
MORPHOLOGIC AND MOLECULAR CHARACTERIZATION OF SPECIES OF *Myrciaria* spp

PEREIRA, Márcio¹
OLIVEIRA, Antonio Luis de²
SENA, Janete Aparecida Desidério³
COSTA, Juliana Regina Vieira da⁴
ALMEIDA, Marcílio de
GONÇALVES, Antonio Natal

Recebido em: 2009-01-26

Aprovado em: 2009-02-28

ISSUE DOI: 10.3738/1982.2278.185

SUMMARY: The jaboticaba tree is considered one of the most typical Brazilian fruit tree, however, there are a little of studies of this plant in the literature and even in the specialized literature, there are controversies about its classification many. The present work makes some comparisons between jaboticaba species, using morphologic markers (organography) and molecular markers (RAPD) technique. The morphologic characteristics of the plants, used as morphologic markers, were compared with specimens present on herbaria from São Paulo and Minas Gerais states and through the revision of specialized literature. Molecular differences between the species were identified by molecular in Piracicaba, Jaboticabal and Ituverava cities, São Paulo, Brasil. Morphologic and molecular differences between the studied plants were identified and arranged in four groups, the identified species were: *Myrciaria cauliflora* (Mart.) O. Berg, *Myrciaria coronata* Mattos, *Myrciaria jaboticaba* (Vell.) O. Berg., *Myrciaria phytrantha* (Kiaersk.) Mattos. The technique of molecular markers with the technique of morphologic markers (organography) showed to be an important tool in the identification of *jaboticaba* tree species.

Keywords: Myrciaria. Biological identification. Morphologic markers. Molecular markers (RAPD)

CARACTERIZAÇÃO MORFOLÓGICA E MOLECULAR DE ESPÉCIES DE *Myrciaria* spp

RESUMO: A Jaboticabeira é considerada como uma das fruteiras mais típicas do Brasil, contudo há poucos estudos desta planta na literatura e mesmo na literatura especializada, existem muitas controvérsias sobre sua classificação. Este trabalho faz comparações entre as espécies de Jaboticabeiras, usando as técnicas de marcadores morfológicos (Organografia) e moleculares (RAPD). As características morfológicas das plantas, usadas como marcadores morfológicos, foram comparadas com espécimes presentes nos herbários dos Estados de São Paulo e Minas Gerais e através da revisão de literatura especializada. As diferenças moleculares entre as espécies foi determinada através do uso de marcadores moleculares (RAPD). O experimento foi realizado nas cidades de Piracicaba, Jaboticabal e Ituverava, São

¹ FAFRAM – Faculdade “Dr. Francisco Maeda” de Ituverava Departamento de Horticultura Rod. Jerônimo Nunes Macedo, Km 01 – 14500-000 – Ituverava, SP – Brasil. UNESP/FCVA – Depto. de Biologia Aplicada à Agropecuária, 14884-900 – Jaboticabal, SP – Brasil.

² UNESP/FCVA – Depto. de Biologia Aplicada à Agropecuária, 14884-900 – Jaboticabal, SP – Brasil.

³ USP/ESALQ – Depto. de Ciências Biológicas, C.P. 9, 13418-900 – Piracicaba, SP – Brasil.

⁴ USP/ESALQ – Depto. Ciências Florestais

* Corresponding author: fafram@feituverava.com.br

Paulo, Brasil. Diferenças morfológicas e moleculares entre as plantas estudadas foram identificadas e as mesmas foram agrupadas em quatro grupos distintos, as espécies identificadas foram: *Myrciaria cauliflora* (Mart.) O. Berg., *Myrciaria coronata* Mattos, *Myrciaria jaboticaba* (Vell.) O. Berg., *Myrciaria phytrantha* (Kiaersk.) Mattos. A técnica de marcadores moleculares aliada à técnica de marcadores morfológicos (organografia), mostrou ser uma ferramenta importante na identificação de espécies de jaboticabeiras.

Palavras-chave: *Myrciaria*. Jaboticabeiras. Marcadores morfológicos. Marcadores moleculares (RAPD)

INTRODUCTION

According to Mattos (1983) the jaboticaba trees haven't always been extensively studied under the taxonomic point of view. There is a lot of confusion concerning the popular names of their fruit. Many times its vulgar name is applied to different species and even different genus, depending on the area where they are found. In that sense the vegetative sprouts play an important part in the identification of the plants, which may happen in several times of the year, but those that appear in the end of the winter and beginning of the spring are the most intense. The new foliage appears in the periphery of the plants and the color varies from light green to purplish, according to the species (DONADIO, 2000). This characteristic with the floral structures are used as a parameter to identify this species.

The identification of species and varieties has been traditionally based on the description of morphologic characteristics (external and internal) of the plants. Without a doubt that description is more restricted, being sometimes necessary the use of controlled conditions to minimize the natural variability. A great progress was the development of genetic techniques based DNA markers.

Nowadays, with the use of efficient techniques to genomic analysis, statistical methods to evaluate the obtained data and efficient software, the study of individuals' genetic identification and populations a good stage has been reached.

Individual organisms differ in its sequences of DNA, and that variation can be considered at the level of individual genes (genic) or genotypes (genotypical). The genetic variation that occurs in time and in space is influenced by the biology and the relationships that the individual has with the environment, influencing in its survival strategy. Measuring the genetic variation and applying genetic models of populations one can infer on the biology of organisms. The processes that affect the individuals, affect the population, influencing in the species and interfering in the taxonomic hierarchy (SUNNUCKS, 2000) .

According to the same author, the adapted choice of the technique for genetics-

analysis is vital for the success in the study of the individual's molecular identification. For that study it can be used the genotypical series for study of multiple locus, being not very accurate, they visualize many anomalous genes at the same time, but they are technically convenient. The gene variation is used for the study of unique locus (simple), they produce more consistent data for a more precise analysis, promoting better comparability. The great usefulness of molecular techniques allows to observe differences with great resolution level, finding differences that would be identical when phenotypic characteristics are used. These differences are obtained independently from the studied part or of the conditions in which the plant was cultivated. To observe the differences of DNA in living beings it is necessary a group of appropriate techniques. Of the existent ones, the polymerase chain reaction (PCR) and its variants are the most important.

The method of amplification of fragments of DNA, RAPD, brought a true democratization of the analysis of molecular polymorphism, when allowing the accomplishment of studies of genetic analysis in species previously not contemplated (WILLIAMS et al., 1993).

For the exposed, and in an attempt of contributing to the solution of the problem of jaboticaba tree identification, the present work has the objective to identify jaboticaba tree species using techniques of morphologic markers (organography) as well as molecular markers.

MATERIAL AND METHODS

To collect the botanical material used in the identification of the jaboticaba species (*Myrciaria* spp.), the procedure recommended by the Institute of Botany of São Paulo (Série Documentos, 1989) was used.

Thus, the collection of fertile branches of the plant was made at a height of 2.5 meters from the level of the soil in the terminal branches, always in the faces east and west.

Due to its quite fine nature, the flowers of the jaboticabeira (jaboticaba tree) were conserved in liquid of Hammarlund (copper sulfate in saturated solution + formaldehyde at 40% = formalin + distilled water in the proportion of 3:0,1:1). The present fruits in some plants were collected and conserved in solution of copper sulfate in aqueous solution at 5%, sulfurous acid in aqueous solution at 5-6% and glycerin

(method of Drummond).

For each collected material, a record was filled with additional data as texture of the peel, load of the plant, size of the peduncle and of the fruit, etc, according to the orientation of the National Council of Scientific and Technological Development, this information was later on transcribed onto an herbarium record.

After the drying out the flowers in absorbent paper, they were pressed and dehydrated in stove at the Laboratory of Botany at Faculdade " Dr. Francisco Maeda " (FAFRAM). The assembly of the dried herbarium specimens was made by placing the flowers in paper bags and labels were placed in the inferior right corner of the assembly cardboard. In the left superior corner, diametrically opposite, an envelope was fastened to contain the parts which eventually fell down from the material during the drying out procedure as well as those necessarily taken for the study of the vegetable.

The collected material was identified with codes:

· The materials from the orchard of the Section of Horticulture of the Department of Vegetable Production of ESALQ-USP:

Codes: Pira1, Pira2, Pira3, E01, E02, E03, E04, P01, and P02.

· The collected materials from the orchard of the Department of Vegetable Production of ESALQ-USP, located in the Section of Engineering:

Codes: P1A, P2A, P3A, P4A, P5A, P6A, P7A, P8A, P9A, and P10A.

· The collected materials from the Section of Fruit-culture at The College " Dr. Francisco Maeda " of Ituverava-SP:

Codes: 1I, 2I, 3I, 4I, 5I, 6I, 7I, 8I, 9I, 10I, 11I and 12I.

REVISION OF HERBARIA

Considering its location and its relationship with the main areas of occurrence, the species of cultivated jaboticabeiras, the dried herbarium specimens of the collected material was compared with other specimens of the list of chosen herbaria and specialized literature.

The herbaria selected for the analysis of the dried herbarium specimens are described in Table 1.

TABLE 1- Herbaria used for revision of the botanical material

State	City	Sigla	Institution	Trustee
SP	São Paulo	SP	Instituto de Botânica de São Paulo – Secretaria de Estado do Meio Ambiente	Dr. ^a Inês Cordeiro
SP	São Paulo	SPB/SPF	Instituto de Biociências – Universidade de São Paulo	Dr. José Rubens Pirani
SP	Campinas	UEC	Universidade de Campinas	Dr. ^o Washington Marcondes Ferreira Neto
SP	Campinas	IAC	Instituto Agrônômico de Campinas	Dr. ^a Sigrid Jung Mendaçolli
SP	Rio Claro	HRCB	UNESP-Rio Claro	Dr. Marco Antônio Assis
SP	Piracicaba	ESA	ESALQ-USP/Piracicaba	Dr. Lindolfo Capellari Júnior
SP	Ituverava	FAFRAM	Faculdade “Dr. Francisco Maeda”	M.Sc. Márcio Pereira
MG	Uberlândia	UFU	Universidade Federal de Uberlândia	Dr. ^o Jimi Nacki Nakagima
MG	Belo Horizonte	UFMG	Universidade Federal de Minas Gerais	Dr. Júlio Lombardi

MOLECULAR CHARACTERIZATION

Of the same plants where the vegetative and reproductive material was collected for morphologic analysis, leaves of new sprouts were collected (not totally expanded), what resulted in approximately 180 mg of fresh tissue. This material was placed in small paper bags, properly identified with the code of the plants and stored in containers with liquid nitrogen at - 86°C and, soon after transported until the laboratory of Genetics of Bacteria of the Department of Applied Biology to the Agriculture of UNESP/FCAV for the extraction of the DNA.

DNA EXTRACTION OF

The extraction of DNA of the leaves of the jaboticabeiras proceeded, at first, in the method described in Lodhi et al. (1994), modified by the staff of the Laboratory of Jaboticabal-SP.

DNA QUANTIFICATION AND QUALITY

The samples were quantified in spectrophotometer DU 640B (Beckman), diluted in the proportion of 2 μL of the solution stock of DNA in 98 μL of TE 10:1 (v/v).

The quantification using the spectrophotometer method allows to estimate the purity of the DNA through average of readings taken at 260 and 280 nm. Pure preparations of DNA have values for this coefficient in the interval from 1.8 to 2.0. Values below 1.8 indicate contamination of the nucleic acid with protein. To evaluate the amount of obtained total DNA it was used the pattern where an absorbency unit at 260 nm is equal to 50 mg of DNA per μL of solution. The final concentration used as work solution was of 10 ng $\cdot \mu\text{L}^{-1}$, necessary for the reactions of RAPD.

The quality analysis of the extracted DNA was carried out in electrophoresis of agarose gels at 0.8%, where it was applied 10 μL of DNA of the samples and 3 μL of charge buffer (Tris - HCL 0,1M, pH 6.8; blue of bromophenol 0.02%; glycerol 50%). To compare the pattern of the bands 8 μL of " 1Kb Plus DNA Ladder " of GIBCO/BRL was used. The buffer used in the prepare of electrophoresis and gel was Tris-borate-EDTA 1X (Tris 89 mM; H₃BO₃ mM; EDTA 2.5 mM, pH 8.2) containing 0.5 mg $\cdot \mu\text{L}^{-1}$ of ethyl bromide. The electrophoresis time was of approximately 2h in tension of 48V. The fragments of genomic DNA were visualized under light UV and documented in a photodocumentator model GEL DOC 2000 (BIO RAD).

DNA AMPLIFICATION

DNA samples were amplified in the Laboratory of Jaboticabal-SP.

The procedure for reactions of amplifications of the DNA, as well as the analysis of PCR with adopted primers, was the same as described by Williams et al. (1990). The primers used in this work came from the collection of the University of British Columbia - Nucleic Acid - Protein Service Unit (Canada), whose access numbers to the collection, as well as the respective sequences, are described in the Table 2.

TABLE 2 - Arbitrary sequences of the initiators used and respective

201. CTG GGG ATT T	226. GGG CCT CTA T
202. GAG CAC TTA C	227. CTA GAG GTC C
203. CAC GGC GAG T	228. GCT GGG CCG A
204. TTC GGG CCG T	230. CGT CGC CCA T
205. CGG TTT GGA A	231. AGG GAG TTC C
206. GAG GAC GTC C	232. CGG TGA CAT C
208. ACG GCC GAC C	233. CTA TGC GCG C
209. TGC ACT GGA G	234. TCC ACG GAC G
212. GCT GCG TGA C	235. CTG AGG CAA A
213. CAG CGA ACT A	236. ATC GTA CGT G
218. CTC AGC CCA G	247. TAC CGA CGG A
219. GTG ACC TCA G	248. GAG TAA GCG G
220. GTC GAT GTC G	279. AGA CAT TAG A
223. GAT CCA TTG C	286. CGG AGC CGG C
225. CGA CTC ACA G	296. CCG CTG GGA G

The amplification reactions were made in a volume of reaction of 20 μL containing: 30 ng of the DNA to be amplified; solution of dNTPs 2.0 mM; 1mM MgCl_2 ; solution lid (1X); 15 ng of each one of the initiators; 1.0 U Taq DNA polymerase and distilled water degree Milli Q (previously sterilized).

The amplification reactions were accomplished using a thermocycler MJ Research, model PTC 100, equipped with circuit "Hot Bonnet". The program adopted for this stage of analysis was set up as described: 4 minutes at 92°C and, later 48 cycles of 1 minute were accomplished at 92°C; 1 minute and 30 seconds at 37°C; 1 minute and 30 seconds at 72°C; and, in the end, 5 minutes at 72°C.

The amplified samples were analyzed in agarose gel 1.5% (p/p), using lid TEB 1X (89 mM of Tris; 2.5 mM of EDTA and 89 mM of Boric Acid, pH 8.3) containing 0.5 $\mu\text{g.L}^{-1}$ of ethyl bromate.

The fragments amplified by PCR and separated by electrophoresis were compared with those of the DNA of known molecular weight ("1 Kb ladder") and it was verified the absence or the presence of electrophoretic migration of the same ones as well as their distances.

RESULTS AND DISCUSSION

MORPHOLOGIC CHARACTERIZATION

By means of the comparison of the collected specimens, both the description of

the specialized bibliography, Mattos (1983), and the specimens conserved in the visited herbaria, it was possible to group the studied plants in four species, allowing to verify from the taxonomic point of view that the species of jaboticabeiras are not very studied, because from all the consulted collections just two species were found in the dried herbarium specimens, *Myrciaria jaboticaba* and *Myrciaria cauliflora*, being nonexistent dried herbarium specimens of the other species (*M. coronata*, *M. phitrantha*, *M. piruviana*, *M. oblongata*, *M. spiritosantensis*, *M. grandiflora*, and *M. aureana*) in the herbaria visited.

Through that research it can also be verified that the collections present vast material of the gender *Myrciaria*, however the species of described Jaboticabeiras are restricted to 2 groups in the herbaria of the State of São Paulo, showing that other tools should be used to aid in the identification of those species.

Of the thirty-one plants studied, the one of code Pira 1, initially identified as *Myrciaria* by the field employees of Piracicaba-SP, it was later identified how the genus *Psidium*. It is an interesting fact that justifies the difficulty and confusion concerning the vulgar names given to the fruits of the species belonging to the gender *Myrciaria*. Even some herbaria bring in their older collections of *Mirciariae*, some jaboticabeiras with different classifications, mainly between the species *Myrciaria cauliflora* and *Myrciaria jaboticaba*.

In literature itself this confusion can be noticed. Mendonça (2000), in a work developed in the jaboticabeiras collection belonging of Viçosa-MG, classified *Myrciaria cauliflora* (Mart.) O. Berg. as *Cultivar Açú* and *Myrciaria jaboticaba* (Vell.) O. Berg as *Cultivar Sabará*. Pio Corrêa (1984), in the classic dictionary of the useful plants of Brazil, classified *Myrciaria jaboticaba* as *Cultivar Açú* and *Myrciaria trunciflora* O. Berg. as belonging to the species *Cultivar Sabará*. Still in the same work the species *Myrciaria cauliflora* (included in the gender *Eugênia*) is treated like “common” or “true” Jaboticaba, having as synonymous jaboticaba of Sabará or jaboticaba of São Paulo.

In the Table 3, the 31 studied plants and their respective identifications are presented, accomplished through the comparison with the specimens of the visited herbaria visited and of the specialized literature.

TABLE 3. Relationship of the species of identified jaboticabeiras, through the comparison of present dried herbarium specimens in the visited herbaria and in the consultation of specialized bibliography

Code	Species
Pira 1	<i>Psidium</i> spp
Pira 2	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
Pira 3	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
1I	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
2I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
3I	<i>Myrciaria coronata</i> Mattos
4I	<i>Myrciaria coronata</i> Mattos
5I	<i>Myrciaria coronata</i> Mattos
6I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
7I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
8I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
9I	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
10I	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
11I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
12I	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
P01	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P02	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P1A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P2A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P3A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P4A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P5A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P6A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P7A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P8A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P9A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
P10A	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
E01	<i>Myrciaria phitrantha</i> (Kiaersk.) Mattos
E02	<i>Myrciaria cauliflora</i> (Mart.) O. Berg
E03	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg
E04	<i>Myrciaria jaboticaba</i> (Vell.) O. Berg

MORPHOLOGIC IDENTIFICATION

The specimens belonging to the species *Myrciaria jaboticaba* (Vell.) O. Berg (Figure 1), presented the bicarpel ovary, infra-axillary placentation, pilose on the base, style surpassing the stamens, captured stigma, numerous stamens, flat terminal branches, green superior surface of leaves and lighter green coloring on the inferior surface, pentamerous actinomorpe corolla, and globular fruits with diameter of 1.6 to 2.2 cm, smooth and black when ripe, and from one to four seeds in its interior. These characteristics are in agreement with the descriptions of Mattos (1983).

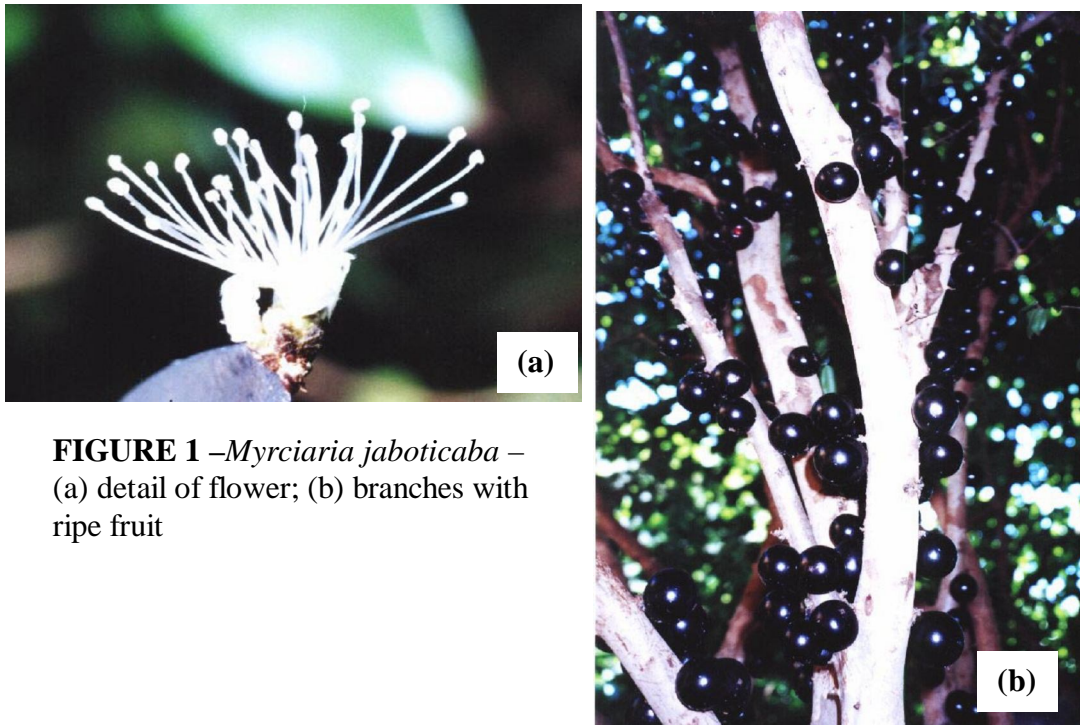


FIGURE 1 –*Myrciaria jaboticaba* – (a) detail of flower; (b) branches with ripe fruit

The species *Myrciaria cauliflora* (Mart.) O. Berg (Figure 2), presented bicarpel ovary, infra-axillary placentation, glabrous, style of 6 mm in length, peltate stigma, flat terminal branches and leaves with central nervure slightly printed in the superior surface and salient epidermis in to inferior surface, pentamerous actinomorpe corolla, glabrous floral button, globular fruit of 2.2 to 2.8 cm of length and 2.2 to 2.9 cm of diameter.



FIGURE 2 - *Myrciaria cauliflora* – Branches with ripe fruit

The *Myrciaria phitrantha* (Kiaersk.) Mattos (Figure 3), presents bicarpel ovary, infra-axillary placentation, long and glabrous style, pentamerous actinomorpe corolla, big fruit (from 3 to 4 cm in diameter) with the “neck” and big leaves as main characteristic (more than twice the size of the other studied species) and pendants.

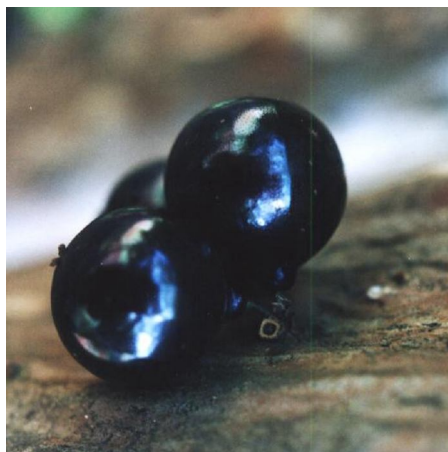


FIGURE 3 - *Myrciaria phitrantha* (Kiaersk.) Mattos - Detail of fruits

The specimens of the species *Myrciaria coronata* presented a captured stigma, bicarpel sericeous ovary, with axillary placentation, flat and grizzly terminal branches, leaf with main nervure printed in the superior surface and salient in the inferior surface, globular fruit and whitish contour of the disk of the apex.

MOLECULAR CHARACTERIZATION

In the analysis of markers RAPD, the 11 selected primers generated 45 polymorphic bands. The more polymorphic primers were the ones of number 203 and 226 (Figures 4 and 5), being the ones that also presented better image in the agarose gels. According to Colombo (1998), 10 to 30 primers, generating 50 to 100 polymorphic bands are enough to estimate genetic relationships intra and interspecies.

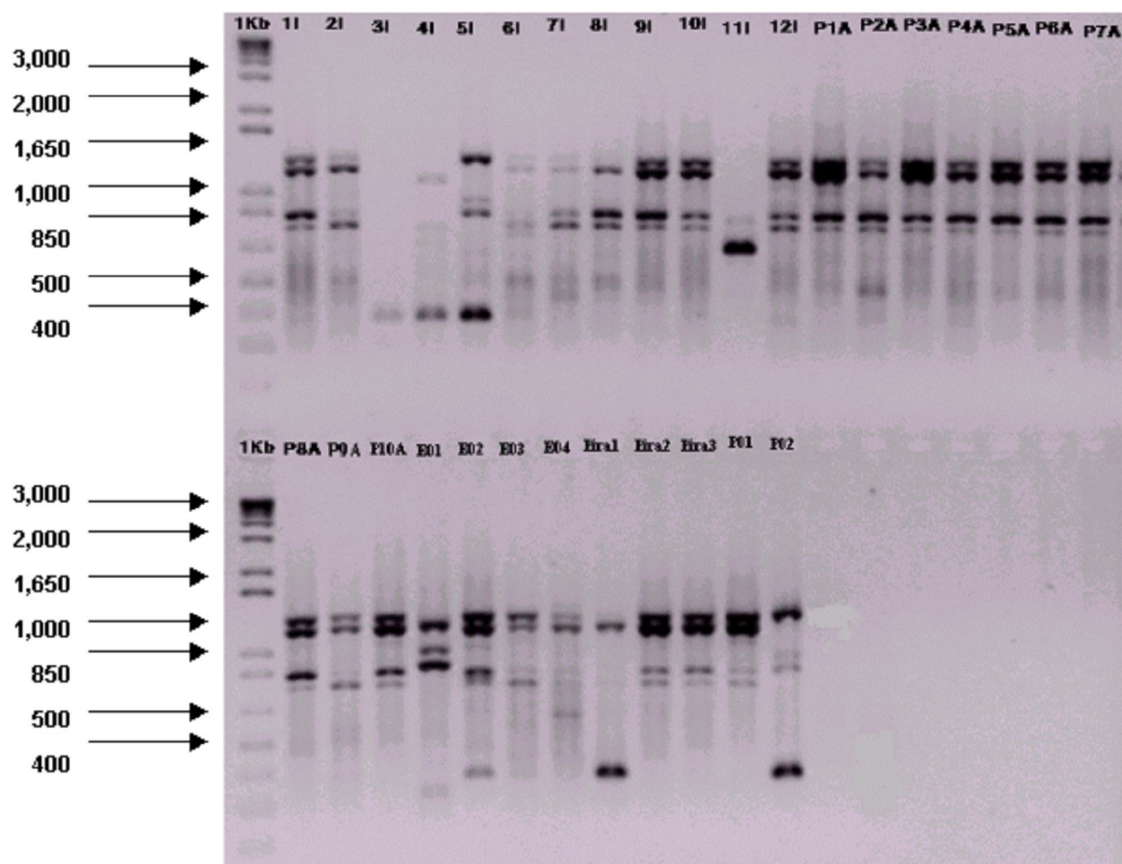


FIGURE 4 - Electropherogram of the plants amplified with primer of number 203, being: MM = " 1Kb Plus DNA Ladder ".

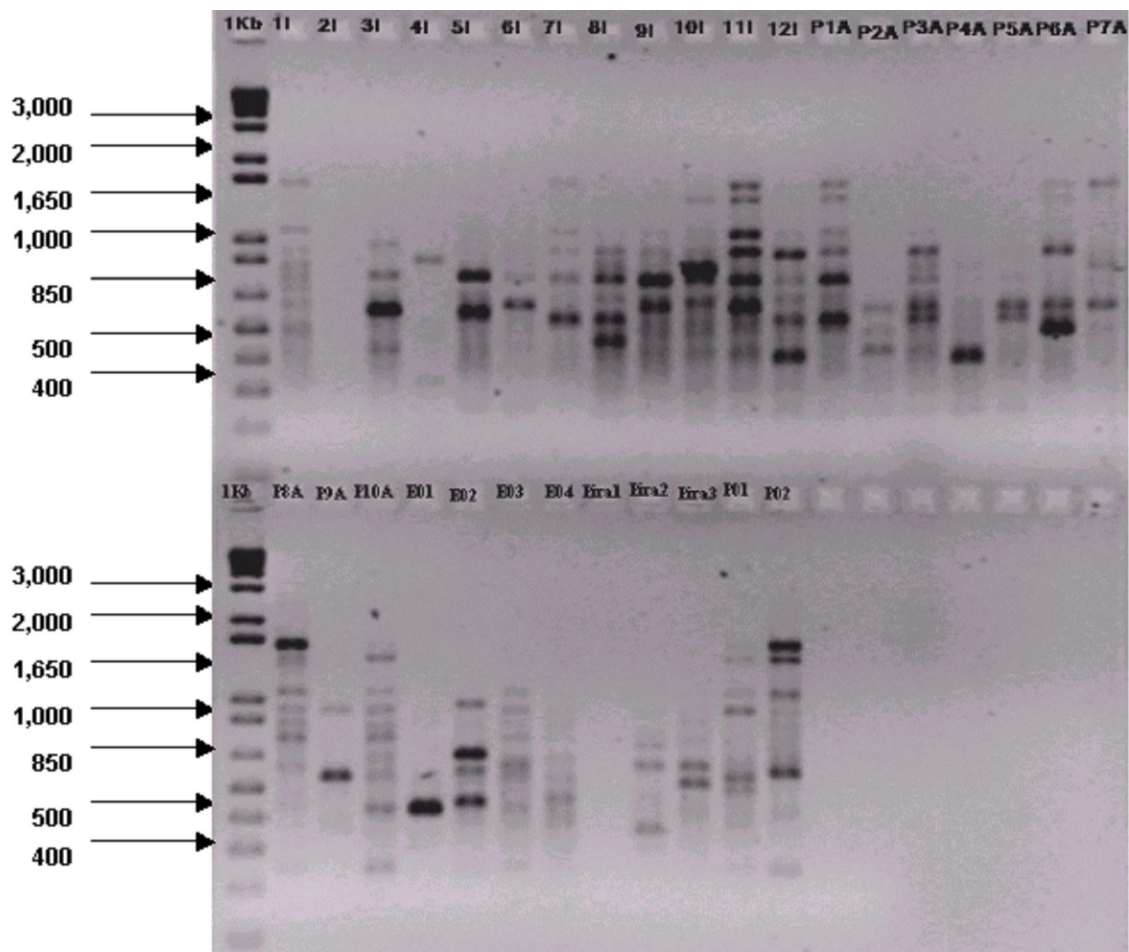


FIGURE 5 - Electropherogram of the plants amplified with primer of number 226, being: MM = " 1Kb Plus DNA Ladder ".

Ashburner et al. (1996) observed that the markers of type " RAPD " were capable for differentiating the coconut populations (*Coco nucifera* L.) coming from the Islands Rennel of those originating from of the continent. Those facts prove that the methodology of the molecular markers applied in the individuals' identification can be a tool to aid in the individuals' grouping for their genetic similarity.

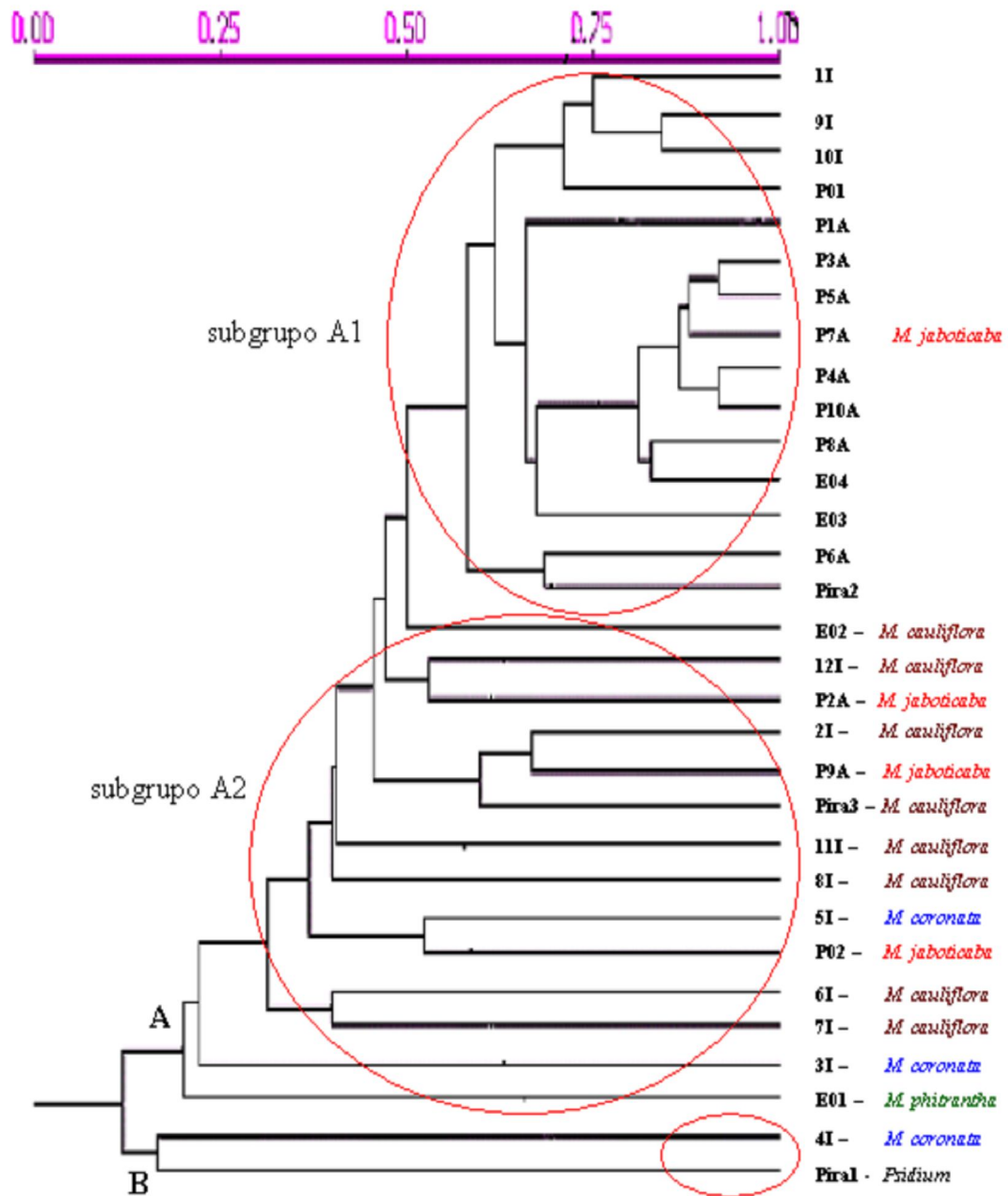


FIGURE 6 – Phylogenetic diagram of genetic similarity among jaboticabeiras individuals, obtained through the coefficient of Jaccard.

In Figure 6 the formation of two main groups is observed: A and B. In group B represents just two plants, whose genetic distance is approximately of 80%. The plant of code " 4I " is morphologically classified, whose species is *Myrciaria coronata* Mattos and the plant of code " Pira 1 ", after having been collected like a *Myrciaria* was

identified as belonging to the family of *Myrtaceae*, but of the gender *Psidium* spp.), confirming, this way, such divergence among them.

In group A are distributed other species (*M. jaboticaba*, *M. cauliflora*, and *M. phitrantha*). Inside this same group, the formation of a sub group is observed (A1), where are the plants of the species *M. jaboticaba*, with a similarity degree varying from 60% to 90%.

Another sub group (A2) was formed with species of *M. cauliflora*, *M. coronata*, and *M. jaboticaba*, with degrees of non similarity from 80% to 30%, however, with very close genetic characteristics.

The species *M. phitrantha* Mattos, presents a genetic divergence of 80% to the other species. It can be observed in the phylogeny diagram in an isolated branch of the other groups.

In this context, it is concluded that technical RAPD-PCR, didn't allow the grouping of the plants at the species level, but it allowed to analyze, by means of generated phylogeny diagram, the degrees of genetic similarity among them, observing that, the species *M. phitrantha* was farther distant to the other species, as well as the plant identified as *Psidium* spp., initially collected like *Myrciaria*, which also had high non similarity degree, checking the reliability of this technique.

At first, if one observes the phylogeny diagram, one may infer that the markers RAPD didn't allow the grouping of the plants, according to sub group A2. However, for the accomplished study, there aren't subsidies to affirm such fact, because the studied material was compared with older collections of *Myrciariae*, in which there are different classifications, mainly between *M. cauliflora* and *M. jaboticaba*, proving the analysis of the sub group A2, in which most of the contained plants belongs the species *M. cauliflora* and, in which are present the plants P2A, P9A, and P02 morphologically classified as *M. jaboticaba*.

Concerning the species *M. coronata* (3I, 4I, and 5I), whose morphologic classification was just based in the Literature (Mattos, 1983), without dried herbarium specimens of this species in the visited herbaria, a more meticulous research is suggested, because those plants can present a different classification.

It is evident in phylogeny diagram that the largest genetic distances involve the plants of sub group A2, confirming the difficulty of a current morphologic classification.

Nowadays, with the use of molecular tools, it could be suggested a revision of the classification of the gender *Myrciaria*, as well as of the dried herbarium specimens deposited in the visited herbaria.

REFERENCES

ASHBURNER, G.R.; THOMPSON, W.K.; HALLORAN, G.M. RAPD analysis of south pacific coconut palm populations. **Crop Science**, v.37, p.992-997, 1997.

COLOMBO, C. Genetic diversity characterization of casava cultivars (*Manihot esculenta* Crantz) with RAPD markers. **Genetics and Molecular Biology**, Ribeirão Preto, v.21, 1, p.105-113, 1998

DONADIO, L. C. **Jaboticaba (*Myrciaria jaboticaba* (Vell.) Berg)**. Jaboticabal: FUNEP, 2000, 55p.

FERREIRA, M.E; GRATTAPAGLIA, D. **Introdução ao uso de marcadores em análise genética**. 2.ed. Brasília: EMBRAPA-CENARGEM, 1995. p.220 (Documento, 20)

LODHI, M.A.; YE, N.G.; WEEDEN, N.F. A simple and efficient method for DNA extraction from grapevine cultivars and *Vitis* species. **Plant Molecular Biology Reporter** v. 12: p6-13, 1994.

MATTOS, J.L.R. **Fruteiras nativas do Brasil: jaboticabeiras**. Porto Alegre: Nobel, 1983. 92p.

MENDONÇA, R.M.N. **Maturação, secagem e armazenamento de sementes e propagação vegetativa de Jaboticabeiras (*Myrciaria* sp)**. Viçosa, 2000. 136p. (Tese de Doutorado). Universidade Federal de Viçosa.

PIO CORRÊA, M. **Dicionário das plantas úteis do Brasil e das exóticas cultivadas**. Rio de Janeiro: IBDF, 1984, v.4, 450p.

SÉRIE DOCUMENTOS. Técnicas de coleta, preservação e herborização de material botânico. **Instituto de Botânica**. São Paulo, 1989. 62p.

SUNNUCKS P. Efficient genetic markers for population biology. **TREE**, v 15, n.5, 2000, p.199-203.

WILLIAMS, J.G.K., KUBELIK, A.R.; LIVAK, K.J J.A. DNA polymorfims amplified by arbitrary primers useful as genetic markers. **Nucleic Acids Research**, v.18, p.6531-6535, 1990.

WILLIAMS, J.G.K.; RAFALSKI, J.A. ; TINGEY, S.V. 1993. Genetic analyss using RAPD markers. **Methods in Enzymology**, v.218, p.704-740, 1993.