# Role of *Bradyrhizobium japonicum* induced by genistein on soybean stressed by water deficit

M. C. Nápoles<sup>1\*</sup>, E. Guevara<sup>2</sup>, F. Montero<sup>3</sup>, A. Rossi<sup>3</sup> and A. Ferreira<sup>3</sup>

<sup>1</sup> Instituto Nacional de Ciencias Agrícolas (INCA). Carretera a Tapaste, Km 3,5, San José de las Lajas, La Habana. Cuba.
<sup>2</sup> Instituto Nacional de Tecnología Agropecuaria (INTA). Ruta 32, Km 4,5. Camino a Salto, Pergamino. Argentina.
<sup>3</sup> Rizobacter Argentina S.A. Ruta 32, Km 1,5, Parque Industrial, Pergamino. Argentina.

#### Abstract

The soybean (*Glycine max* (L.) Merr.) is a crop mainly grown under rain fed conditions although irrigation is increasingly being used. Water deficiency is the main factor limiting seed production. The symbiosis process is also negatively affected by water stress. The isoflavone genistein have been recognized as a powerful inducer of Nod factors production by *Bradyrhizobium* and its addition to inocula has been shown to increase nodule number and promote soybean nitrogen (N) fixation at low temperatures. This study looks for answers about the possible role of genistein in countering the stress on nodulation produced by water deficit in soybeans. *Bradyrhizobium japonicum* SEMIA 5079 was grown in culture media induced or not induced with genistein. Inocula were applied to plants growing at different moisture levels. The effect of the treatments on nodulation and N content was evaluated. An improved response to drought stress was observed when the bacteria were grown in presence of genistein as a Nod factors inducer. Nodulation values under moisture stress differed from 8.9 nodules plant<sup>-1</sup> with genistein at 10  $\mu$ M to 1.8 nodules plant<sup>-1</sup> when no inducer was used. Genistein reduced the negative effect on nodulation caused by water deficiency.

Additional key words: humidity, inoculants, nitrogen fixation, nodulation, soil.

#### Resumen

#### Papel de Bradyrhizobium japonicum inducido por genisteína en soja estresada por déficit hídrico

El cultivo de la soja (*Glycine max* (L.) Merr.) se realiza fundamentalmente bajo condiciones de lluvia, pero la irrigación, específicamente el riego suplementado, es cada vez más utilizado. El proceso de simbiosis también se daña con el estrés hídrico. La isoflavona genisteína ha sido reconocida como un potente inductor de la producción de factores de nodulación por *Bradyrhizobium* y su adición a los inoculantes ha mostrado incrementos en el número de nódulos y la fijación del nitrógeno en soja a bajas temperaturas. Este estudio busca respuestas sobre el posible papel de la genisteína frente al estrés en la nodulación producido por el déficit de agua en plantas de soja. Se cultivó *Bradyrhizobium japonicum* SEMIA 5079 en medios conteniendo genisteína o no como inductor, y se inoculó posteriormente en plantas sometidas a diferentes niveles de humedad. Se evaluó el efecto sobre la nodulación y el contenido de nitrógeno en los diferentes tratamientos. Se obtuvo una mejor respuesta a la sequía cuando las bacterias fueron multiplicadas en presencia de genisteína como inductor de los genes de nodulación. Los valores de nodulación en condiciones de estrés fueron tan diferentes como 8,9 nódulos por planta con el empleo de genisteína 10  $\mu$ M con respecto a 1,8 nódulos por planta cuando no se utilizó ningún inductor. Se detectó un efecto importante de la genisteína en disminuir la caída de la nodulación por la deficiencia de agua.

Palabras clave adicionales: fijación de nitrógeno, humedad, inoculantes, nodulación, suelo.

Abbreviations used: DAS (days after sowing), LCO (lipochitooligosaccharides), LSD (least significant difference), *nod* (nodulation genes), RWC (relative water content), YMA (yeast mannitol agar).

<sup>\*</sup> Corresponding author: mariacaridad.napoles@gmail.com; tere@inca.edu.cu Received: 14-11-07. Accepted: 29-06-09.

# Introduction

Soybean (*Glycine max* (L.) Merr.) is one of the most important world crops. It is mainly cultivated for oil and protein and is a major food and feed source. As with other grain legumes, soybean is very sensitive to drought stress which leads to reduced yield and seed quality (Bosniols *et al.*, 1986; Frederick *et al.*, 2001; Purcell *et al.*, 2004). Negative effects of water stress on growth, photosynthesis, and photoassimilate translocation in soybean were demonstrated by Ohashi *et al.* (2000) and Fulai *et al.* (2004).

Because of their ability, as a legume, to interact with rhizobia, soybean forms root nodules where the bacteria fix atmospheric nitrogen (N) to ammonia, the substrate for protein biosynthesis (Gresshoff, 1993; Morgan *et al.*, 2005). The rhizobia and the plant cooperate to develop a symbiosis controlled by both partners. However, environmental conditions such as low temperature, high salinity, low pH and drought can negatively affect the rhizobia–legume symbiosis.

Drought stress is a major factor affecting symbiosis and leads to decreased nodule formation, reduced nodule size and N fixation (Serraj et al., 1999; King and Purcell, 2001; Serraj, 2003; Streeter, 2003; Tajima et al., 2004). Several mechanisms have been reported to be involved in the physiological response, carbon (C) shortage and nodule C metabolism, oxygen limitation, and feedback regulation by the accumulation of N fixation products, which results in poor nodulation and reduced amounts of fixed N (Zahran, 1999; Kurdalai et al., 2002; Serraj, 2003). Modifications in the activity of key nodule enzymes such as sucrose synthase and isocitrate dehydrogenase and in nodular malate content also occur. The decline in nodule water potential results in a cell redox imbalance (Marino et al., 2007). Considerable work has been conduced aimed at understanding the physiological mechanisms involved in the soybean-Bradyrhizobium symbiosis when subjected to drought stress. Ladrera et al. (2007) demonstrated that drought reduced C flux and N accumulation in nodules, but not in shoots.

In the molecular exchange between the plant and the microorganism, legume roots are known to exude flavonoid and isoflavonoid molecules that induce expression of *nod* (nodulation) genes in the rhizobia (Long, 2001). Genistein and daidzein are the major isoflavones in soybean exudates, responsible for *nod* gene induction in *Bradyrhizobium* (Kosslak *et al.*, 1990). As a result, the bacteria produce lipo-oligosaccharide Nod factors, whose precise structure determines the host range and specificity

of the association and induces several plant responses until complete nodule formation (Vijn *et al.*, 1993; Stokkermans and Peters, 1994; Heidstra *et al.*, 1997; Spaink, 2000; Geurts *et al.*, 2005). The structure and function of the lipo-chitooligosaccharides as essential signal for symbiosis with legumes have been extensively reported (Spaink *et al.*, 1998; Broughton *et al.*, 2000; Perret *et al.*, 2000; Gage, 2004; Jones *et al.*, 2007).

The addition of genistein to *Bradyrhizobium* inoculants has been shown to increase nodule number and dry matter (DM) and N fixation in soybean at low temperatures (Zhang and Smith, 1996, 1997). The biological activity of Nod factor, from *B. japonicum*, added to soybean roots under abiotic stress was also evaluated by Duzan *et al.* (2004). They found that addition of high levels of Nod factor was able to overcome the effects of low pH and temperature stress, but not salinity. The impact of a lipochitinoligosaccharide spray application on the physiology and productivity of "water stressed" soybean plants was evaluated by Atti *et al.* (2005).

This work studied the influence of three inoculants of *B. japonicum*, two of them induced with genistein, at different concentrations, on soybean nodulation at different water supply levels.

# Material and methods

#### **Bacterial culture**

Bradyrhizobium japonicum SEMIA 5079 was obtained from the FEPAGRO culture collection. The strain was selected because it is widely used in commercial *B. japonicum* inoculants in Argentina and Brazil. The culture for inoculation was grown at 28°C in 100 mL of Jap medium (Cozzi and Benintende, 1989) and the determination of cellular concentration in yeast mannitol agar (YMA, Vincent, 1970). The Jap medium was supplemented in two treatments by the addition of 5 and 10  $\mu$ M genistein respectively, to induce Nod factor synthesis. Flasks were shaken at 150 rpm for 60 hours on an incubator orbital shaker. Bacterial cultures (1.4-1.8 x 10<sup>9</sup> CFU mL<sup>-1</sup>) were used to inoculate seeds at rate of 25 mL 100 g<sup>-1</sup> seed.

#### **Plant growth conditions**

Soybean seed cv. DM 4600 RR was inoculated with the three inoculants, one non-induced culture and two induced with genistein. A fourth treatment was non-inoculated seed as a control.

A soil in which soybean had not been grown was used for the assay. Plants were cultivated in a growth chamber with 5000 lux of illumination, temperature of 25-30°C and 60-70% of relative humidity (Montero and Sagardoy, 2005) for 31 days.

#### **Application of water stress**

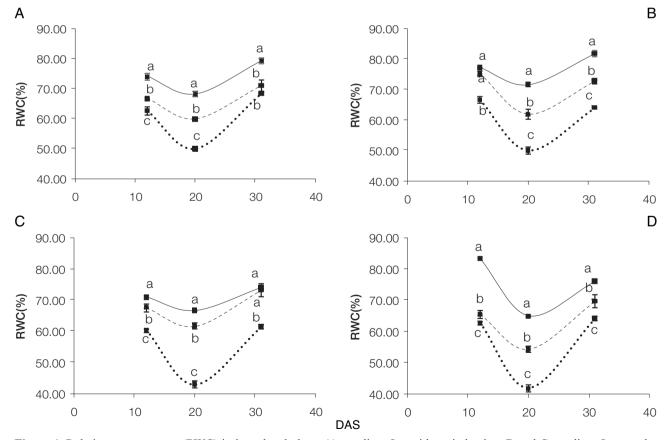
Plants were grown at three soil moisture levels: 30, 60 and 90% of field capacity. During the experiment water was supplied every two days to maintain these moisture levels. The relative water content (RWC) was measured at 12, 20 and 31 days after sowing (DAS), 48 hours after irrigation. The RWC was determined on three plants treatment<sup>-1</sup>, with three repetitions using the method of Pomper and Breen (1997).

#### Nodule biomass and plant nitrogen content

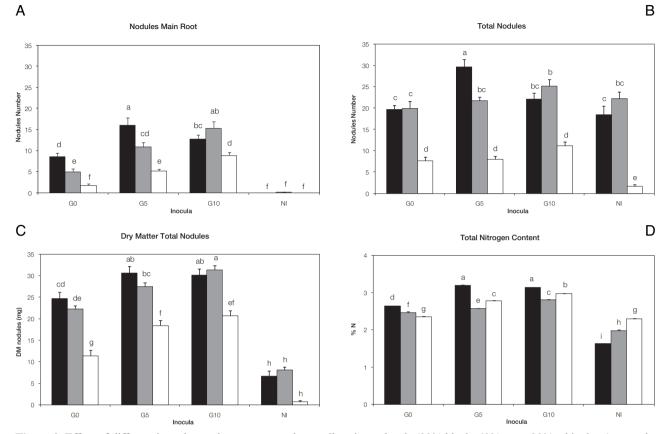
Plants were harvested at stage V3 (31 DAS); corresponding to the vegetative stage with at least three fully developed trifoliate leaves (Fehr *et al.*, 1971). The number and DM of nodules on main, secondary and total root, were determined. Shoot total N content was by the Kjeldahl method.

#### Experimental design and statistical analysis

The experiments were conducted twice. Results from the second assay are shown. The treatments were a completely randomized design with 20 replicates. The Info-Stat program (InfoStat, 2003) was used for two-ways analysis of variance. Where a significant treatment effect ( $p \le 0.05$ ) was observed by ANOVA, a least significant difference (LSD) test was conducted to determine differences among means at  $p \le 0.05$ .



**Figure 1.** Relative water content (RWC) in inoculated plants (A: medium Jap without induction, B and C: medium Jap supplemented with genistein at 5 and 10  $\mu$ M, respectively) and (D) non inoculated at three irrigation levels (line in points: 30% field capacity, suspense line: 60% field capacity, continuous line: 90% field capacity) at 12, 20 and 31 days after sowing (DAS). Vertical bars indicate ± standard error (SE). Treatments with different letters are significantly different, ANOVA (p < 0.05) LSD<sub>0.05</sub>.



**Figure 2.** Effect of different inocula on plants grown at three soil moisture levels (90% black, 60% grey, 30% white bars) on nodulation and nitrogen accumulation. G0: uninduced medium, G5 and G10: medium Jap supplemented with genistein at 5 and 10  $\mu$ M, respectively. NI: un-inoculated control. Treatments with different letters are significantly different, ANOVA (p < 0.05) LSD<sub>0.05</sub>.

### Results

#### Effect of irrigation level on relative water content

Evaluation of the RWC showed the effect of soil moisture level on the plant water relationship (Figure 1). The RWC fell considerably in all treatments after 12 days. After 20 days, plants had values of 40 to 50% RWC at 30% field capacity indicating moisture stress. However, there was some recovery from stress by the end of the assay. The RWC showed significant differences in treatments between each stress level, and over time 48 hours after irrigation.

# Effect of irrigation level on nodulation and plant nitrogen content

Generally, genistein increased nodulation and plant N content (Figure 2). The number of nodules on the main

root (Figure 2A), total nodule DM (Figure 2C) and N content (Figure 2D) of stressed plants (30 and 60% field capacity) were increased by use of genistein. There was no difference in total nodulation between genistein and uninduced inoculum for 10  $\mu$ M at 60% field capacity (Figure 2B), but with greater water availability (60 and 90% field capacity), the number and weight of nodules increased considerably in all treatments (Figure 2A, B and C).

The uninduced inoculum produced 8.6 nodules plant<sup>-1</sup> at 90% field capacity 5.0 at 60% and to 1.8 at the most severe stress level (Figure 2A). Nevertheless, the induced media decreased to 5.2 and 8.9 nodules plant<sup>-1</sup> in the most stressed conditions with 5 and 10  $\mu$ M of genistein, respectively. Similar results were observed for nodule DM and N content, which indicates a correlation between the effect on nodulation and its efficiency.

At 90% of field capacity, genistein at 5  $\mu$ M was enough to obtain the best results, while under stress (30 and 60% of field capacity) more genistein was required (10  $\mu M).$ 

# Discussion

Both nodulation and N fixation in soybean are sensitive to drying soil and this can have a negative effect on yield (Serraj and Sinclair, 1998). Nodule number, total plant dry weight and shoot-N pool declined as drought stress increased. Serraj *et al.* (1999) established that drought stress leads to decreased N fixation, mainly as a result ureide accumulation in shoots and asparagine in nodules.

In all treatments lowest RWC values were observed at 20 days (Figure 1). This suggests that the period when nodules were formed was the most critical stress phase.

The results indicate that the flavonoid genistein positively influences nodulation, its efficiency and N fixation.

Although no differences were observed in total nodulation between genistein and the uninduced medium (Figure 2B), the abundant nodulation on lateral roots in response to medium Jap without genistein, the N content and nodule weight demonstrated that they were less effective.

The highest soil moisture level guaranteed better nodulation and a higher efficiency of this process was modulated by the inducer. Williams and De Mallorca (1984) demonstrated that the magnitude of stress effects and the rate of inhibition of symbiosis usually depended on the growth and development phase, as well as stress severity. In their results, mild water stress only reduced nodule number on soybean roots. Moderate and severe water stress reduced both nodule number and size.

An effect on N content in induced treatments at 60% field capacity was rare.

The results showed a positive effect of genistein on nodulation, its efficiency and contribution to plant N nutrition at all soil moisture levels and was specially marked under the adverse conditions of drought stress.

Extensive research has focused on decreasing yield losses during soybean crop production. Atti *et al.* (2005) found that foliar application of lipochitinoligosaccharides on soybean gave a positive effects on growth under moderate stress. Their results agree with ours, considering that they used LCO direct. In our work it was used as a *nod* gene inducer, which lead to synthesis of Nod factors in the inocula.

The effect of water lack on nodulation has been extensively documented (Franson et al., 1991; Sellstedt

*et al.*, 1993; Serraj *et al.*, 1999). It is important to produce inoculants which have been obtained from induced media, because they will not only increase nodulation and N fixation, but can also help under adverse conditions of water stress. Other factors may be considered, such as plant growth stage. Peña-Cabriales and Castellanos (1993) found that water stress during vegetative growth was more detrimental to nodulation and N fixation than at the reproduction stage.

In conclusion, after evaluating the effect of genistein as an inducer of *Bradyrhizobium japonicum* inoculants under water stress it was possible to show an important influence of this isoflavonoid on reducing the effect of water stress on nodulation.

# Acknowledgements

This work was supported by Rizobacter Argentina S.A. The authors also thank Fernando Incerti for his help in the experiments.

# References

- ATTI S., BONNELL R., PRASHER S., SMITH D.L., 2005. Response of soybean {*Glycine max* (L.) Merr.} under chronic water deficit to LCO application during flowering and pod filling. Irrig Drain 54, 15-30. doi:10.1002/ird.153.
- BOSNIOLS A., CHAMALET A., LAGACHERIE B., MER-RIEN A., OBATON M., 1986. Le soja. Physiologie de la plante et adaptation aux conditions françaises. Nutrition azotée du soja: limites et améliorations de la fixation symbiotique. CETIOM 1867, 157-165. [In French].
- BROUGHTON W.J., JABBOURI S., PERRET X., 2000. Keys to symbiotic harmony. J Bacteriol 182, 5641-5652. doi:10.1128/JB.182.20.5641-5652.2000.
- COZZI J., BENINTENDE G., 1989. Influencia de la esterilización de la turba en la supervivencia de *Bradyrhizobium japonicum*. Rev Latinoam Microbiol 31, 275-278.
- DUZAN H.M., ZHOU X., SOULEIMANOV A., SMITH D.L., 2004. Perception of *Bradyrhizobium japonicum* Nod factor by soybean [*Glycine max* (L.) Merr.] root hairs under abiotic stress conditions. J Exp Bot 55,2641-2646. doi:10.1093/jxb/erh265.
- FEHR W., CAVINESS C., BURMOOD D., PENNINGTON J., 1971. Stage of development descriptions for soybeans, *Glycine max* (L.) Merrill. Crop Sci 11, 929-931.
- FRANSON R., BROWN M., BETHLENFALVAY G., 1991. The *Glycine-Glomus-Bradyrhizobium* symbiosis. XI. Nod-

ule gas exchange and efficiency as a function of soil and root water status in mycorrhizal soybean. Physiol Plant 83, 476-482. doi:10.1111/j.1399-3054.1991.tb00123.x.

- FREDERICK J., CAMP C., BAUER P., 2001. Drought-stress effects on branch and mainstem seed yield and yield components of determinate soybean. Crop Sci 41, 759-763.
- FULAI L., JENSEN C., ANDERSEN M., 2004. Drought stress effect on carbohydrate concentration in soybean leaves and pods during early reproductive development: its implication in altering podset. Field Crop Res 86, 1-13.
- GAGE D.J., 2004. Infection and invasion of roots by symbiotic, nitrogen-fixing rhizobia during nodulation of temperature legumes. Microbiol Mol Biol Rev 68, 280-300. doi: 10.1128/MMBR.68.2.280-300.2004.
- GEURTS R., FEDOROVA E., BISSELING T., 2005. Nod factor signaling genes and their function in the early stages of *Rhizobium* infection. Curr Opin Plant Biol 346-352.
- GRESSHOFF P., 1993. Molecular genetic analysis of nodulation genes in soybean. Plant Breeding Rev 11, 275-318.
- HEIDSTRA R., YANG W., YALCIN Y., PECK S., EMONS A., VAN KAMMEN A., BISSELING T., 1997. Ethylene provides positional information on cortical cell division but is not involved in Nod factor-induced root hair tip growth in Rhizobium-legume interaction. Development 124, 1781-1787.
- INFOSTAT, 2003. InfoStat version 1.5. Grupo InfoStat, FCA, Universidad Nacional de Córdoba, Argentina, 232 p. [In Spanish].
- JONES K.M., KOBAYASHI H., DAVIES B.W., TAGA M.E., WALKER G.C., 2007. How rhizobial symbionts invade plants: the *Sinorhizobium-Medicago* model. Nat Rev Microbiol 5, 619-633. doi:10.1038/nrmicro1705.
- KING C.A., PURCELL L.C., 2001. Soybean nodule size and relationship to nitrogen fixation response to water deficit. Crop Sci 41, 1099–1107.
- KOSSLAK R.M., BARKEI J., PAAREN H.E., APPELBAUM E.R., 1990. Strain-specific inhibition of nod gene induction in *Bradyrhizobium japonicum* by flavonoid compounds. Appl Environ Microbiol 56, 1333-1341.
- KURDALAI F., AL-AIN F., AL-SHAMMA M., 2002. Nodulation, dry matter production, and N<sub>2</sub> fixation by faba bean and chickpea as affected by soil moisture and potassium fertilizer. J Plant Nutr 25, 355-368. doi:10.1081/PLN-100108841.
- LADRERA R., MARINO D., LARRAINZAR S., GONZÁLEZ E., ARRESE-IGOR C., 2007. Reduced carbon availability to bacteroids and elevated ureides in nodules, but not in shoots, are involved in the nitrogen fixation response to early drought in soybean. Plant Physiol 145, 539-546. doi:10.1104/pp.107.102491.

- LONG S., 2001. Genes and signals in the *Rhizobium*-legume symbiosis. Plant Physiol 125, 69-72. doi:10.1104/pp.125. 1.69.
- MARINO D., PIERRE F., LADRERA R., ZABALZA A., PUPPO A., ARRESE-IGOR C., GONZÁLEZ E., 2007. Nitrogen fixation control under drought stress. Localized or systemic? Plant Physiol 143, 1968-1974. doi:10.1104/ pp.107.097139.
- MONTERO F., SAGARDOY M., 2005. Resultados del uso de un protector bacteriano sobre la nodulación de plantas de soja tratadas con inoculante líquido, insecticidas, fungicida y micronutrientes. V Reunión Nacional Científico Técnica de Biología del Suelo - V Encuentro sobre Fijación Biológica de Nitrógeno. Fac. Ciencias Agrarias, UNJu, San Salvador de Jujuy, Argentina. [In Spanish].
- MORGAN J., BENDING G., WHITE P., 2005. Biological costs and benefits to plant–microbe interactions in the rhizosphere. J Exp Bot 56, 1729-1739. doi:10.1093/jxb/eri205.
- OHASHI Y., SANEOKA H., FUJITA K., 2000. Effect of water stress on growth, photosynthesis, and photoassimilate translocation in soybean and tropical pasture legume siratro. Soil Sci Plant Nutr 46, 417-425.
- PEÑA-CABRIALES J., CASTELLANOS J., 1993. Effect of water stress on N<sub>2</sub> fixation and grain yield of *Phaseolus vulgaris* L. Plant Soil 152, 151-155. doi:10.1007/ BF00016345.
- PERRET X., STAHELIN C., BROUGHTON W.J., 2000. Molecular basis of symbiotic promiscuity. Microbiol Mol Biol Rev 64, 180-201. doi:10.1128/MMBR.64.1.180-201. 2000.
- POMPER W., BREEN P., 1997. Expansion and osmotic adjustment of strawberry fruits during water stress. J Amer Soc Hort Sci 122, 183-189.
- PURCELL L.C., SERRAJ R., SINCLAIR T.R., DE A., 2004. Soybean N<sub>2</sub> fixation estimates, ureide concentration, and yield responses to drought. Crop Sci 44, 484-492.
- SELLSTEDT A., STAAHL L., MATTSSON M., JONSSON K., HOEGBERG P., 1993. Can the <sup>15</sup>N dilution technique be used to study N<sub>2</sub> fixation in tropical tree symbioses as affected by water deficit? J Exp Bot 44, 1749-1755. doi:10.1093/jxb/44.12.1749.
- SERRAJ R., 2003. Effects of drought stress on legume symbiotic nitrogen fixation: Physiological mechanisms. Indian J Exp Biol 41, 1136-1141.
- SERRAJ R., SINCLAIR T., 1998. Soybean cultivar variability for nodule formation and growth under drought. Plant Soil 202, 159-166. doi: 10.1023/A:1004300819535.
- SERRAJ R., SINCLAIR T., PURCELL L., 1999. Symbiotic N<sub>2</sub> fixation response to drought. J Exp Bot 50, 143-155. doi:10.1093/jexbot/50.331.143.

- SPAINK H.P., 2000. Root nodulation and infection factors produced by rhizobial bacteria. Annu Rev Microbiol 54, 257-288. doi:10.1146/annurev.micro.54.1.257.
- SPAINK H.P., KONDOROSI A., HOOYKAAS P.J.J., 1998. The *Rhizobiaceae*. Kluwer Academic Publishers, Dordrecht.
- STOKKERMANS T., PETERS N., 1994. *Bradyrhizobium elkanii* lipo-oligosaccharide signals induce complete nodule structures on *Glycine soja*. Planta 193, 413-420. doi:10.1007/BF00201821.
- STREETER J.G., 2003. Effects of drought on nitrogen fixation in soybean root nodules. Plant Cell Environ 26, 1199-1204. doi:10.1046/j.1365-3040.2003.01041.x.
- TAJIMA S., NOMURA M., KOUCHI H., 2004. Ureide biosynthesis in legume nodules. Front Biosci 9, 1374–1381. doi:10.2741/1345.
- VIJN I., DAS NEVES L., VAN KAMMEN A., FRANSSEN H., BISSELING T., 1993. Nod factors and nodulation in plants. Science 260, 1764-1765. doi:10.1126/science. 8511583.

- VINCENT J.M., 1970. A manual for the practical study of root nodule bacteria. Oxford, Blackwell, Oxford.
- WILLIAMS P., DE MALLORCA M., 1984. Effect of osmotically induced leaf moisture stress on nodulation and nitrogenase activity of *Glycine max*. Plant Soil 80, 267-283. doi:10.1007/BF02161183.
- ZAHRAN H., 1999. Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. Microbiol Mol Biol Rev 63, 968-989.
- ZHANG F., SMITH D.L., 1996. Inoculation of soybean (*Glycine max* (L.) Merr.) with genistein-preincubated *Bradyrhizobium japonicum* or genistein directly into soil increases soybean protein and dry matter yield under short season conditions. Plant Soil 179, 233-241. doi:10.1007/ BF00009333.
- ZHANG F., SMITH D.L., 1997. Application of genistein to inocula and soil to overcome low spring soil temperature inhibition of soybean nodulation and nitrogen fixation. Plant Soil 192, 141-151. doi:10.1023/A:1004232417658.