising business for a country like Colombia, since according to the BACEX Foreign Trade Data Bank of Colombian government, during the last 9 years, different segments of the Colombian Industry have imported USD 68 Million in coloring pigments for food (carotenoids, phycocyanins, chlorophylls, and others). The latter represents an essential segment due to the growing demand by the pharmaceutical, food, and cosmetic industries, where there are no Colombian companies that produce these metabolites.

Among all the products obtainable from microalgae, dyes (especially phycocyanins and carotenoids) are the most required raw materials

in the national pharmaceutical and food industry^(1,2); which are imported in considerable quantities due to the lack of companies present in the national territory dedicated to the production of this type of raw materials.

Carotenoids are fat-soluble substances with colors ranging from brown, red, orange to yellow. They perform two critical functions in photosynthesis: 1) absorb light in regions of the visible spectrum, in which chlorophyll is not efficiently absorbed; 2) protect photosynthetic systems. The photoprotection mechanisms eliminate the more active states of chlorophyll, as a result of the excessive absorption of light radiation. The latter hinders the formation of reactive oxygen species (ROS), makes carotenoids good antioxidants⁽³⁾. The main carotenoids of microalgae are β -carotene, lycopene, astaxanthin, zeaxanthin, violaxanthin, and lutein.

Carotenoids are essential for microalgae since they act as a protective barrier against high light radiations and dissipate excess energy in the form of heat⁽⁴⁾. According to the literature, strains such as *Muriellopsis* sp⁽⁵⁾, *Scenedesmus* sp⁽⁶⁾, *Chlorella zofingensis*⁽⁷⁾, *Chlorella*

protothecoides⁽⁸⁾, *Chlorella sorokiniana*^(9,10), *Coccomyxa acidophila*⁽¹¹⁾ and *Scenedesmus almeriensis*⁽⁶⁾ are potential candidates for obtaining at the industrial level of carotenoids with industrial interest; However, it is necessary to make adjustments to the cultivation conditions (light, concentration of nutrients, carbon source) that allow improving the metabolism of the product of interest without affecting the total cost of the process.

The carotenoid market has grown steadily in recent years, registering values close to USD 1.24 trillion in 2016 and is expected to reach USD 1.53 trillion in 2021. The β -carotene market is approximately 1200 tons per year (261 million dollars in 2010) and is expected to be 334 million dollars in 2018; currently, its sale price ranges from USD 300/kg to USD 3000/kg, depending mainly on the final concentration and the type of product^(12,13). On the other hand, natural Astaxanthin has a market of 300 tons per year with an approximate cost of 1.2 million USD and can reach a sale price of up to USD 7000/kg⁽¹⁴⁾.

Nitrogen (both NO_3^- and NH_4^+) is one of the critical nutrients in the production of biomass and different algal metabolites⁽¹⁵⁾. Several algal biotechnological processes need to exploit this feature to improve the performance of specific metabolites such as carbohydrates⁽¹⁶⁾ lipids⁽¹⁷⁾, hydrocarbons⁽¹⁸⁾ and biomass⁽¹⁹⁾. The objective of this work is the evaluation of the carbon/nitrogen ratio on the production of microalgae-based carotenoids in a strain of *Scenedesmus* sp.

2. Methodology

2.1. Microorganism

Scenedesmus sp Scene_UFPS01 was obtained from the collection of microorganisms of the Francisco de Paula University Santander

(Colombia) and maintained in Bold Basal medium⁽²⁰⁾. Initially, the strain was cultured in 500 mL glass reactors with 250 mL of sterile basal Bold medium, agitated by the injection of prefiltered air at an approximate flow of 150 mL air/min, constant radiation of $110 \mu\text{molm}^{-2}\text{s}^{-1}$ and a light-dark cycle of 12:12 hours.

2.2. Carbon/Nitrogen ratio

The effect of the Carbon/nitrogen ratio on the biomass growth and total carotenoids deposition was tested. First, three different carbon sources were evaluated: sodium acetate, sodium carbonate, and, sodium bicarbonate. The application of a 3^2 Central non-factorial design (3 levels, two factors) with response surface experimental design was applied for each of the carbon sources. The experimental design was evaluated using STATISTICA 7.0 software⁽²¹⁾ (Table 1).

Table 1. Variables and levels for the evaluation of C/N ratio

Factors	Levels		
Carbon source (g/L)	0.3	0.4	0.5
Sodium nitrate (mL/L)	6	7	8

The final concentration of available carbon on each of the carbon sources was adjusted according to their chemical formula (Table 2- 4). The nitrogen stock used in the Bold basal media (25 g/L of NaNO_3) was used. As a control, Bold Basal medium, without the addition of extra carbon and a NaNO_3 concentration of 10 mL/L (0.25 g/L) was employed.

Each of the experiments was carried out using 250 mL of Bold basal medium⁽²⁰⁾ with the adjusted nitrogen and carbon source and 30 mL of pre-cultured *Scenedesmus* sp (12% v/v,

alga/medium). Each of the experiments was maintained agitated by injecting prefiltered air at an approximate flow of 150 mL air/min, constant radiation of $110 \mu\text{mol m}^{-2} \text{s}^{-1}$ and a light:dark cycle of 12:12 hours during 40 days.

Table 2. Experiments for Sodium Acetate/Sodium nitrate ratio

Experiment	Sodium Acetate (g/250mL)	Sodium nitrate (mL/250mL)
8	0.26	1.40
5	0.26	1.75
3	0.34	1.50
1	0.17	1.50
2	0.17	2.00
9	0.26	2.10
6	0.14	1.75
7	0.38	1.75
4	0.34	2.00

Table 3. Experiments for Sodium bicarbonate/Sodium nitrate ratio.

Experiment	Sodium bicarbonate (g/250 mL)	Sodium nitrate (mL/250mL)
8	0.53	1.40
5	0.53	1.75
3	0.70	1.50
1	0.35	1.50
2	0.35	2.00
9	0.53	2.10
6	0.28	1.75
7	0.77	1.75
4	0.70	2.00

Table 4. Experiments for Sodium carbonate/Sodium nitrate ratio.

Experiment	Sodium carbonate (g/250 mL)	Sodium nitrate (mL/250mL)
8	0.66	1.40
5	0.66	1.75
3	0.88	1.50
1	0.44	1.50
2	0.44	2.00
9	0.66	2.10
6	0.35	1.75
7	0.97	1.75
4	0.88	2.00

2.3. Biomass and total carotenoids quantification

Total biomass (in dry weight) was measured according to the method described by ⁽²²⁾ GF/C filters were pre-combusted for 1 hour at 100°C and stored in a desiccator for up to 4 hours, their weight (Cell-free) was recorded employing a 6-digit analytical balance and stored in Petri dishes on a silica gel bed until their use. Once the experiments were completed, 80 mL of medium was filtered (by triplicate) and dried at 60 ° C overnight. The next day the samples were taken to desiccator until a constant weight was reached, and the final weight was recorded using a 6-digit analytical balance.

The extraction and quantification of total carotenoids were developed using the method proposed by ⁽²³⁾. The filtered biomass obtained from the previous step was suspended in 20 mL of phosphate buffer (8 mM Na₂HPO₄, 2 mM NaH₂PO₄, 140 mM NaCl, pH 7.4) and mixed with 10 mg of 0.2 mm glass beads. The mixture was stirred using a vortex at maximum speed for 10 minutes. To separate the carotenoids, 5 mL of chloroform was added and centrifuged at 3400 RPM for 8 minutes. The process was repeated

until the pellet was colorless. The chloroform fraction was stirred and concentrated by rotoevaporation. Concentrated carotenoids were suspended again in chloroform and read spectrophotometrically using Eq 1.

$$\begin{aligned} \text{Total carotenoids (mg/L)} \\ = \text{Abs}_{464nm} - 0.0222/0.0325 \end{aligned} \quad (\text{Eq 1})$$

3. Results and discusión

Scenedesmus sp is an alga belonging to the class Chlorophyceae, which is a small and immobile colony-forming cell in which they are aligned in the form of a plate. The cells contain a single nucleus, which consists of a chloroplast in the central part. Figures 1 and 2 show the different cultures carried out with Scene_UFPS01, in the same way, it is possible to recognize the characteristic morphology of the genus.

In recent years, the design of culture media that adjust to the needs of the strain to be cultured has become increasingly frequent, as an alternative to traditional media. Therefore, they are designed using specific concentrations of sodium nitrate, potassium phosphate, sodium acetate, and other critical nutrients for the production of biomass and deposition of metabolites of industrial interest. It has been found that, for media with acetate, there are no significant variables that influence the production of proteins, while, in media with carbonate, sodium nitrate and potassium phosphate significantly influence the production of this metabolite ⁽²⁴⁾.

Under stress conditions, some microalgae positively regulate specific biosynthetic pathways, which leads to the accumulation of specific compounds. For example, the change in the composition of nutrients can induce stress in

the physiological activities of algae, which can trigger a marked increase in the production of carotenoids.



Figure 1. (a) Production under lab conditions

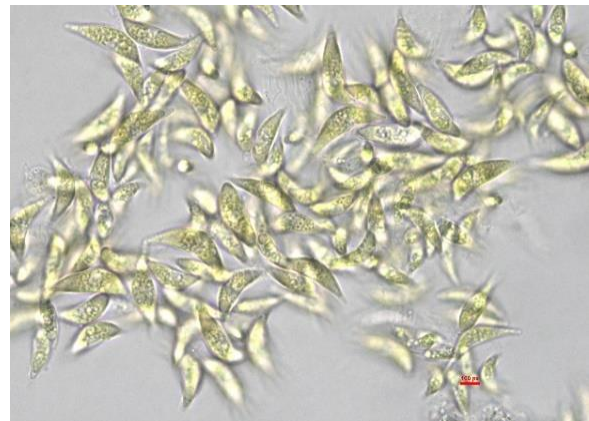


Figure 1. (b) Microscopic revision of monoalgal culture

For example, physiological stresses induced by nitrogen (N) limitation, high salinity, or extreme light intensity can initiate the synthesis of secondary carotenoids ⁽²⁵⁾. Recent studies indicate possibilities that microalgae can be used as a natural source of bioactive compounds, such as carotenoids, which have high antioxidant properties. These products can have several applications in nutrition and human or animal pharmacology ⁽²⁶⁾.

According to the results presented in Figure 2; by the adjustment of the carbon (either carbonate, bicarbonate or acetate) and nitrogen source (sodium nitrate), it is possible to increase the concentration of biomass. In the case studied, values of up to 0.8 g/L of biomass (in dry weight) were obtained, which is double that of the control (0.4 g/L).

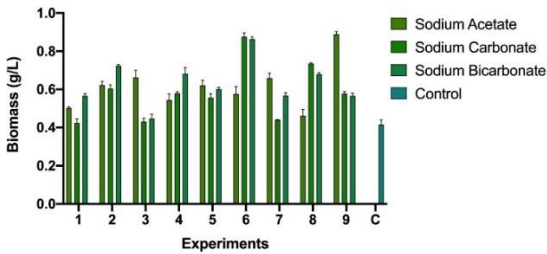


Figure 2. Biomass concentration for the three carbon sources evaluated.

Figure 3 shows the content of carotenoids concerning the biomass obtained for the strain Scene_UFPS01. According to the results, sodium carbonate has the highest values of total carotenoids in all its experiments, with values higher than the control (0.302% w/w) and up to 0.66% P / P. On the other hand, sodium acetate, and sodium bicarbonate obtained values very close to the control. Přibyl, et al. (23) obtained higher concentrations of carotenoids for *Dunaliella salina* (8-10% w/w) and *Haematococcus pluvialis* (6% w/w) under stress due to high deficiency of light, salinity and nutrients, which allows appreciate that the implementation of other stressors may be necessary to encourage the synthesis of total carotenoids.

The different results obtained through the application of the design of experiments are better understood through statistical analyzes obtained in the Pareto diagram and response surface graphs, which allow establishing the

influence of the variables (carbon vs. sodium nitrate). For the study case, the total carotenoid concentration (in% w/w) was the response variable, which can be seen in Figure 4 (a, b and c). Figure 4b for sodium carbonate shows the significant influence of the carbon concentration in linear function since it exceeds the threshold $p = 0.5$; however, for the other carbon sources, the variables do not present a significant influence.

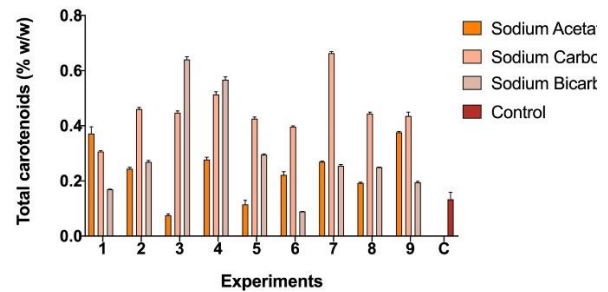


Figure 3. Total carotenoids concentration for the three carbon sources evaluated.

Regarding the data obtained from the response surfaces Figure 4 (a, b, and c), it is sodium acetate that reaches values of up to 1% in high concentrations of sodium and carbon nitrate or deficient concentrations of both, exceeding the values obtained in control.

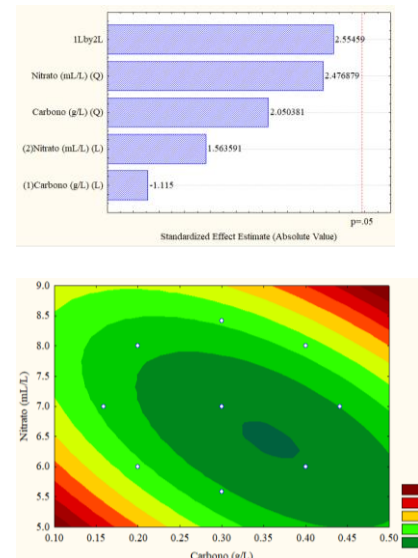


Figure 4a. Pareto diagram and surface response graphic for sodium acetate/sodium nitrate

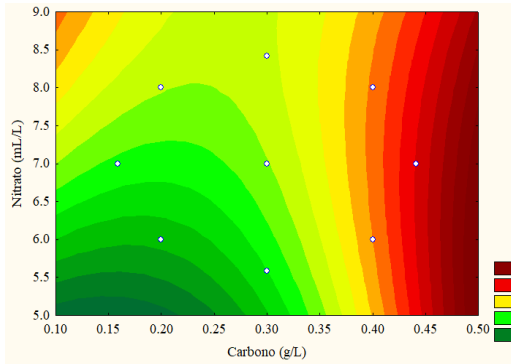
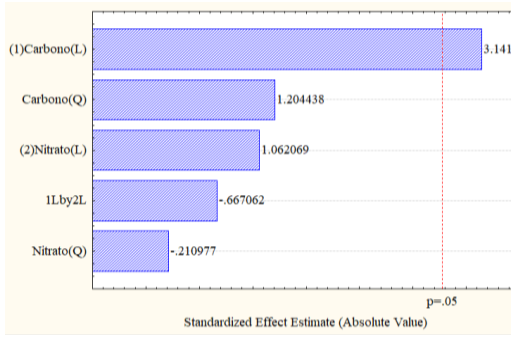


Figure 4b. Pareto diagram and surface response graphic for sodium carbonate/sodium nitrate

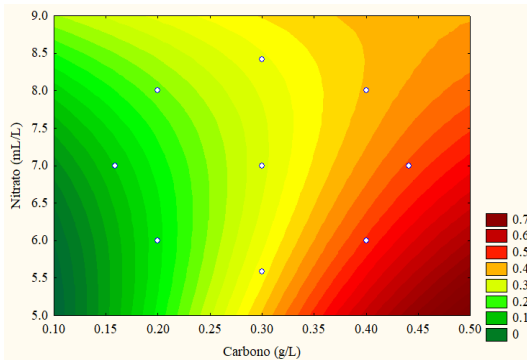
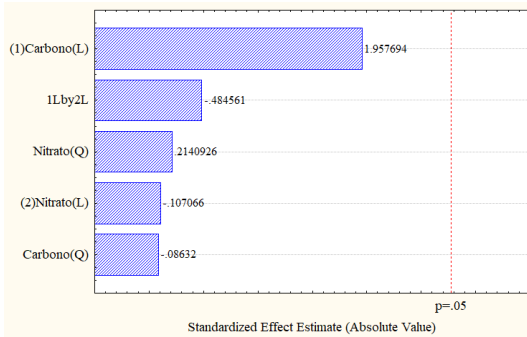


Figure 4c. Pareto diagram and surface response graphic for sodium bicarbonate/sodium nitrate

Over the last 20 years, different and unique strains of *Scenedesmus* sp has been tested for the production of lipids and carotenoids. Table 5 summarize the most relevant studies on this area.

Table 5. Comparison table on the concentration of total carotenoids for different strains.

Microalgae	Total Carotenoids	Reference
<i>Scenedesmus oblicuos</i>	17.4 mg/L	Bishop, 1996 (27)
<i>Scenedesmus quadricauda</i>	1.75 mg/L	Song & Pei, 2018 (28)
<i>Scenedesmus bijugus</i>	2.9 mg/g ó 0.47 mg/L	Minhas et al., 2016 (29)
<i>Scenedesmus almeriensis</i>	3.8 mg/L	Sanchez et al., 2008 (6)
<i>Scenedesmus oblicuus</i>	0.98 mg/g	Qin et al., 2008 (30)
<i>Scenedesmus quadricauda</i>	6.74 mg/g	Kozlova et al., 2017 (31)
<i>Scenedesmus bajacalifornicus</i>	25.07 mg/L	Patil & Kaliwal, 2016 (32)
<i>Scenedesmus</i> sp	20 mg/L	Přibyl et al., 2015 (33)
<i>Scenedesmus</i> sp	20-65 mg/L	This work Scene_UFPS01

3.1. Optimization of Carbon/Nitrogen ratio for biomass and total carotenoids production

Taking into account the results obtained, we obtained a series of equations that represent the mathematical models of carotenoid production according to the concentration of the carbon source and the source of nitrogen used. Table 6 presents the equations obtained from the response surface graphs and the operating conditions for each of the cases.

Table 6. Equations of C/N ratio.

Experiment	Equation % P/P	Carbon source (g/L)	Sodium nitrate (mL/L)
Control	-	-	10
Acetate 1	Z = 6.0479974522161 - 10.003528882418x + 6.3576591263102x ²	0.1	2
Acetate 2	- 1.292605102042y + 0.07680108027039y + 0.84679798968709y + 0	0.7	9.5
Carbonate 1	Z = -0.4214753703782 + 0.043354041205401 + 3.736567958183x ²	0.6	5
Carbonate 2	+ 0.1829099473951y - 0.00654521124834y - 0.22123319211982x + 0	1	5
Bicarbonate 1	Z = -0.1328636195980 + 4.7572139901267x - 0.73017190853271x - 0.12892867758564y + 0.018109867293955 - 0.43818540495544y + 0.0	0.6	2

The results obtained are presented in Figure 5, in which higher concentrations of total carotenoids were achieved than those obtained with the control (0.13% w/w) for acetate 2 and carbonate 1, which is related to the results obtained previously, due to the presence of concentrations less than 1 g/L of carbon and medium-high concentrations of sodium nitrate. Similar behaviors were found by ⁽³⁴⁾ who managed to improve the production of carotenoids with the use of sodium carbonate. On the other hand, ⁽³⁵⁾

did not find essential results in the implementation of diverse carbon sources since the content of carotenoids (Lutein) did not vary significantly.

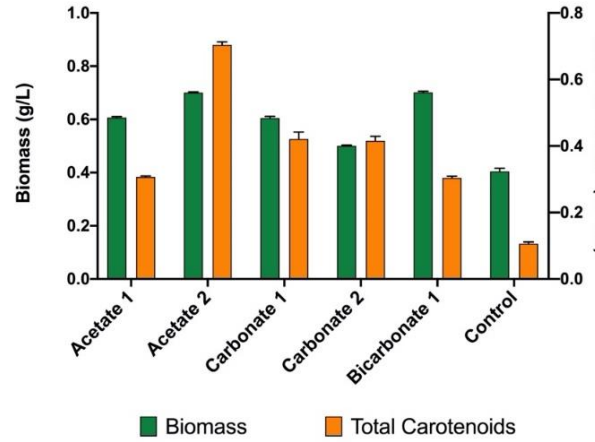


Figure 5. Biomass and total carotenoids for optimal conditions

4. Conclusions

The results of this study indicate that carbon/nitrate ratio It is an exciting process to improve the production of biomass and metabolites of industrial interest such as carotenoids. Concerning to carbon source, results shown that the alga can effectively use the different concentrations of sodium carbonate, sodium bicarbonate and sodium acetate, with an increase on biomass production. Of up to 0.7 g/L (with a control of 0.4 g/L). However, is the specific ratio between the carbon source employed and the concentration of the nitrogen source shows that an outstanding increase on the final biomass and the concentration of total carotenoids that can be produced. Finally, the effect of well-known strategies such as light, salinity and pH, coupled with C/N ratio must be studied to achieve a proper method to stress the cell culture and enhance the synthesis of carotenoids in *Scenedesmus* sp.

5. Funding Source Declaration

This research was partially funded by GEN FOUNDATION (England) with the project: ISOLATION OF TERMO-TOLERANT ALGAE AS NOVEL SOURCE OF FOOD COLORANTS. FINU (44-2018) project: USE OF RESIDUAL GLYCEROL FOR THE PRODUCTION OF HIGH-VALUE PRODUCTS FROM MICROALGAE-BASED MIXOTROPHIC CULTURES. and by National program for young research talents Jóvenes Investigadores e Innovadores COLCIENCIAS (753-2016).

6. Acknowledgments

We would like to express our sincere gratitude to Universidad Francisco de Paula Santander for providing equipment for successfully conclude this research and the Departamento Administrativo de Ciencia, Tecnología e Innovación COLCIENCIAS, for its Francisco José de Caldas scholarship program to support national PhD doctorates and its program for young research talents “Jovenes Investigadores e Innovadores”.

7. References

- (1) Ciccone MM, Cortese F, Gesualdo M, Carbonara S, Zito A, Ricci G, et al. Dietary Intake of Carotenoids and Their Antioxidant and Anti-Inflammatory Effects in Cardiovascular Care. *Mediators Inflamm.* 2013;2013:1–11. Doi: 10.1155/2013/782137.
- (2) Del Campo JA, García-González M, Guerrero MG. Outdoor cultivation of microalgae for carotenoid production: current state and perspectives. *Appl Microbiol Biotechnol.* 2007;74(6):1163–74. Doi: 10.1007/s00253-007-0844-9.
- (3) D’Alessandro EB, Filho NRA. Concepts and studies on lipid and pigments of microalgae: A review. *Renew Sustain Energy Rev.* 2016;58:832–41. Doi: 10.1016/j.rser.2015.12.162.
- (4) Hu J, Nagarajan D, Zhang Q, Chang JS, Lee DJ. Heterotrophic cultivation of microalgae for pigment production: A review. *Biotechnol adv.* 2018;36(1), 54–67. Doi: 10.1016/j.biotechadv.2017.09.009.
- (5) Del Campo JA, Moreno J, Rodríguez H, Vargas MA, Rivas J, Guerrero MG. Carotenoid content of chlorophycean microalgae: factors determining lutein accumulation in *Muriellopsis* sp. (Chlorophyta). *J Biotechnol.* 2000;76(1):51–9. Doi: 10.1016/S0168-1656(99)00178-9.
- (6) Sánchez JF, Fernández JM, Ación FG, Rueda A, Pérez-Parra J, Molina E. Influence of culture conditions on the productivity and lutein content of the new strain *Scenedesmus almeriensis*. *Process Biochem.* 2008;43(4):398–405. Doi: 10.1016/j.procbio.2008.01.004.
- (7) Del Campo JA, Rodríguez H, Moreno J, Vargas MA, Rivas J, Guerrero MG. Accumulation of astaxanthin and lutein in *Chlorella zofingiensis* (Chlorophyta). *Appl Microbiol Biotechnol.* 2004;64(6):848–854. Doi: 10.1007/s00253-003-1510-5.
- (8) Shi X-M, Zhang X-W, Chen F. Heterotrophic production of biomass and lutein by *Chlorella protothecoides* on various nitrogen sources. *Enzyme Microb Technol.* 2000;27(3–5):312–8. Doi: 10.1016/S0141-0229(00)00208-8.

- (9) Chen C-Y, Jesisca, Hsieh C, Lee D-J, Chang C-H, Chang J-S. Production, extraction and stabilization of lutein from microalga *Chlorella sorokiniana* MB-1. *Bioresour Technol.* 2016;200:500–5. Doi: 10.1016/j.biortech.2015.10.071.
- (10) Chen J-H, Chen C-Y, Hasunuma T, Kondo A, Chang C-H, Ng I-S, et al. Enhancing lutein production with mixotrophic cultivation of *Chlorella sorokiniana* MB-1-M12 using different bioprocess operation strategies. *Bioresour Technol.* 2019;278:17–25. Doi: 10.1016/j.biortech.2019.01.041.
- (11) Casal C, Cuaresma M, Vega JM, Vilchez C. Enhanced Productivity of a Lutein-Enriched Novel Acidophile Microalga Grown on Urea. *Mar Drugs.* 2011;9(1):29–42. Doi: 10.3390/md9010029.
- (12) García-González M, Moreno J, Cañavate JP, Anguis V, Prieto A, Manzano C, et al. Conditions for open-air outdoor culture of *Dunaliella salina* in southern Spain. *J Appl Phycol.* 2003;15(2–3):177–184. Doi: 10.1023/a:1023892520443.
- (13) León R, Martín M, Vígara J, Vilchez C, Vega JM. Microalgae mediated photoproduction of β -carotene in aqueous–organic two phase systems. *Biomol Eng.* 2003;20(4–6):177–82. Doi: 10.1016/S1389-0344(03)00048-0.
- (14) Gómez PI, Inostroza I, Pizarro M, Pérez J. From genetic improvement to commercial-scale mass culture of a Chilean strain of the green microalga *Haematococcus pluvialis* with enhanced productivity of the red ketocarotenoid astaxanthin. *AoB Plants.* 2013;5(plt026). Doi: 10.1093/aobpla/plt026.
- (15) Solovchenko AE, Khozin-Goldberg I, Didi-Cohen S, Cohen Z, Merzlyak MN. Effects of light intensity and nitrogen starvation on growth, total fatty acids and arachidonic acid in the green microalga *Parietochloris incisa*. *J Appl Phycol.* 2008;20(3):245–251. Doi: 10.1007/s10811-007-9233-0.
- (16) Jerez–Mogollón SJ, Rueda–Quiñonez LV, Alfonso–Velazco LY, Barajas–Solano AF, Barajas–Ferreira C, Kafarov V. Improvement of lab-scale production of microalgal carbohydrates for biofuel production. *CT&F - Ciencia, Tecnol y Futur.* 2012;5(1):103–16.
- (17) Lin C., Lay C. Carbon/nitrogen-ratio effect on fermentative hydrogen production by mixed microflora. *Int J Hydrogen Energy.* 2004;29(1):41–5. Doi: 10.1016/s0360-3199(03)00083-1.
- (18) Barajas–Solano AF, Guzman-Monsalve A, Kafarov V. Effect of Carbon–Nitrogen Ratio for the Biomass Production, Hydrocarbons and Lipids on *Botryococcus braunii* UIS 003. *Chem Eng Trans.* 2016;49:247–52. Doi: 10.3303/CET1649042.
- (19) Estévez-Landazábal L-L, Barajas–Solano AF, Barajas–Ferreira C, Kafarov V. Improvement of lipid productivity on *Chlorella vulgaris* using waste glycerol and sodium acetate. *CT&F - Ciencia, Tecnol y Futur.* 2013;5(2):113–26.
- (20) Andersen RA, editor. Recipes for Freshwater and Seawater Media. In: *Algal Culturing Techniques*. 1st ed. Elsevier Academic Press; 2005. p. 429–538.

- (21) TIBCO Statistica™. TIBCO Software Inc; 2004. Available from: <https://www.tibco.com/sites/tibco/files/resources/ds-statistica-tech-brief-big-data-analytics-final.pdf>.
- (22) Moheimani NR, Borowitzka MA, Isdepsky A, Sing SF. Standard Methods for Measuring Growth of Algae and Their Composition. *Algae for Biofuels and Energy*. 2013;5:265–84. Doi: 10.1007/978-94-007-5479-9_16.
- (23) Příbyl P, Pilný J, Cepák V, Kaštánek P. The role of light and nitrogen in growth and carotenoid accumulation in *Scenedesmus* sp. *Algal Res*. 2016;16:69–75. Doi: 10.1016/j.algal.2016.02.028.
- (24) González-Delgado AD, Barajas-Solano AF, Ardila-Álvarez AM. Biomass and protein production of *Chlorella vulgaris* Beyerinck (Chlorellales: Chlorellaceae) via the design of selective culture media. *Cienc y Tecnol Agropecu*. 2017;18(3):451–61. Doi: 10.21930/rcta.vol18_num3_art:736.
- (25) Bhosale P. Environmental and cultural stimulants in the production of carotenoids from microorganisms. *Appl Microbiol Biotechnol*. 2004;63(4):351–361. Doi: 10.1007/s00253-003-1441-1.
- (26) Pirastru L, Darwish M, Chu FL, Perreault F, Sirois L, Sleno L, et al. Carotenoid production and change of photosynthetic functions in *Scenedesmus* sp. exposed to nitrogen limitation and acetate treatment. *J Appl Phycol*. 2012;24(1):117–124. Doi: 10.1007/s10811-011-9657-4.
- (27) Bishop NI. The β , ϵ -carotenoid, lutein, is specifically required for the formation of the oligomeric forms of the light harvesting complex in the green alga, *Scenedesmus obliquus*. *J Photochem Photobiol B Biol*. 1996;36(3):279–83. Doi: 10.1016/S1011-1344(96)07381-2.
- (28) Song M, Pei H. The growth and lipid accumulation of *Scenedesmus quadricauda* during batch mixotrophic/heterotrophic cultivation using xylose as a carbon source. *Bioresour technol*. 2018;263:525–531. Doi: 10.1016/j.biortech.2018.05.020.
- (29) Minhas AK, Hodgson P, Barrow CJ, Sashidhar B, Adholeya A. The isolation and identification of new microalgal strains producing oil and carotenoid simultaneously with biofuel potential. *Bioresour Technol*. 2016;211:556–65. Doi: 10.1016/j.biortech.2016.03.121.
- (30) Qin S, Liu G-X, Hu Z-Y. The accumulation and metabolism of astaxanthin in *Scenedesmus obliquus* (Chlorophyceae). *Process Biochem*. 2008;43(8):795–802. Doi: 10.1016/j.procbio.2008.03.010.
- (31) Kozlova TA, Hardy BP, Krishna P, Levin DB. Effect of phytohormones on growth and accumulation of pigments and fatty acids in the microalgae *Scenedesmus quadricauda*. *Algal Res*. 2017;27:325–34. Doi: 10.1016/j.algal.2017.09.020.
- (32) Patil L, Kaliwal B. Effect of CO₂ Concentration on Growth and Biochemical Composition of Newly Isolated Indigenous Microalga *Scenedesmus bajacalifornicus* BBKLP-07. *Appl Biochem Biotechnol*. 2017;182(1):335–348. Doi: 10.1007/s12010-016-2330-2.

- (33) Příbyl P, Cepák V, Kaštánek P, Zachleder V. Elevated production of carotenoids by a new isolate of *Scenedesmus* sp. Algal Res. 2015;11:22–7. Doi: 10.1016/j.algal.2015.05.020. 2017;55(10):702–10. Available from: <http://nopr.niscair.res.in/handle/123456789/42843>.
- (34) Anusree V, Sujitha B, Anand J, Arumugam M. Dissolved inorganic carbonate sustain the growth, lipid and biomass yield of *Scenedesmus quadricauda* under nitrogen starved condition. Indian J Exp Biol. 2014;152:275–82. Doi: 10.1016/j.biortech.2013.11.031.
- (35) Ho SH, Chan MC, Liu CC, Chen CY, Lee WL, Lee DJ, et al. Enhancing lutein productivity of an indigenous microalga *Scenedesmus obliquus* FSP-3 using light-related strategies. Bioresour Technol. 2014;152:275–82. Doi: 10.1016/j.biortech.2013.11.031.



Este trabajo está licenciado bajo una [Licencia Internacional Creative Commons Reconocimiento-
NoComercial-CompartirIgual 4.0](https://creativecommons.org/licenses/by-nc-sa/4.0/)